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NUCLEIC ACID AND CORRESPONDING PROTEIN ENTITLED 101P3A11 USEFUL IN TREATMENT AND DETECTION OF CANCER

RELATED APPLICATIONS

This application claims benefit of priority from U.S. Provisional Application Serial Number 60/291,118, filed May 15, 2001 and is a continuation-in-part of U.S. Application Serial Number 09/680,728, filed October 5, 2000, which claims benefit of priority from U.S. Provisional Application Serial Number 60/157,902, filed October 5, 1999. All three of these applications are hereby incorporated in their entireties as if fully set forth.

FIELD OF THE INVENTION

The invention described herein relates to a novel gene and its encoded protein, termed 101P3A11, and to diagnostic and therapeutic methods and compositions useful in the management of various cancers that express 101P3A11 (also referred to as PHOR-1).

BACKGROUND OF THE INVENTION

Cancer is the second leading cause of human death next to coronary disease. Worldwide, millions of people die from cancer every year. In the United States alone, as reported by the American Cancer Society, cancer causes the death of well over a half-million people annually, with over 1.2 million new cases diagnosed per year. While deaths from heart disease have been declining significantly, those resulting from cancer generally are on the rise. In the early part of the next century, cancer is predicted to become the leading cause of death.

Worldwide, several cancers stand out as the leading killers. In particular, carcinomas of the lung, prostate, breast, colon, pancreas, and ovary represent the primary causes of cancer death. These and virtually all other carcinomas share a common lethal feature. With very few exceptions, metastatic disease from a carcinoma is fatal. Moreover, even for those cancer patients who initially survive their primary cancers, common experience has shown that their lives are dramatically altered. Many cancer patients experience strong anxieties driven by the awareness of the potential for recurrence or treatment failure. Many cancer patients experience physical debilitations following treatment. Furthermore, many cancer patients experience a recurrence.

Worldwide, prostate cancer is the fourth most prevalent cancer in men. In North America and Northern Europe, it is by far the most common cancer in males and is the second leading cause of cancer death in men. In the United States alone, well over 30,000 men die annually of this disease - second only to lung cancer. Despite the magnitude of these figures, there is still no effective treatment for metastatic prostate cancer. Surgical prostatectomy, radiation therapy, hormone ablation therapy, surgical castration and chemotherapy continue to be the main treatment sd-63712

modalities. Unfortunately, these treatments are ineffective for many and are often associated with undesirable consequences.

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On the diagnostic front, the lack of a prostate tumor marker that can accurately detect early-stage, localized tumors remains a significant limitation in the diagnosis and management of this disease. Although the serum prostate specific antigen (PSA) assay has been a very useful tool, however its specificity and general utility is widely regarded as lacking in several important respects.

Progress in identifying additional specific markers for prostate cancer has been improved by the generation of prostate cancer xenografts that can recapitulate different stages of the disease in mice. The LAPC (Los Angeles Prostate Cancer) xenografts are prostate cancer xenografts that have survived passage in severe combined immune deficient (SCID) mice and have exhibited the capacity to mimic the transition from androgen dependence to androgen independence (Klein et al., 1997, Nat. Med. 3:402). More recently identified prostate cancer markers include PCTA-1 (Su et al., 1996, Proc. Natl. Acad. Sci. USA 93: 7252), prostate-specific membrane (PSM) antigen (Pinto et al., Clin Cancer Res 1996 Sep 2 (9): 1445-51), STEAP (Hubert, et al., Proc Natl Acad Sci U S A. 1999 Dec 7; 96(25): 14523-8) and prostate stem cell antigen (PSCA) (Reiter et al., 1998, Proc. Natl. Acad. Sci. USA 95: 1735).

While previously identified markers such as PSA, PSM, PCTA and PSCA have facilitated efforts to diagnose and treat prostate cancer, there is need for the identification of additional markers and therapeutic targets for prostate and related cancers in order to further improve diagnosis and therapy.

Renal cell carcinoma (RCC) accounts for approximately 3 percent of adult malignancies. Once adenomas reach a diameter of 2 to 3 cm, malignant potential exists. In the adult, the two principal malignant renal tumors are renal cell adenocarcinoma and transitional cell carcinoma of the renal pelvis or ureter. The incidence of renal cell adenocarcinoma is estimated at more than 29,000 cases in the United States, and more than 11,600 patients died of this disease in 1998. Transitional cell carcinoma is less frequent, with an incidence of approximately 500 cases per year in the United States.

Surgery has been the primary therapy for renal cell adenocarcinoma for many decades. Until recently, metastatic disease has been refractory to any systemic therapy. With recent developments in systemic therapies, particularly immunotherapies, metastatic renal cell carcinoma may be approached aggressively in appropriate patients with a possibility of durable responses. Nevertheless, there is a remaining need for effective therapies for these patients.

Of all new cases of cancer in the United States, bladder cancer represents approximately 5 percent in men (fifth most common neoplasm) and 3 percent in women (eighth most common neoplasm). The incidence is increasing slowly, concurrent with an increasing older population. In 1998, there was an estimated 54,500 cases, including 39,500 in men and 15,000 in women. The age-adjusted incidence in the United States is 32 per 100,000 for men and 8 per 100,000 in women. The historic male/female ratio of 3:1 may be decreasing related to smoking patterns in women. There were an estimated 11,000 deaths from bladder cancer in 1998 (7,800 in men and 3,900 in women). Bladder cancer incidence and mortality strongly increase with age and will be an increasing problem as the population becomes more elderly.

Most bladder cancers recur in the bladder. Bladder cancer is managed with a combination of transurethral resection of the bladder (TUR) and intravesical chemotherapy or immunotherapy. The multifocal and recurrent nature of bladder cancer points out the limitations of TUR. Most muscle-invasive cancers are not cured by TUR alone. Radical cystectomy and urinary diversion is the most effective means to eliminate the cancer but carry an undeniable

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impact on urinary and sexual function. There continues to be a significant need for treatment modalities that are beneficial for bladder cancer patients.

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An estimated 130,200 cases of colorectal cancer occurred in 2000 in the United States, including 93,800 cases of colon cancer and 36,400 of rectal cancer. Colorectal cancers are the third most common cancers in men and women. Incidence rates declined significantly during 1992-1996 (-2.1% per year). Research suggests that these declines have been due to increased screening and polyp removal, preventing progression of polyps to invasive cancers. There were an estimated 56,300 deaths (47,700 from colon cancer, 8,600 from rectal cancer) in 2000, accounting for about 11% of all U.S. cancer deaths.

At present, surgery is the most common form of therapy for colorectal cancer, and for cancers that have not spread, it is frequently curative. Chemotherapy, or chemotherapy plus radiation, is given before or after surgery to most patients whose cancer has deeply perforated the bowel wall or has spread to the lymph nodes. A permanent colostomy (creation of an abdominal opening for elimination of body wastes) is occasionally needed for colon cancer and is infrequently required for rectal cancer. There continues to be a need for effective diagnostic and treatment modalities for colorectal cancer.

There were an estimated 164,100 new cases of lung and bronchial cancer in 2000, accounting for 14% of all U.S. cancer diagnoses. The incidence rate of lung and bronchial cancer is declining significantly in men, from a high of 86.5 per 100,000 in 1984 to 70.0 in 1996. In the 1990s, the rate of increase among women began to slow. In 1996, the incidence rate in women was 42.3 per 100,000.

Lung and bronchial cancer caused an estimated 156,900 deaths in 2000, accounting for 28% of all cancer deaths. During 1992–1996, mortality from lung cancer declined significantly among men (-1.7% per year) while rates for women were still significantly increasing (0.9% per year). Since 1987, more women have died each year of lung cancer than breast cancer, which, for over 40 years, was the major cause of cancer death in women. Decreasing lung cancer incidence and mortality rates most likely resulted from decreased smoking rates over the previous 30 years; however, decreasing smoking patterns among women lag behind those of men. Of concern, although the declines in adult tobacco use have slowed, tobacco use in youth is increasing again.

Treatment options for lung and bronchial cancer are determined by the type and stage of the cancer and include surgery, radiation therapy, and chemotherapy. For many localized cancers, surgery is usually the treatment of choice. Because the disease has usually spread by the time it is discovered, radiation therapy and chemotherapy are often needed in combination with surgery. Chemotherapy alone or combined with radiation is the treatment of choice for small cell lung cancer; on this regimen, a large percentage of patients experience remission, which in some cases is long lasting. There is however, an ongoing need for effective treatment and diagnostic approaches for lung and bronchial cancers.

An estimated 182,800 new invasive cases of breast cancer were expected to occur among women in the United States during 2000. Additionally, about 1,400 new cases of breast cancer were expected to be diagnosed in men in 2000. After increasing about 4% per year in the 1980s, breast cancer incidence rates in women have leveled off in the 1990s to about 110.6 cases per 100,000.

In the U.S. alone, there were an estimated 41,200 deaths (40,800 women, 400 men) in 2000 due to breast cancer. Breast cancer ranks second among cancer deaths in women. According to the most recent data, mortality rates

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declined significantly during 1992–1996 with the largest decreases in younger women, both white and black. These decreases were probably the result of earlier detection and improved treatment.

Taking into account the medical circumstances and the patient's preferences, treatment of breast cancer may involve lumpectomy (local removal of the tumor) and removal of the lymph nodes under the arm; mastectomy (surgical removal of the breast) and removal of the lymph nodes under the arm; radiation therapy; chemotherapy; or hormone therapy. Often, two or more methods are used in combination. Numerous studies have shown that, for early stage disease, long-term survival rates after lumpectomy plus radiotherapy are similar to survival rates after modified radical mastectomy. Significant advances in reconstruction techniques provide several options for breast reconstruction after mastectomy. Recently, such reconstruction has been done at the same time as the mastectomy.

Local excision of ductal carcinoma in situ (DCIS) with adequate amounts of surrounding normal breast tissue may prevent the local recurrence of the DCIS. Radiation to the breast and/or tamoxifen may reduce the chance of DCIS occurring in the remaining breast tissue. This is important because DCIS, if left untreated, may develop into invasive breast cancer. Nevertheless, there are serious side effects or sequelae to these treatments. There is, therefore, a need for efficacious breast cancer treatments.

There were an estimated 23,100 new cases of ovarian cancer in the United States in 2000. It accounts for 4% of all cancers among women and ranks second among gynecologic cancers. During 1992–1996, ovarian cancer incidence rates were significantly declining. Consequent to ovarian cancer, there were an estimated 14,000 deaths in 2000. Ovarian cancer causes more deaths than any other cancer of the female reproductive system.

Surgery, radiation therapy, and chemotherapy are treatment options for ovarian cancer. Surgery usually includes the removal of one or both ovaries, the fallopian tubes (salpingo-oophorectomy), and the uterus (hysterectomy). In some very early tumors, only the involved ovary will be removed, especially in young women who wish to have children. In advanced disease, an attempt is made to remove all intra-abdominal disease to enhance the effect of chemotherapy. There continues to be an important need for effective treatment options for ovarian cancer.

There were an estimated 28,300 new cases of pancreatic cancer in the United States in 2000. Over the past 20 years, rates of pancreatic cancer have declined in men. Rates among women have remained approximately constant but may be beginning to decline. Pancreatic cancer caused an estimated 28,200 deaths in 2000 in the United States. Over the past 20 years, there has been a slight but significant decrease in mortality rates among men (about -0.9% per year) while rates have increased slightly among women.

Surgery, radiation therapy, and chemotherapy are treatment options for pancreatic cancer. These treatment options can extend survival and/or relieve symptoms in many patients but are not likely to produce a cure for most. There is a significant need for additional therapeutic and diagnostic options for pancreatic cancer.

SUMMARY OF THE INVENTION

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The present invention relates to a gene, designated 101P3A11, that is over-expressed in the normal tissues and cancers listed in Table I. Northern blot expression analysis of 101P3A11 gene expression in normal tissues shows a restricted expression pattern in adult tissues. The nucleotide (Figure 2) and amino acid (Figure 2, and Figure 3) sequences of 101P3A11 are provided. The tissue-related profile of 101P3A11 in normal adult tissues, combined with the over-expression observed in tumors of the tissues listed in Table I, shows that 101P3A11 is aberrantly over-

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expressed in certain some cancers, and thus serves as a useful diagnostic, prophylactic, prognostic, and/or therapeutic target for cancers that express the protein, such as those listed in Table I.

The invention provides polynucleotides corresponding or complementary to all or part of the 101P3A11 genes, mRNAs, and/or coding sequences, preferably in isolated form, including polynucleotides encoding 101P3A11-related proteins and fragments of 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, or more than 25 contiguous amino acids; at least 30, 35, 40, 45, 50, 55, 60, 65, 70, 80, 85, 90, 95, 100 or more than 100 contiguous amino acids of a 101P3A11-related protein, as well as the peptides/proteins themselves; DNA, RNA, DNA/RNA hybrids, and related molecules, polynucleotides or oligonucleotides complementary or having at least a 90% homology to the 101P3A11 genes or mRNA sequences or parts thereof, and polynucleotides or oligonucleotides that hybridize to the 101P3A11 genes, mRNAs, or to 101P3A11-encoding polynucleotides. Also provided are means for isolating cDNAs and the genes encoding 101P3A11. Recombinant DNA molecules containing 101P3A11 polynucleotides, cells transformed or transduced with such molecules, and host-vector systems for the expression of 101P3A11 gene products are also provided. The invention further provides antibodies that bind to 101P3A11 proteins and polypeptide fragments thereof, including polyclonal and monoclonal antibodies, murine and other mammalian antibodies, chimeric antibodies, humanized and fully human antibodies, and antibodies labeled with a detectable marker. In certain embodiments there is a proviso that the entire nucleic acid sequence of Figure 2 is not encoded and/or the entire amino acid sequence of Figure 2 is not prepared. In certain embodiments, the entire nucleic acid sequence of Figure 2 is encoded and/or the entire amino acid sequence of Figure 2 is prepared, either of which are in respective human unit dose forms.

The invention further provides methods for detecting the presence and status of 101P3A11 polynucleotides and proteins in various biological samples, as well as methods for identifying cells that express 101P3A11. A typical embodiment of this invention provides methods for monitoring 101P3A11 gene products in a tissue or hematology sample having or suspected of having some form of growth dysregulation such as cancer.

The invention further provides various immunogenic or therapeutic compositions and strategies for treating cancers that express 101P3A11 such as cancers of tissues listed in Table I, including therapies aimed at inhibiting the transcription, translation, processing or function of 101P3A11 as well as cancer vaccines. In one aspect, the invention provides compositions, and methods comprising them, for treating a cancer that expresses 101P3A11 (PHOR-1) in a human subject wherein the composition comprises a carrier suitable for human use and a human unit dose of one or more than one agent that inhibits the production or function of 101P3A11 (PHOR-1). Preferably, the carrier is a uniquely human carrier. In another aspect of the invention, the agent is a moiety that is immunoreactive with 101P3A11 protein. Non-limiting examples of such moieties include, but are not limited to, antibodies (such as single chain, monoclonal, polyclonal, humanized, or human antibodies), functional equivalents thereof (whether naturally occurring or synthetic), and combinations thereof. In another aspect, the agent is a small molecule as defined herein.

In another aspect, the agent comprises one or more than one peptide which comprises a cytotoxic T lymphocyte (CTL) epitope that binds an HLA class I molecule in a human to elicit a CTL response to 101P3A11 and/or one or more than one peptide which comprises a helper T lymphocyte (HTL) epitope which binds an HLA class II molecule in a human to elicit an HTL response. In a further aspect of the invention, the agent comprises one or more than one nucleic acid molecule that expresses one or more than one of the CTL or HTL response stimulating peptides as described above. In yet another aspect of the invention, the one or more than one nucleic acid molecule may express a

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moiety that is immunologically reactive with 101P3A11 as described above. The one or more than one nucleic acid molecule may also be, or encodes, a molecule that inhibits production of 101P3A11. Non-limiting examples of such molecules include, but are not limited to, those complementary to a nucleotide sequence essential for production of 101P3A11 (e.g. antisense sequences or molecules that form a triple helix with a nucleotide double helix essential for 101P3A11 production) or a ribozyme effective to lyse 101P3A11 mRNA.

BRIEF DESCRIPTION OF THE FIGURES

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Figure 1. 101P3A11 SSH sequence. The 101P3A11 SSH sequence is shown as SEQ ID NO: 2864.

Figures 2A-2D. The cDNA (SEQ ID. NO. :—: 2865) and amino acid sequence (SEQ ID. NO. :—: 2866) of 101P3A11. The start methionine is underlined. The open reading frame extends from nucleic acid 133 to 1086 including the stop codon (the codon for the initial M is omitted as the shorter peptide has a more favorable Kozak sequence).

Figure 3. Amino acid sequence of 101P3A11 (SEQ ID. NO. :____: 2866). The 101P3A11 protein has 317 amino acids.

Figure 4. Alignment of 101P3A11 (Sbjct — SEQ ID NO: 2867) with mouse olfactory receptor S25 (Query—SEQ ID NO: 2868.) The transmembrane regions of 101P3A11 and mouse olfactory receptor S25 (ORS25) predicted using the TMHMM algorithm are highlighted in gray. The amino acids of ORS25 predicted (Floriano, W.B., et al, 2000, Proc. Natl. Acad. Sci., USA, 97:10712-10716) to be involved in binding of the ligand hexanol and/or involved in the formation of the ligand binding pocket are italicized and bolded in the Figure, and are: Leu 131, Val 134, Val 135, Gly 138, Thr139, Ser 193, Ser 197, Phe 225, Ala 230, Ile 231, Gly 234, Thr 284, Phe 287, Gln 300, Lys 302.

Figure 5. Hydrophilicity amino acid profile of 101P3A11 determined by computer algorithm sequence analysis using the method of Hopp and Woods (Hopp T.P., Woods K.R., 1981. Proc. Natl. Acad. Sci. U.S.A. 78:3824-3828) accessed on the Protscale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 6. Hydropathicity amino acid profile of 101P3A11 determined by computer algorithm sequence analysis using the method of Kyte and Doolittle (Kyte J., Doolittle R.F., 1982. J. Mol. Biol. 157:105-132) accessed on the ProtScale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 7. Percent accessible residues amino acid profile of 101P3A11 determined by computer algorithm sequence analysis using the method of Janin (Janin J., 1979 Nature 277:491-492) accessed on the ProtScale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 8. Average flexibility amino acid profile of 101P3A11 determined by computer algorithm sequence analysis using the method of Bhaskaran and Ponnuswamy (Bhaskaran R., and Ponnuswamy P.K., 1988. Int. J. Pept. Protein Res. 32:242-255) accessed on the ProtScale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

Figure 9. Beta-turn amino acid profile of 101P3A11 determined by computer algorithm sequence analysis using the method of Deleage and Roux (Deleage, G., Roux B. 1987 Protein Engineering 1:289-294) accessed on the ProtScale website (www.expasy.ch/cgi-bin/protscale.pl) through the ExPasy molecular biology server.

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Figure 10A. Expression of 101P3A11 by RT-PCR. First strand cDNA was prepared from vital pool 1 (VP1: liver, lung and kidney), vital pool 2 (VP2, pancreas, colon and stomach), prostate xenograft pool, prostate cancer pool, kidney cancer pool, colon cancer pool, breast cancer pool, and cancer metastasis pool. Normalization was performed by PCR using primers to actin and GAPDH. Semi-quantitative PCR, using primers to 101P3A11, was performed at 30 cycles of amplification. Expression of 101P3A11 was observed in prostate xenograft pool, prostate cancer pool, kidney cancer pool, colon cancer pool, breast cancer pool, and cancer metastasis pool, but not in VP1 and VP2.

Figure 10B. Expression of 101P3A11 in human cancers demonstrated by dot blot analysis of tumor RNA (T) and normal RNA (N) matched samples using patient-derived amplified cDNAs. Up-regulation of PHOR-1 expression was found in 3 of 3 prostate cancer patients, 6 of 14 kidney cancer patients, 2 of 8 uterine cancer patients, 3 of 8 stomach cancer patients and 7 of 7 rectal cancer patients.

Figure 11. Expression of 101P3A11 in human patient cancer specimens. RNA was extracted from a pool of three prostate cancer tumors, kidney cancer tumors, colon cancer tumors, breast cancer tumors, and a cancer metastasis pool derived from cancer patients, as well as from normal prostate (NP), normal bladder (NB), normal kidney (NK) and normal colon (NC). Northern blots with 10 μg of total RNA/lane were probed with a 101P3A11 fragment. Size standards in kilobases (kb) are indicated on the side. The results showed expression of 101P3A11 in prostate cancer tumors, kidney cancer tumors, colon cancer tumors, breast cancer tumors, cancer metastasis pool, bladder cancer pool, and in the normal prostate but not in the other normal tissues. A picture of the ethidium-bromide staining of the RNA gel is also presented.

Figure 12A. Expression of 101P3A11 in prostate cancer patient specimens. RNA was extracted from prostate tumors (T) and their normal adjacent tissues (Nat) derived from prostate cancer patients. Northern blots with 10 μg of total RNA/lane were probed with 101P3A11 sequences. Results show upregulated expression of 101P3A11 in 8 of 10 tumor specimens.

Figure 12B. Photomicrograph showing 101P3A11 expression in prostatic intraepithelial neoplasia (PIN) by in situ hybridization with an anti-sense 101P3A11 riboprobe.

Figure 12C. Photomicrograph showing 101P3A11 expression in prostate cancer tissue by in situ hybridization with an anti-sense 101P3A11 riboprobe.

Figure 12D. Photomicrograph showing 101P3A11 expression in prostate cancer by in situ hybridization with an anti-sense 101P3A11 riboprobe. Note up-regulation of expression relative to normal prostate, FIG. 12E.

Figure 12E. Photomicrograph showing 101P3A11 expression in normal prostate by in situ hybridization with an anti-sense 101P3A11 riboprobe.

Figure 13. Expression of 101P3A11 in colon cancer patient specimens. RNA was extracted from colon tumors (T) and their normal adjacent tissues (Nat) derived from colon cancer patients. Northern blots with 10 µg of total RNA/lane were probed with 101P3A11 sequences. Size standards in kilobases (kb) are indicated on the side. Results showed expression of 101P3A11 in colon tumors but not in normal tissues. Expression was also seen in the colon cancer cell line T84. A picture of the ethidium-bromide staining of the RNA gel is also presented.

Figure 14. Expression of 101P3A11 in kidney cancer patient specimens. RNA was extracted from kidney tumors (T) and their normal adjacent tissues (Nat) derived from kidney cancer patients. Northern blots with $10 \mu g$ of total RNA/lane were probed with 101P3A11 sequences. Size standards in kilobases (kb) are indicated on the side. The

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results showed expression of 101P3A11 in five of six kidney tumor specimens. The expression detected in normal adjacent tissues (isolated from diseased tissues) but not in normal tissues (isolated from healthy donors) indicates that these tissues are not fully normal and that 101P3A11 is expressed in early stage tumors. A picture of the ethidium-bromide staining of the RNA gel is also presented.

Figures 15A-15C. Androgen regulation of 101P3A11 in tissue culture cells. LAPC-9 cells were grown in charcoal-stripped medium and stimulated with the synthetic androgen mibolerone, for either 14 or 24 hours. Northern blots with 10 µg of total RNA/lane were probed with 101P3A11 sequences (Figure 15A). A picture of the ethidium-bromide staining of the RNA gel is also presented (Figure 15C). Results showed expression of 101P3A11 was not regulated by androgen. The experimental samples were confirmed by testing for the expression of the androgen-regulated prostate cancer gene PSA (Figure 15B). This experiment showed that, as expected, PSA levels go down in presence of charcoal-stripped serum, and expression is induced at 14 and 24 hours in presence of mibolerone.

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Figure 16. Androgen regulation of 101P3A11 in vivo. Male mice were injected with LAPC-9AD tumor cells. When tumors reached a palpable size (0.3-0.5cm in diameter), mice were castrated and tumors harvested at different time points following the castration. RNA was isolated from the xenograft tissues. Northern blots with 10 μg of total RNA/lane were probed with 101P3A11sequences. Size standards in kilobases (kb) are indicated on the side. A picture of the ethidium-bromide staining of the RNA gel is also presented. The results showed that expression of 101P3A11 is not androgen regulated.

Figure 17. Expression and detection of 101P3A11(159-202)-psecFc fusion protein. The 101P3A11(159-202)-psecFc vector was constructed. The recombinant expression vector DNA was transfected into either 293T cells or Cos-7 cells. Cells as well as culture supernatants (media) were harvested 24 hours later. The cells were lysed, and run on SDS-PAGE gel along with the media samples. The gel was transferred to nitrocellulose, stained with HRP-labeled anti-human IgG and developed using the ECL chemiluminescence detection kit. Results showed expression of the 101P3A11(159-202)-psecFc fusion protein in the lysates of both 293T and Cos-7 cells. The 101P3A11(159-202)-psecFc fusion protein was also secreted and detected in the culture supernatants of both cell types.

Figure 18. Expression of 101P3A11 in 300.19 cells following retroviral-mediated gene delivery. 300.19 cells were transduced with the pSRα retroviral vector encoding the 101P3A11 gene. Following selection with neomycin, the cells were expanded and RNA was extracted. A Northern blot with 10 μg of total RNA/lane was probed with the 101P3A11 sequence. Size standards in kilobases (kb) are indicated on the side. Results showed expression of the 101P3A11 transcript driven from the retroviral LTR. LAPC-4AD and LAPC-9AD showed expression of the endogenous 101P3A11 transcript. The figure shows results of a short exposure of the autoradiogram.

Figure 19A: The secondary structure of 101P3A11 protein was predicted using the HNN - Hierarchical Neural Network method (Guermeur, 1997, http://pbil.ibcp.fr/cgi-bin/npsa_automat.pl?page=npsa_nn.html), accessed from the ExPasy molecular biology server (http://www.expasy.ch/tools/). This method predicts the presence and location of alpha helices, extended strands, and random coils from the primary protein sequnce. The percent of the protein in a given secondary structure is also given. Figure 19B is a schematic representation of the probability of existence of transmembrane regions and orientation of 101P3A11 based on the TMpred algorithm of Hofmann and Stoffel which utilizes TMBASE (K. Hofmann, W. Stoffel. TMBASE - A database of membrane spanning protein segments Biol.

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Chem. Hoppe-Seyler 374:166, 1993). Figure 19C is a schematic representation of the probability of the existence of transmembrane regions and the extracellular and intracellular orientation of 101P3A11 based on the TMHMM algorithm of Sonnhammer, von Heijne, and Krogh (Erik L.L. Sonnhammer, Gunnar von Heijne, and Anders Krogh: A hidden Markov model for predicting transmembrane helices in protein sequences. In Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, p 175-182 Ed J. Glasgow, T. Littlejohn, F. Major, R. Lathrop, D. Sankoff, and C. Sensen Menlo Park, CA: AAAI Press, 1998). The TMpred and TMHMM algorithms are accessed from the ExPasy molecular biology server (http://www.expasy.ch/tools/). The results of the transmembrane prediction programs presented in Figure 19B and 19C depict 101P3A11 as containing 7 transmembrane domains consistent with that of a Gprotein coupled receptor.

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Figure 20. Expression of 101P3A11 in NIH-3T3 Tumors. Mice were injected subcutaneously with control 3T3-neo or NIH3T3 cells expressing 101P3A11. Tumors were allowed to grow, the mice were then sacrificed and tumors harvested. RNA was isolated from LAPC-4AD and LAPC-4AI xenografts, 3T3-neo and 3T3-101P3A11 cells grown in culture were used as controls. RNA isolated from six different tumors derived from 3T3-101P3A11 cells (Tumor #1-3) were compared by Northern blotting. Northern blots with 10 μg of total RNA/lane were probed with 101P3A11 sequence. A picture of the ethidium-bromide staining of the RNA gel is also presented. Results showed expression of 101P3A11 in all 3T3-101P3A11 tumors as well as in 3T3/101P3A11 cells used to derive the tumors, but not in the negative control cells 3T3/neo cells.

Figure 21. 101P3A11 Induces Tumor Formation of 3T3 Cells. Injection of 106 3T3-neo, 3T3-Ras or 3T3-101P3A11 cells (106 of the indicated cells mixed with Matrigel) subcutaneously into 6 male SCID mice (right flank) revealed that 6/6 3T3-v12Ras-injected mice formed tumors, 6/6 3T3-101P3A11- injected mice formed tumors, and 0/6 3T3-neo-injected mice formed tumors. Each data point represents the mean tumor volume (n = 6) in each group.

Figure 22. PTX reduces the in vivo growth of 3T3-101P3A11 Tumors. Pertussis toxin was found to inhibit the sub-cutaneous growth of 3T3-101P3A11 tumors in SCID mice in a dose dependent manner.

Figure 23. Alignment of 101P3A11-PHOR-1 (Phor<u>-SEQ ID NO: 2869</u>) with the rat GPCR RA1C <u>-SEQ ID NO: 2870</u> (gi|3420759). Identities = 179/299 (59%), Positives = 231/299 (76%), Gaps = 1/299 (0%).

Figure 24. Alignment of 101P3A11-PHOR-1 (Phor—SEQ ID NO: 2871) with the human prostate specific GPCR—SEQ ID NO: 2872 (gi|13540539). Identities = 179/299 (59%), Positives = 233/299 (77%), Gaps = 1/299 (0%).

Figure 25. Alignment of 101P3A11-PHOR-1 (Phor<u>SEQ ID NO: 2873</u>) with human olfactory receptor 5II12, HOR5<u>SEQ ID NO: 2874</u>, (gi|14423836). Identities = 163/304 (53%), Positives = 214/304 (69%), Gaps = 1/304 (0%).

Figure 26. 101P3A11 Modulated Tyrosine Phosphorylation in NIH-3T3 Cells. 101P3A11 mediated the dephosphorylation of proteins at 200, 120-140, 85-90 and 55 kDa. 101P3A11 induced the phosphorylation of proteins at 80 and 29 kDa in NIH-3T3 cells.

Figure 27. ERK Phosphorylation by PCR ligands in 101P3A11 Expressing Cells. FBS, lipophosphatidic acid, gastrin releasing peptide, leukotriene and platelet activating factor induced the phosphorylation of ERK in 101P3A11 expressing cells.

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- Figure 28. Inhibition of 101P3A11-Mediated ERK Activation by PD98059. ERK phosphorylation was inhibited by a MEK specific(PD98059) but not a p38 specific (SB203580) inhibitor in PC3-101P3A11 cells.
- **Figure 29.** Enhanced ERK Phosphorylation in Sodium Orthovanadate Treated PC3-101P3A11 Cells. Sodium orthovanadate induced increased ERK phosphorylation in PC3-101P3A11 cells relative to PC3-neo cells.
- Figure 30. Inhibition of 101P3A11-Mediated ERK Phosphorylation by AG1517. The EGFR inhibitor, AG1517, inhibits EGF-mediated ERK phosphorylation in control and 101P3A11-expressing PC3 cells. AG1517 partially inhibits 101P3A11 mediated ERK phosphorylation in PC3 cells.

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- Figures 31A-31B. Activation of p38 in PC3-101P3A11 Cells. Expression of 101P3A11 mediates p38 phosphorylation in cells treated with 10% FBS as shown by blotting with antibodies to phospho-p38 (Figure 31A) compared to p38 (Figure 31B).
- Figure 32. 101P3A11 Induced Accumulation of cAMP in PC3 Cells. Expression of 101P3A11 increased the accumulation of cAMP in cells treated with 0.1% and 10% FBS. FBS-induced cAMP accumulation in 101P3A11 cells was inhibited by pertussis toxin.
- Figure 33. Pertussis Toxin Inhibits 101P3A11 Mediated ERK Phosphorylation. Pertussis toxin inhibited FBS- mediated ERK phosphorylation in 101P3A11 expressing cells.
- Figure 34. Pertussis Toxin Inhibited ERK Phosphorylation in PC3-101P3A11 Cells. Pertussis toxin inhibited FBS- mediated ERK phosphorylation in 101P3A11 expressing cells. The inhibitory activity of pertussis toxin on ERK phosphorylation was more dramatic in FBS-treated than EGF or GRP-treated PC3-101P3A11 cells
- Figure 35. Inhibition of 101P3A11-mediated signaling by Suranim, a G protein inhibitor. Control NIH 3T3 and 3T3-101P3A11 cells were grown in the presence of absence of G protein inhibitors Surinam and NF449. Proliferation was analyzed by Alamar blue after 72 hours. Suranim and NF449 inhibited the proliferation of 101P3A11 expressing but not control cells.
- **Figures 36A-36B.** 101P3A11 Mediated ERK Phosphorylation By Conditioned Media. Figure 36A: blotting with anti-phospho ERK antibodies; Figure 36B: blotting with anti-ERK antibodies. Supernatants from PC3, PC3-101P3A11, PrEC and LAPC42 cells induce ERK phosphorylation in PC3 101P3A11 but not PC3 cells. Supernatants from 3T3 and 293T cells had little specific effect on ERK phosphorylation.
- Figure 37. 101P3A11 Enhances the Proliferation of 3T3 Cells. Control NIH 3T3 and 3T3-101P3A11 cells were grown in the presence of absence 0.5 or 10% FBS. Proliferation was analyzed by Alamar blue after 48 hours. Expression of 101P3A11 induced a 6 fold increase in the proliferation of 3T3 cells grown in 0.5% FBS.
- Figure 38. Inhibition of 101P3A11 Mediated ERK Phosphorylation by 101P3A11 Specific Antibodies. Expression of 101P3A11 induced ERK phosphorylation in 293T cells. Anti-101P3A11 pAb inhibited ERK Phosphorylation in 293T-101P3A11 cells.
- Figure 39. Anti-101P3A11 Ab Mediated cAMP Accumulation in PC3-101P3A11 Cells. Control PC3 cells and cells expressing 101P3A11 were treated with anti-101P3A11 pAb for 2 min and evaluated for intracellular cAMP content. The assay was performed in duplicate.
- Figures 40A-40F. Photomicrographs showing immunohistochemical analysis using anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded prostate cancer tissues (Figure 40A); anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded prostate

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cancer cell line, LNCaP (Figure 40B); anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded prostate cancer tissues (Figure 40C); anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded normal prostate (Figure 40D); anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded prostate cancer tissues (Figure 40E); and anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded normal prostate (Figure 40F).

Figures 41A-41F. Photomicrographs showing immunohistochemical analysis using anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded prostate cancer tissues (Figure 41A); anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded bladder cancer tissues (Figure 41B); anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded kidney cancer tissues (Figure 41C); anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded colon cancer tissues (Figure 41D); anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded lung cancer tissues (Figure 41E); and anti-101P3A11 (peptide 1; amino acids 1-14) rabbit polyclonal antibody on formalin fixed and paraffin embedded breast cancer tissues (Figure 41F).

Figure 42 shows demonstrates that 101P3A11 induces orthotopic growth of tumors. $5x10^5$ cells were injected orthotopically into SCID mice, 7 mice per group; tumor weight was evaluated 24-25 days post cell injection.

Figure 43 shows that 101P3A11 induces colony formation in a soft agar assay.

DETAILED DESCRIPTION OF THE INVENTION

Outline of Sections

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- I.) Definitions
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 - II.A.2.) Antisense Embodiments
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 - IV.) 101P3A11 Antibodies
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 - X.C.1. Minigene Vaccines
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 - X.C.4. Vaccine Compositions Comprising DC Pulsed with CTL and/or HTL Peptides
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- XI.) Diagnostic and Prognostic Embodiments of 101P3A11.
- XII.) Inhibition of 101P3A11 Protein Function
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 - XII.C.) Inhibition of 101P3A11 Transcription or Translation
 - XII.D.) General Considerations for Therapeutic Strategies
- XIII.) KITS

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I.) Definitions:

Unless otherwise defined, all terms of art, notations and other scientific terms or terminology used herein are intended to have the meanings commonly understood by those of skill in the art to which this invention pertains. In some cases, terms with commonly understood meanings are defined herein for clarity and/or for ready reference, and the inclusion of such definitions herein should not necessarily be construed to represent a substantial difference over what is generally understood in the art. Many of the techniques and procedures described or referenced herein are well understood and commonly employed using conventional methodology by those skilled in the art, such as, for example, the widely utilized molecular cloning methodologies described in Sambrook et al., Molecular Cloning: A Laboratory Manual 2nd. edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. As appropriate, procedures involving the use of commercially available kits and reagents are generally carried out in accordance with manufacturer defined protocols and/or parameters unless otherwise noted.

The terms "advanced prostate cancer", "locally advanced prostate cancer", "advanced disease" and "locally advanced disease" mean prostate cancers that have extended through the prostate capsule, and are meant to include stage C disease under the American Urological Association (AUA) system, stage C1 - C2 disease under the Whitmore-Jewett system, and stage T3 - T4 and N+ disease under the TNM (tumor, node, metastasis) system. In general, surgery is not recommended for patients with locally advanced disease, and these patients have substantially less favorable outcomes compared to patients having clinically localized (organ-confined) prostate cancer. Locally advanced disease is clinically identified by palpable evidence of induration beyond the lateral border of the prostate, or asymmetry or

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induration above the prostate base. Locally advanced prostate cancer is presently diagnosed pathologically following radical prostatectomy if the tumor invades or penetrates the prostatic capsule, extends into the surgical margin, or invades the seminal vesicles.

"Altering the native glycosylation pattern" is intended for purposes herein to mean deleting one or more carbohydrate moieties found in native sequence 101P3A11 (either by removing the underlying glycosylation site or by deleting the glycosylation by chemical and/or enzymatic means), and/or adding one or more glycosylation sites that are not present in the native sequence 101P3A11. In addition, the phrase includes qualitative changes in the glycosylation of the native proteins, involving a change in the nature and proportions of the various carbohydrate moieties present.

The term "analog" refers to a molecule which is structurally similar or shares similar or corresponding attributes with another molecule (e.g. a 101P3A11-related protein). For example an analog of the 101P3A11 protein can be specifically bound by an antibody or T cell that specifically binds to 101P3A11.

The term "antibody" is used in the broadest sense. Therefore an "antibody" can be naturally occurring or manmade such as monoclonal antibodies produced by conventional hybridoma technology. Anti-101P3A11 antibodies comprise monoclonal and polyclonal antibodies as well as fragments containing the antigen-binding domain and/or one or more complementarity determining regions of these antibodies.

An "antibody fragment" is defined as at least a portion of the variable region of the immunoglobulin molecule that binds to its target, i.e., the antigen-binding region. In one embodiment it specifically covers single anti-101P3A11 antibodies and clones thereof (including agonist, antagonist and neutralizing antibodies) and anti-101P3A11 antibody compositions with polyepitopic specificity.

The term "codon optimized sequences" refers to nucleotide sequences that have been optimized for a particular host species by replacing any codons having a usage frequency of less than about 20%. Nucleotide sequences that have been optimized for expression in a given host species by elimination of spurious polyadenylation sequences, elimination of exon/intron splicing signals, elimination of transposon-like repeats and/or optimization of GC content in addition to codon optimization are referred to herein as an "expression enhanced sequences."

The term "cytotoxic agent" refers to a substance that inhibits or prevents the function of cells and/or causes destruction of cells. The term is intended to include radioactive isotopes chemotherapeutic agents, and toxins such as small molecule toxins or enzymatically active toxins of bacterial, fungal, plant or animal origin, including fragments and/or variants thereof. Examples of cytotoxic agents include, but are not limited to maytansinoids, yttrium, bismuth, ricin, ricin A-chain, doxorubicin, daunorubicin, taxol, ethidium bromide, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicine, dihydroxy anthracin dione, actinomycin, diphtheria toxin, Pseudomonas exotoxin (PE) A, PE40, abrin, abrin A chain, modeccin A chain, alpha-sarcin, gelonin, mitogellin, retstrictocin, phenomycin, enomycin, curicin, crotin, calicheamicin, sapaonaria officinalis inhibitor, and glucocorticoid and other chemotherapeutic agents, as well as radioisotopes such as At²¹¹, I¹³¹, I¹²⁵, Y⁹⁰, Re¹⁸⁶, Re¹⁸⁸, Sm¹⁵³, Bi²¹², P³² and radioactive isotopes of Lu. Antibodies may also be conjugated to an anti-cancer pro-drug activating enzyme capable of converting the pro-drug to its active form.

The term "homolog" refers to a molecule which exhibits homology to another molecule, by for example, having sequences of chemical residues that are the same or similar at corresponding positions.

"Human Leukocyte Antigen" or "HLA" is a human class I or class II Major Histocompatibility Complex (MHC) protein (see, e.g., Stites, et al., IMMUNOLOGY, 8TH ED., Lange Publishing, Los Altos, CA (1994).

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The terms "hybridize", "hybridizing", "hybridizes" and the like, used in the context of polynucleotides, are meant to refer to conventional hybridization conditions, preferably such as hybridization in 50% formamide/6XSSC/0.1% SDS/100 µg/ml ssDNA, in which temperatures for hybridization are above 37 degrees C and temperatures for washing in 0.1XSSC/0.1% SDS are above 55 degrees C.

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The phrases "isolated" or "biologically pure" refer to material which is substantially or essentially free from components which normally accompany the material as it is found in its native state. Thus, isolated peptides in accordance with the invention preferably do not contain materials normally associated with the peptides in their *in situ* environment. For example, a polynucleotide is said to be "isolated" when it is substantially separated from contaminant polynucleotides that correspond or are complementary to genes other than the 101P3A11 gene or that encode polypeptides other than 101P3A11 gene product or fragments thereof. A skilled artisan can readily employ nucleic acid isolation procedures to obtain an isolated 101P3A11 polynucleotide. A protein is said to be "isolated," for example, when physical, mechanical or chemical methods are employed to remove the 101P3A11 protein from cellular constituents that are normally associated with the protein. A skilled artisan can readily employ standard purification methods to obtain an isolated 101P3A11 protein. Alternatively, an isolated protein can be prepared by chemical means.

The term "mammal" refers to any organism classified as a mammal, including mice, rats, rabbits, dogs, cats, cows, horses and humans. In one embodiment of the invention, the mammal is a mouse. In another embodiment of the invention, the mammal is a human.

The terms "metastatic prostate cancer" and "metastatic disease" mean prostate cancers that have spread to regional lymph nodes or to distant sites, and are meant to include stage D disease under the AUA system and stage TxNxM+ under the TNM system. As is the case with locally advanced prostate cancer, surgery is generally not indicated for patients with metastatic disease, and hormonal (androgen ablation) therapy is a preferred treatment modality. Patients with metastatic prostate cancer eventually develop an androgen-refractory state within 12 to 18 months of treatment initiation. Approximately half of these androgen-refractory patients die within 6 months after developing that status. The most common site for prostate cancer metastasis is bone. Prostate cancer bone metastases are often osteoblastic rather than osteolytic (i.e., resulting in net bone formation). Bone metastases are found most frequently in the spine, followed by the femur, pelvis, rib cage, skull and humerus. Other common sites for metastasis include lymph nodes, lung, liver and brain. Metastatic prostate cancer is typically diagnosed by open or laparoscopic pelvic lymphadenectomy, whole body radionuclide scans, skeletal radiography, and/or bone lesion biopsy.

The term "monoclonal antibody" refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the antibodies comprising the population are identical except for possible naturally occurring mutations that are present in minor amounts.

A "motif", as in biological motif of an 101P3A11-related protein, refers to any pattern of amino acids forming part of the primary sequence of a protein, that is associated with a particular function (e.g. protein-protein interaction, protein-DNA interaction, etc) or modification (e.g. that is phosphorylated, glycosylated or amidated), or localization (e.g. secretory sequence, nuclear localization sequence, etc.) or a sequence that is correlated with being immunogenic, either humorally or cellularly. A motif can be either contiguous or capable of being aligned to certain positions that are generally correlated with a certain function or property. In the context of HLA motifs, "motif" refers to the pattern of residues in a peptide of defined length, usually a peptide of from about 8 to about 13 amino acids for a class I HLA

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motif and from about 6 to about 25 amino acids for a class II HLA motif, which is recognized by a particular HLA molecule. Peptide motifs for HLA binding are typically different for each protein encoded by each human HLA allele and differ in the pattern of the primary and secondary anchor residues.

A "pharmaceutical excipient" comprises a material such as an adjuvant, a carrier, pH-adjusting and buffering agents, tonicity adjusting agents, wetting agents, preservative, and the like.

"Pharmaceutically acceptable" refers to a non-toxic, inert, and/or composition that is physiologically compatible with humans or other mammals.

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The term "polynucleotide" means a polymeric form of nucleotides of at least 10 bases or base pairs in length, either ribonucleotides or deoxynucleotides or a modified form of either type of nucleotide, and is meant to include single and double stranded forms of DNA and/or RNA. In the art, this term if often used interchangeably with "oligonucleotide". A polynucleotide can comprise a nucleotide sequence disclosed herein wherein thymidine (T) (as shown for example in SEQ ID NO: 702) can also be uracil (U); this definition pertains to the differences between the chemical structures of DNA and RNA, in particular the observation that one of the four major bases in RNA is uracil (U) instead of thymidine (T).

The term "polypeptide" means a polymer of at least about 4, 5, 6, 7, or 8 amino acids. Throughout the specification, standard three letter or single letter designations for amino acids are used. In the art, this term is often used interchangeably with "peptide" or "protein".

An HLA "primary anchor residue" is an amino acid at a specific position along a peptide sequence which is understood to provide a contact point between the immunogenic peptide and the HLA molecule. One to three, usually two, primary anchor residues within a peptide of defined length generally defines a "motif" for an immunogenic peptide. These residues are understood to fit in close contact with peptide binding groove of an HLA molecule, with their side chains buried in specific pockets of the binding groove. In one embodiment, for example, the primary anchor residues for an HLA class I molecule are located at position 2 (from the amino terminal position) and at the carboxyl terminal position of a 8, 9, 10, 11, or 12 residue peptide epitope in accordance with the invention. In another embodiment, for example, the primary anchor residues of a peptide that will bind an HLA class II molecule are spaced relative to each other, rather than to the termini of a peptide, where the peptide is generally of at least 9 amino acids in length. The primary anchor positions for each motif and supermotif are set forth in Table IV. For example, analog peptides can be created by altering the presence or absence of particular residues in the primary and/or secondary anchor positions shown in Table IV. Such analogs are used to modulate the binding affinity and/or population coverage of a peptide comprising a particular HLA motif or supermotif.

A "recombinant" DNA or RNA molecule is a DNA or RNA molecule that has been subjected to molecular manipulation *in vitro*.

"Stringency" of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe length, washing temperature, and salt concentration. In general, longer probes require higher temperatures for proper annealing, while shorter probes need lower temperatures. Hybridization generally depends on the ability of denatured nucleic acid sequences to reanneal when complementary strands are present in an environment below their melting temperature. The higher the degree of desired homology between the probe and hybridizable sequence, the higher the relative temperature that can be used. As a result, it

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follows that higher relative temperatures would tend to make the reaction conditions more stringent, while lower temperatures less so. For additional details and explanation of stringency of hybridization reactions, see Ausubel et al., Current Protocols in Molecular Biology, Wiley Interscience Publishers, (1995).

"Stringent conditions" or "high stringency conditions", as defined herein, are identified by, but not limited to, those that: (1) employ low ionic strength and high temperature for washing, for example 0.015 M sodium chloride/0.0015 M sodium citrate/0.1% sodium dodecyl sulfate at 50°C; (2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1% bovine serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/50 mM sodium phosphate buffer at pH 6.5 with 750 mM sodium chloride, 75 mM sodium citrate at 42 °C; or (3) employ 50% formamide, 5 x SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt's solution, sonicated salmon sperm DNA (50 μg/ml), 0.1% SDS, and 10% dextran sulfate at 42 °C, with washes at 42°C in 0.2 x SSC (sodium chloride/sodium. citrate) and 50% formamide at 55 °C, followed by a high-stringency wash consisting of 0.1 x SSC containing EDTA at 55 °C. "Moderately stringent conditions" are described by, but not limited to, those in Sambrook et al., Molecular Cloning: A Laboratory Manual, New York: Cold Spring Harbor Press, 1989, and include the use of washing solution and hybridization conditions (e.g., temperature, ionic strength and %SDS) less stringent than those described above. An example of moderately stringent conditions is overnight incubation at 37°C in a solution comprising; 20% formamide, 5 x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5 x Denhardt's solution, 10% dextran sulfate, and 20 mg/mL denatured sheared salmon sperm DNA, followed by washing the filters in 1 x SSC at about 37-50°C. The skilled artisan will recognize how to adjust the temperature, ionic strength, etc. as necessary to accommodate factors such as probe length and the like.

An HLA "supermotif" is a peptide binding specificity shared by HLA molecules encoded by two or more HLA alleles.

As used herein "to treat" or "therapeutic" and grammatically related terms, refer to any improvement of any consequence of disease, such as prolonged survival, less morbidity, and/or a lessening of side effects which are the byproducts of an alternative therapeutic modality; full eradication of disease is not required.

A "transgenic animal" (e.g., a mouse or rat) is an animal having cells that contain a transgene, which transgene was introduced into the animal or an ancestor of the animal at a prenatal, e.g., an embryonic stage. A "transgene" is a DNA that is integrated into the genome of a cell from which a transgenic animal develops.

As used herein, an HLA or cellular immune response "vaccine" is a composition that contains or encodes one or more peptides of the invention. There are numerous embodiments of such vaccines, such as a cocktail of one or more individual peptides; one or more peptides of the invention comprised by a polyepitopic peptide; or nucleic acids that encode such individual peptides or polypeptides, *e.g.*, a minigene that encodes a polyepitopic peptide. The "one or more peptides" can include any whole unit integer from 1-150 or more, e.g., at least 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 or more peptides of the invention. The peptides or polypeptides can optionally be modified, such as by lipidation, addition of targeting or other sequences. HLA class I peptides of the invention can be admixed with, or linked to, HLA class II

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peptides, to facilitate activation of both cytotoxic T lymphocytes and helper T lymphocytes. HLA vaccines can also comprise peptide-pulsed antigen presenting cells, *e.g.*, dendritic cells.

The term "variant" refers to a molecule that exhibits a variation from a described type or norm, such as a protein that has one or more different amino acid residues in the corresponding position(s) of a specifically described protein (e.g. the 101P3A11 protein shown in Figure 2 or Figure 3). An analog is an example of a variant protein.

The 101P3A11-related proteins of the invention include those specifically identified herein, as well as allelic variants, conservative substitution variants, analogs and homologs that can be isolated/generated and characterized without undue experimentation following the methods outlined herein or readily available in the art. Fusion proteins that combine parts of different 101P3A11 proteins or fragments thereof, as well as fusion proteins of a 101P3A11 protein and a heterologous polypeptide are also included. Such 101P3A11 proteins are collectively referred to as the 101P3A11-related proteins, the proteins of the invention, or 101P3A11. The term "101P3A11-related protein" refers to a polypeptide fragment or an 101P3A11 protein sequence of 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, or more than 25 amino acids; or, at least 30, 35, 40, 45, 50, 55, 60, 65, 70, 80, 85, 90, 95, 100 or more than 100 amino acids.

II.) 101P3A11 Polynucleotides

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One aspect of the invention provides polynucleotides corresponding or complementary to all or part of an 101P3A11 gene, mRNA, and/or coding sequence, preferably in isolated form, including polynucleotides encoding an 101P3A11-related protein and fragments thereof, DNA, RNA, DNA/RNA hybrid, and related molecules, polynucleotides or oligonucleotides complementary to an 101P3A11 gene or mRNA sequence or a part thereof, and polynucleotides or oligonucleotides that hybridize to an 101P3A11 gene, mRNA, or to an 101P3A11 encoding polynucleotide (collectively, "101P3A11 polynucleotides"). In all instances when referred to in this section, T can also be U in Figure 2.

Embodiments of a 101P3A11 polynucleotide include: a 101P3A11 polynucleotide having the sequence shown in Figure 2, the nucleotide sequence of 101P3A11 as shown in Figure 2, wherein T is U; at least 10 contiguous nucleotides of a polynucleotide having the sequence as shown in Figure 2; or, at least 10 contiguous nucleotides of a polynucleotide having the sequence as shown in Figure 2 where T is U. For example, embodiments of 101P3A11 nucleotides comprise, without limitation:

- (a) a polynucleotide comprising, consisting essentially of, or consisting of the sequence as shown in Figure 2 (SEQ ID NO.: 2865), wherein T can also be U;
- (b) a polynucleotide comprising, consisting essentially of, or consisting of the sequence as shown in Figure 2 (SEQ ID NO.: 2865), from nucleotide residue number 133 through nucleotide residue number 1086, wherein T can also be U;
- (c) a polynucleotide that encodes a 101P3A11-related protein whose sequence is encoded by the cDNAs contained in the plasmid designated p101P3A11, clone GTH10 deposited with American Type Culture Collection as Accession No. PTA-312 on July 2, 1999;
- (d) a polynucleotide that encodes an 101P3A11-related protein that is at least 90% homologous to the entire amino acid sequence shown in Figure 2 (SEO ID NO.: 2866);

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- (e) a polynucleotide that encodes an 101P3A11-related protein that is at least 90% identical to the entire amino acid sequence shown in Figure 2 (SEQ ID NO: 2866);
- (f) a polynucleotide that encodes at least one peptide set forth in Tables V-XVIII or XXVI-XXVIII;
- (g) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 317 that includes an amino acid position having a value greater than 0.5 in the Hydrophilicity profile of Figure 5;
 - (h) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 317 that includes an amino acid position having a value less than 0.5 in the Hydropathicity profile of Figure 6;
- (i) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 317 that includes an amino acid position having a value greater than 0.5 in the Percent Accessible Residues profile of Figure 7;
 - (j) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 317 that includes an amino acid position having a value greater than 0.5 in the Average Flexibility profile on Figure 8;
 - (k) a polynucleotide that encodes a peptide region of at least 5 amino acids of Figure 3 in any whole number increment up to 317 that includes an amino acid position having a value greater than 0.5 in the Betaturn profile of Figure 9;
 - (l) a polynucleotide that is fully complementary to a polynucleotide of any one of (a)-(k);
 - (m) a polynucleotide that selectively hybridizes under stringent conditions to a polynucleotide of (a)-(l);
 - (n) a peptide that is encoded by any of (a)-(k); and,
- 25 (o) a polynucleotide of any of (a)-(m)or peptide of (n) together with a pharmaceutical excipient and/or in a human unit dose form.

As used herein, a range is understood to specifically disclose all whole unit positions thereof.

Typical embodiments of the invention disclosed herein include 101P3A11 polynucleotides that encode specific portions of the 101P3A11 mRNA sequence (and those which are complementary to such sequences) such as those that encode the protein and fragments thereof, for example of 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 205, 210, 215, 220, 225, 250, 275, 300, or 317 contiguous amino acids.

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For example, representative embodiments of the invention disclosed herein include: polynucleotides and their encoded peptides themselves encoding about amino acid 1 to about amino acid 10 of the 101P3A11 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 10 to about amino acid 20 of the 101P3A11 protein shown in Figure 2, or Figure 3, polynucleotides encoding about amino acid 20 to about amino acid 30 of the 101P3A11 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 30 to about amino acid 40 of the 101P3A11 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 40 to about amino acid 50 of the 101P3A11 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 50 to about amino acid 60 of the 101P3A11 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 60 to about amino acid 70 of the 101P3A11 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 70 to about amino acid 80 of the 101P3A11 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 80 to about amino acid 90 of the 101P3A11 protein shown in Figure 2 or Figure 3, polynucleotides encoding about amino acid 90 to about amino acid 100 of the 101P3A11 protein shown in Figure 2 or Figure 3, in increments of about 10 amino acids, ending at the carboxyl terminal amino acid set forth in Figure 2 or Figure 3. Accordingly polynucleotides encoding portions of the amino acid sequence (of about 10 amino acids), of amino acids 100 through the carboxyl terminal amino acid of the 101P3A11 protein are embodiments of the invention. Wherein it is understood that each particular amino acid position discloses that position plus or minus five amino acid residues.

Polynucleotides encoding relatively long portions of the 101P3A11 protein are also within the scope of the invention. For example, polynucleotides encoding from about amino acid 1 (or 20 or 30 or 40 etc.) to about amino acid 20, (or 30, or 40 or 50 etc.) of the 101P3A1100 protein shown in Figure 2 or Figure 3 can be generated by a variety of techniques well known in the art. These polynucleotide fragments can include any portion of the 101P3A11 sequence as shown in Figure 2 or Figure 3.

Additional illustrative embodiments of the invention disclosed herein include 101P3A11 polynucleotide fragments encoding one or more of the biological motifs contained within the 101P3A11 protein sequence, including one or more of the subsequences of the 101P3A11 protein set forth in Tables V-XVIII or XXVI-XXVIII. In another embodiment, typical polynucleotide fragments of the invention encode one or more of the regions of 101P3A11 that exhibit homology to a known molecule. In another embodiment of the invention, typical polynucleotide fragments can encode one or more of the 101P3A11 N-glycosylation sites, cAMP and cGMP-dependent protein kinase phosphorylation sites, casein kinase II phosphorylation sites or N-myristoylation site and amidation sites.

II.A.) Uses of 101P3A11 Polynucleotides

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II.A.1.) Monitoring of Genetic Abnormalities

The polynucleotides of the preceding paragraphs have a number of different specific uses. The human 101P3A11 gene maps to the chromosomal location set forth in Table XXV. For example, because the 101P3A11 gene maps to this chromosome, polynucleotides that encode different regions of the 101P3A11 protein are used to characterize cytogenetic abnormalities of this chromosomal locale, such as abnormalities that are identified as being associated with various cancers. In certain genes, a variety of chromosomal abnormalities including rearrangements have been identified as frequent cytogenetic abnormalities in a number of different cancers (see e.g. Krajinovic et al., Mutat. Res. 382(3-4): 81-83 (1998); Johansson et al., Blood 86(10): 3905-3914 (1995) and Finger et al., P.N.A.S.

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85(23): 9158-9162 (1988)). Thus, polynucleotides encoding specific regions of the 101P3A11 protein provide new tools that can be used to delineate, with greater precision than previously possible, cytogenetic abnormalities in the chromosomal region that encodes 101P3A11 that may contribute to the malignant phenotype. In this context, these polynucleotides satisfy a need in the art for expanding the sensitivity of chromosomal screening in order to identify more subtle and less common chromosomal abnormalities (see e.g. Evans et al., Am. J. Obstet. Gynecol 171(4): 1055-1057 (1994)).

Furthermore, as 101P3A11 was shown to be highly expressed in prostate and other cancers, 101P3A11 polynucleotides are used in methods assessing the status of 101P3A11 gene products in normal versus cancerous tissues. Typically, polynucleotides that encode specific regions of the 101P3A11 protein are used to assess the presence of perturbations (such as deletions, insertions, point mutations, or alterations resulting in a loss of an antigen etc.) in specific regions of the 101P3A11 gene, such as such regions containing one or more motifs. Exemplary assays include both RT-PCR assays as well as single-strand conformation polymorphism (SSCP) analysis (see, e.g., Marrogi et al., J. Cutan. Pathol. 26(8): 369-378 (1999), both of which utilize polynucleotides encoding specific regions of a protein to examine these regions within the protein.

II.A.2.) Antisense Embodiments

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Other specifically contemplated nucleic acid related embodiments of the invention disclosed herein are genomic DNA, cDNAs, ribozymes, and antisense molecules, as well as nucleic acid molecules based on an alternative backbone, or including alternative bases, whether derived from natural sources or synthesized, and include molecules capable of inhibiting the RNA or protein expression of 101P3A11. For example, antisense molecules can be RNAs or other molecules, including peptide nucleic acids (PNAs) or non-nucleic acid molecules such as phosphorothioate derivatives, that specifically bind DNA or RNA in a base pair-dependent manner. A skilled artisan can readily obtain these classes of nucleic acid molecules using the 101P3A11 polynucleotides and polynucleotide sequences disclosed herein.

Antisense technology entails the administration of exogenous oligonucleotides that bind to a target polynucleotide located within the cells. The term "antisense" refers to the fact that such oligonucleotides are complementary to their intracellular targets, e.g., 101P3A11. See for example, Jack Cohen, Oligodeoxynucleotides, Antisense Inhibitors of Gene Expression, CRC Press, 1989; and Synthesis 1:1-5 (1988). The 101P3A11 antisense oligonucleotides of the present invention include derivatives such as S-oligonucleotides (phosphorothioate derivatives or S-oligos, see, Jack Cohen, supra), which exhibit enhanced cancer cell growth inhibitory action. S-oligos (nucleoside phosphorothioates) are isoelectronic analogs of an oligonucleotide (O-oligo) in which a nonbridging oxygen atom of the phosphate group is replaced by a sulfur atom. The S-oligos of the present invention can be prepared by treatment of the corresponding O-oligos with 3H-1,2-benzodithiol-3-one-1,1-dioxide, which is a sulfur transfer reagent. See Iyer, R. P. et al., J. Org. Chem. 55:4693-4698 (1990); and Iyer, R. P. et al., J. Am. Chem. Soc. 112:1253-1254 (1990). Additional 101P3A11 antisense oligonucleotides of the present invention include morpholino antisense oligonucleotides known in the art (see, e.g., Partridge et al., 1996, Antisense & Nucleic Acid Drug Development 6: 169-175).

The 101P3A11 antisense oligonucleotides of the present invention typically can be RNA or DNA that is complementary to and stably hybridizes with the first 100 5' codons or last 100 3' codons of the 101P3A11 genomic sequence or the corresponding mRNA. Absolute complementarity is not required, although high degrees of complementarity are preferred. Use of an oligonucleotide complementary to this region allows for the selective

hybridization to 101P3A11 mRNA and not to mRNA specifying other regulatory subunits of protein kinase. In one embodiment, 101P3A11 antisense oligonucleotides of the present invention are 15 to 30-mer fragments of the antisense DNA molecule that have a sequence that hybridizes to 101P3A11 mRNA. Optionally, 101P3A11 antisense oligonucleotide is a 30-mer oligonucleotide that is complementary to a region in the first 10 5' codons or last 10 3' codons of 101P3A11. Alternatively, the antisense molecules are modified to employ ribozymes in the inhibition of 101P3A11 expression, see, e.g., L. A. Couture & D. T. Stinchcomb; *Trends Genet* 12: 510-515 (1996).

II.A.3.) Primers and Primer Pairs

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Further specific embodiments of this nucleotides of the invention include primers and primer pairs, which allow the specific amplification of polynucleotides of the invention or of any specific parts thereof, and probes that selectively or specifically hybridize to nucleic acid molecules of the invention or to any part thereof. Probes can be labeled with a detectable marker, such as, for example, a radioisotope, fluorescent compound, bioluminescent compound, a chemiluminescent compound, metal chelator or enzyme. Such probes and primers are used to detect the presence of a 101P3A11 polynucleotide in a sample and as a means for detecting a cell expressing a 101P3A11 protein.

Examples of such probes include polypeptides comprising all or part of the human 101P3A11 cDNA sequence shown in Figure 2. Examples of primer pairs capable of specifically amplifying 101P3A11 mRNAs are also described in the Examples. As will be understood by the skilled artisan, a great many different primers and probes can be prepared based on the sequences provided herein and used effectively to amplify and/or detect a 101P3A11 mRNA.

The 101P3A11 polynucleotides of the invention are useful for a variety of purposes, including but not limited to their use as probes and primers for the amplification and/or detection of the 101P3A11 gene(s), mRNA(s), or fragments thereof; as reagents for the diagnosis and/or prognosis of prostate cancer and other cancers; as coding sequences capable of directing the expression of 101P3A11 polypeptides; as tools for modulating or inhibiting the expression of the 101P3A11 gene(s) and/or translation of the 101P3A11 transcript(s); and as therapeutic agents.

The present invention includes the use of any probe as described herein to identify and isolate a 101P3A11 or 101P3A11 related nucleic acid sequence from a naturally occurring source, such as humans or other mammals, as well as the isolated nucleic acid sequence *per se*, which would comprise all or most of the sequences found in the probe used.

II.A.4.) Isolation of 101P3A11-Encoding Nucleic Acid Molecules

The 101P3A11 cDNA sequences described herein enable the isolation of other polynucleotides encoding 101P3A11 gene product(s), as well as the isolation of polynucleotides encoding 101P3A11 gene product homologs, alternatively spliced isoforms, allelic variants, and mutant forms of the 101P3A11 gene product as well as polynucleotides that encode analogs of 101P3A11-related proteins. Various molecular cloning methods that can be employed to isolate full length cDNAs encoding an 101P3A11 gene are well known (see, for example, Sambrook, J. et al., Molecular Cloning: A Laboratory Manual, 2d edition, Cold Spring Harbor Press, New York, 1989; Current Protocols in Molecular Biology. Ausubel et al., Eds., Wiley and Sons, 1995). For example, lambda phage cloning methodologies can be conveniently employed, using commercially available cloning systems (e.g., Lambda ZAP Express, Stratagene). Phage clones containing 101P3A11 gene cDNAs can be identified by probing with a labeled 101P3A11 cDNA or a fragment thereof. For example, in one embodiment, the 101P3A11 cDNA (Figure 2) or a portion thereof can be synthesized and used as a probe to retrieve overlapping and full-length cDNAs corresponding to a 101P3A11 gene. The 101P3A11 gene itself can be isolated by

screening genomic DNA libraries, bacterial artificial chromosome libraries (BACs), yeast artificial chromosome libraries (YACs), and the like, with 101P3A11 DNA probes or primers.

II.A.5.) Recombinant Nucleic Acid Molecules and Host-Vector Systems

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The invention also provides recombinant DNA or RNA molecules containing an 101P3A11 polynucleotide, a fragment, analog or homologue thereof, including but not limited to phages, plasmids, phagemids, cosmids, YACs, BACs, as well as various viral and non-viral vectors well known in the art, and cells transformed or transfected with such recombinant DNA or RNA molecules. Methods for generating such molecules are well known (see, for example, Sambrook et al, 1989, supra).

The invention further provides a host-vector system comprising a recombinant DNA molecule containing a 101P3A11 polynucleotide, fragment, analog or homologue thereof within a suitable prokaryotic or eukaryotic host cell. Examples of suitable eukaryotic host cells include a yeast cell, a plant cell, or an animal cell, such as a mammalian cell or an insect cell (e.g., a baculovirus-infectible cell such as an Sf9 or HighFive cell). Examples of suitable mammalian cells include various prostate cancer cell lines such as DU145 and TsuPr1, other transfectable or transducible prostate cancer cell lines, primary cells (PrEC), as well as a number of mammalian cells routinely used for the expression of recombinant proteins (e.g., COS, CHO, 293, 293T cells). More particularly, a polynucleotide comprising the coding sequence of 101P3A11 or a fragment, analog or homolog thereof can be used to generate 101P3A11 proteins or fragments thereof using any number of host-vector systems routinely used and widely known in the art.

A wide range of host-vector systems suitable for the expression of 101P3A11 proteins or fragments thereof are available, see for example, Sambrook et al., 1989, supra; Current Protocols in Molecular Biology, 1995, supra). Preferred vectors for mammalian expression include but are not limited to pcDNA 3.1 myc-His-tag (Invitrogen) and the retroviral vector pSRαtkneo (Muller et al., 1991, MCB 11:1785). Using these expression vectors, 101P3A11 can be expressed in several prostate cancer and non-prostate cell lines, including for example 293, 293T, rat-1, NIH 3T3 and TsuPr1. The host-vector systems of the invention are useful for the production of a 101P3A11 protein or fragment thereof. Such host-vector systems can be employed to study the functional properties of 101P3A11 and 101P3A11 mutations or analogs.

Recombinant human 101P3A11 protein or an analog or homolog or fragment thereof can be produced by mammalian cells transfected with a construct encoding a 101P3A11-related nucleotide. For example, 293T cells can be transfected with an expression plasmid encoding 101P3A11 or fragment, analog or homolog thereof, the 101P3A11 or related protein is expressed in the 293T cells, and the recombinant 101P3A11 protein is isolated using standard purification methods (e.g., affinity purification using anti-101P3A11 antibodies). In another embodiment, a 101P3A11 coding sequence is subcloned into the retroviral vector pSRαMSVtkneo and used to infect various mammalian cell lines, such as NIH 3T3, TsuPr1, 293 and rat-1 in order to establish 101P3A11 expressing cell lines. Various other expression systems well known in the art can also be employed. Expression constructs encoding a leader peptide joined in frame to the 101P3A11 coding sequence can be used for the generation of a secreted form of recombinant 101P3A11 protein.

As discussed herein, redundancy in the genetic code permits variation in 101P3A11 gene sequences. In particular, it is known in the art that specific host species often have specific codon preferences, and thus one can adapt the disclosed sequence as preferred for a desired host. For example, preferred analog codon sequences typically have

rare codons (i.e., codons having a usage frequency of less than about 20% in known sequences of the desired host) replaced with higher frequency codons. Codon preferences for a specific species are calculated, for example, by utilizing codon usage tables available on the INTERNET such as at URL www.dna.affrc.go.jp/~nakamura/codon.html.

Additional sequence modifications are known to enhance protein expression in a cellular host. These include elimination of sequences encoding spurious polyadenylation signals, exon/intron splice site signals, transposon-like repeats, and/or other such well-characterized sequences that are deleterious to gene expression. The GC content of the sequence is adjusted to levels average for a given cellular host, as calculated by reference to known genes expressed in the host cell. Where possible, the sequence is modified to avoid predicted hairpin secondary mRNA structures. Other useful modifications include the addition of a translational initiation consensus sequence at the start of the open reading frame, as described in Kozak, *Mol. Cell Biol.*, 9:5073-5080 (1989). Skilled artisans understand that the general rule that eukaryotic ribosomes initiate translation exclusively at the 5' proximal AUG codon is abrogated only under rare conditions (see, e.g., Kozak PNAS 92(7): 2662-2666, (1995) and Kozak NAR 15(20): 8125-8148 (1987)).

III.) 101P3A11-related Proteins

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Another aspect of the present invention provides 101P3A11-related proteins. Specific embodiments of 101P3A11 proteins comprise a polypeptide having all or part of the amino acid sequence of human 101P3A11 as shown in Figure 2 or Figure 3. Alternatively, embodiments of 101P3A11 proteins comprise variant, homolog or analog polypeptides that have alterations in the amino acid sequence of 101P3A11 shown in Figure 2 or Figure 3.

In general, naturally occurring allelic variants of human 101P3A11 share a high degree of structural identity and homology (e.g., 90% or more homology). Typically, allelic variants of the 101P3A11 protein contain conservative amino acid substitutions within the 101P3A11 sequences described herein or contain a substitution of an amino acid from a corresponding position in a homologue of 101P3A11. One class of 101P3A11 allelic variants are proteins that share a high degree of homology with at least a small region of a particular 101P3A11 amino acid sequence, but further contain a radical departure from the sequence, such as a non-conservative substitution, truncation, insertion or frame shift. In comparisons of protein sequences, the terms, similarity, identity, and homology each have a distinct meaning as appreciated in the field of genetics. Moreover, orthology and paralogy can be important concepts describing the relationship of members of a given protein family in one organism to the members of the same family in other organisms.

Amino acid abbreviations are provided in Table II. Conservative amino acid substitutions can frequently be made in a protein without altering either the conformation or the function of the protein. Proteins of the invention can comprise 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15 conservative substitutions. Such changes include substituting any of isoleucine (I), valine (V), and leucine (L) for any other of these hydrophobic amino acids; aspartic acid (D) for glutamic acid (E) and vice versa; glutamine (Q) for asparagine (N) and vice versa; and serine (S) for threonine (T) and vice versa. Other substitutions can also be considered conservative, depending on the environment of the particular amino acid and its role in the three-dimensional structure of the protein. For example, glycine (G) and alanine (A) can frequently be interchangeable, as can alanine (A) and valine (V). Methionine (M), which is relatively hydrophobic, can frequently be interchanged with leucine and isoleucine, and sometimes with valine. Lysine (K) and arginine (R) are frequently interchangeable in locations in which the significant feature of the amino acid residue is its charge and the differing pK's of these two amino acid residues are not significant. Still other changes can be considered "conservative"

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in particular environments (see, e.g. Table III herein; pages 13-15 "Biochemistry" 2nd ED. Lubert Stryer ed (Stanford University); Henikoff et al., PNAS 1992 Vol 89 10915-10919; Lei et al., J Biol Chem 1995 May 19; 270(20):11882-6).

Embodiments of the invention disclosed herein include a wide variety of art-accepted variants or analogs of 101P3A11 proteins such as polypeptides having amino acid insertions, deletions and substitutions. 101P3A11 variants can be made using methods known in the art such as site-directed mutagenesis, alanine scanning, and PCR mutagenesis. Site-directed mutagenesis (Carter et al., *Nucl. Acids Res., 13*:4331 (1986); Zoller et al., *Nucl. Acids Res., 10*:6487 (1987)), cassette mutagenesis (Wells et al., Gene, 34:315 (1985)), restriction selection mutagenesis (Wells et al., *Philos. Trans. R. Soc. London SerA*, 317:415 (1986)) or other known techniques can be performed on the cloned DNA to produce the 101P3A11 variant DNA.

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Scanning amino acid analysis can also be employed to identify one or more amino acids along a contiguous sequence that is involved in a specific biological activity such as a protein-protein interaction. Among the preferred scanning amino acids are relatively small, neutral amino acids. Such amino acids include alanine, glycine, serine, and cysteine. Alanine is typically a preferred scanning amino acid among this group because it eliminates the side-chain beyond the beta-carbon and is less likely to alter the main-chain conformation of the variant. Alanine is also typically preferred because it is the most common amino acid. Further, it is frequently found in both buried and exposed positions (Creighton, *The Proteins*, (W.H. Freeman & Co., N.Y.); Chothia, J. Mol. Biol., 150:1 (1976)). If alanine substitution does not yield adequate amounts of variant, an isosteric amino acid can be used.

As defined herein, 101P3A11 variants, analogs or homologs, have the distinguishing attribute of having at least one epitope that is "cross reactive" with a 101P3A11 protein having the amino acid sequence of SEQ ID NO: 703. As used in this sentence, "cross reactive" means that an antibody or T cell that specifically binds to an 101P3A11 variant also specifically binds to the 101P3A11 protein having the amino acid sequence of SEQ ID NO: 703. A polypeptide ceases to be a variant of the protein shown in SEQ ID NO: 703 when it no longer contains any epitope capable of being recognized by an antibody or T cell that specifically binds to the 101P3A11 protein. Those skilled in the art understand that antibodies that recognize proteins bind to epitopes of varying size, and a grouping of the order of about four or five amino acids, contiguous or not, is regarded as a typical number of amino acids in a minimal epitope. See, e.g., Nair et al., J. Immunol 2000 165(12): 6949-6955; Hebbes et al., Mol Immunol (1989) 26(9):865-73; Schwartz et al., J. Immunol (1985) 135(4):2598-608.

Another class of 101P3A11-related protein variants share 70%, 75%, 80%, 85% or 90% or more similarity with the amino acid sequence of SEQ ID NO: 703 or a fragment thereof. Another specific class of 101P3A11 protein variants or analogs comprise one or more of the 101P3A11 biological motifs described herein or presently known in the art. Thus, encompassed by the present invention are analogs of 101P3A11 fragments (nucleic or amino acid) that have altered functional (e.g. immunogenic) properties relative to the starting fragment. It is to be appreciated that motifs now or which become part of the art are to be applied to the nucleic or amino acid sequences of Figure 2 or Figure 3.

As discussed herein, embodiments of the claimed invention include polypeptides containing less than the full amino acid sequence of the 101P3A11 protein shown in Figure 2 or Figure 3. For example, representative embodiments of the invention comprise peptides/proteins having any 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15 or more contiguous amino acids of the 101P3A11 protein shown in Figure 2 or Figure 3. With respect to any aspect or embodiment of the

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invention, a peptide or peptides may be on one or more than one molecule and may include "pseudopeptides" made of amino acid -like residues that are linked by non-peptide bonds such as, but not limited to, -CH₂-NH-.

Moreover, representative embodiments of the invention disclosed herein include polypeptides consisting of about amino acid 1 to about amino acid 10 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 10 to about amino acid 20 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 20 to about amino acid 30 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 30 to about amino acid 40 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 40 to about amino acid 50 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 50 to about amino acid 60 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 60 to about amino acid 70 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 70 to about amino acid 80 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 80 to about amino acid 90 of the 101P3A11 protein shown in Figure 2 or Figure 3, polypeptides consisting of about amino acid 90 to about amino acid 100 of the 101P3A11 protein shown in Figure 2 or Figure 3, etc. throughout the entirety of the 101P3A11 amino acid sequence. Moreover, polypeptides consisting of about amino acid 1 (or 20 or 30 or 40 etc.) to about amino acid 20, (or 130, or 140 or 150 etc.) of the 101P3A11 protein shown in Figure 2 or Figure 3 are embodiments of the invention. It is to be appreciated that the starting and stopping positions in this paragraph refer to the specified position as well as that position plus or minus 5 residues.

101P3A11-related proteins are generated using standard peptide synthesis technology or using chemical cleavage methods well known in the art. Alternatively, recombinant methods can be used to generate nucleic acid molecules that encode a 101P3A11-related protein. In one embodiment, nucleic acid molecules provide a means to generate defined fragments of the 101P3A11 protein (or variants, homologs or analogs thereof).

III.A.) Motif-bearing Protein Embodiments

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Additional illustrative embodiments of the invention disclosed herein include 101P3A11 polypeptides comprising the amino acid residues of one or more of the biological motifs contained within the 101P3A11 polypeptide sequence set forth in Figure 2 or Figure 3. Various motifs are known in the art, and a protein can be evaluated for the presence of such motifs by a number of publicly available Internet sites (see, e.g., URL addresses: pfam.wustl.edu/; searchlauncher.bcm.tmc.edu/seq-search/struc-predict.html; psort.ims.u-tokyo.ac.jp/; www.cbs.dtu.dk; www.ebi.ac.uk/interpro/scan.html; www.expasy.ch/tools/scnpsit1.html; EpimatrixTM and EpimerTM, Brown University, www.brown.edu/Research/TB-HIV Lab/epimatrix/epimatrix.html; and BIMAS, bimas.dcrt.nih.gov/.).

Motif bearing subsequences of the 101P3A11 protein are set forth and identified in Table XIX.

Table XX sets forth several frequently occurring motifs based on pfam searches (see URL address pfam.wustl.edu/). The columns of Table XX list (1) motif name abbreviation, (2) percent identity found amongst the different member of the motif family, (3) motif name or description and (4) most common function; location information is included if the motif is relevant for location.

Polypeptides comprising one or more of the 101P3A11 motifs discussed above are useful in elucidating the specific characteristics of a malignant phenotype in view of the observation that the 101P3A11 motifs discussed above are associated with growth dysregulation and because 101P3A11 is expressed or overexpressed in certain cancers (See,

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e.g., Table I). Casein kinase II, cAMP and camp-dependent protein kinase, and Protein Kinase C, for example, are enzymes known to be associated with the development of the malignant phenotype (see e.g. Chen et al., Lab Invest., 78(2): 165-174 (1998); Gaiddon et al., Endocrinology 136(10): 4331-4338 (1995); Hall et al., Nucleic Acids Research 24(6): 1119-1126 (1996); Peterziel et al., Oncogene 18(46): 6322-6329 (1999) and O'Brian, Oncol. Rep. 5(2): 305-309 (1998)). Moreover, both glycosylation and myristoylation are protein modifications also associated with cancer and cancer progression (see e.g. Dennis et al., Biochem. Biophys. Acta 1473(1):21-34 (1999); Raju et al., Exp. Cell Res. 235(1): 145-154 (1997)). Amidation is another protein modification also associated with cancer and cancer progression (see e.g. Treston et al., J. Natl. Cancer Inst. Monogr. (13): 169-175 (1992)).

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In another embodiment, proteins of the invention comprise one or more of the immunoreactive epitopes identified in accordance with art-accepted methods, such as the peptides set forth in Tables V-XVIII or XXVI-XXVIII. CTL epitopes can be determined using specific algorithms to identify peptides within an 101P3A11 protein that are capable of optimally binding to specified HLA alleles (e.g., Table IV; Epimatrix™ and Epimer™, Brown University, URL brown.edu/Research/TB-HIV_Lab/epimatrix/epimatrix.html; and BIMAS, URL bimas.dcrt.nih.gov/.) Moreover, processes for identifying peptides that have sufficient binding affinity for HLA molecules and which are correlated with being immunogenic epitopes, are well known in the art, and are carried out without undue experimentation. In addition, processes for identifying peptides that are immunogenic epitopes, are well known in the art, and are carried out without undue experimentation either *in vitro* or *in vivo*.

Also known in the art are principles for creating analogs of such epitopes in order to modulate immunogenicity. For example, one begins with an epitope that bears a CTL or HTL motif (see, e.g., the HLA Class I and HLA Class II motifs/supermotifs of Table IV). The epitope is analoged by substituting out an amino acid at one of the specified positions, and replacing it with another amino acid specified for that position. For example, one can substitute out a deleterious residue in favor of any other residue, such as a preferred residue as defined in Table IV; substitute a less-preferred residue with a preferred residue as defined in Table IV; or substitute an originally-occurring preferred residue with another preferred residue as defined in Table IV. Substitutions can occur at primary anchor positions or at other positions in a peptide; see, e.g., Table IV.

A variety of references reflect the art regarding the identification and generation of epitopes in a protein of interest as well as analogs thereof. See, for example, WO 9733602 to Chesnut et al.; Sette, Immunogenetics 1999 50(3-4): 201-212; Sette et al., J. Immunol. 2001 166(2): 1389-1397; Sidney et al., Hum. Immunol. 1997 58(1): 12-20; Kondo et al., Immunogenetics 1997 45(4): 249-258; Sidney et al., J. Immunol. 1996 157(8): 3480-90; and Falk et al., Nature 351: 290-6 (1991); Hunt et al., Science 255:1261-3 (1992); Parker et al., J. Immunol. 149:3580-7 (1992); Parker et al., J. Immunol. 152:163-75 (1994)); Kast et al., 1994 152(8): 3904-12; Borras-Cuesta et al., Hum. Immunol. 2000 61(3): 266-278; Alexander et al., J. Immunol. 2000 164(3); 164(3): 1625-1633; Alexander et al., PMID: 7895164, UI: 95202582; O'Sullivan et al., J. Immunol. 1991 147(8): 2663-2669; Alexander et al., Immunol. Res. 1998 18(2): 79-92.

Related embodiments of the inventions include polypeptides comprising combinations of the different motifs set forth in Table XIX, and/or, one or more of the predicted CTL epitopes of Table V through Table XVIII, and/or, one or more of the T cell binding motifs known in the art. Preferred embodiments contain no insertions, deletions or substitutions either within the motifs or the intervening sequences of the polypeptides. In addition, embodiments which

include a number of either N-terminal and/or C-terminal amino acid residues on either side of these motifs may be desirable (to, for example, include a greater portion of the polypeptide architecture in which the motif is located). Typically the number of N-terminal and/or C-terminal amino acid residues on either side of a motif is between about 1 to about 100 amino acid residues, preferably 5 to about 50 amino acid residues.

101P3A11-related proteins are embodied in many forms, preferably in isolated form. A purified 101P3A11 protein molecule will be substantially free of other proteins or molecules that impair the binding of 101P3A11 to antibody, T cell or other ligand. The nature and degree of isolation and purification will depend on the intended use. Embodiments of a 101P3A11-related proteins include purified 101P3A11-related proteins and functional, soluble 101P3A11-related proteins. In one embodiment, a functional, soluble 101P3A11 protein or fragment thereof retains the ability to be bound by antibody, T cell or other ligand.

The invention also provides 101P3A11 proteins comprising biologically active fragments of the 101P3A11 amino acid sequence shown in Figure 2 or Figure 3. Such proteins exhibit properties of the 101P3A11 protein, such as the ability to elicit the generation of antibodies that specifically bind an epitope associated with the 101P3A11 protein; to be bound by such antibodies; to elicit the activation of HTL or CTL; and/or, to be recognized by HTL or CTL.

101P3A11-related polypeptides that contain particularly interesting structures can be predicted and/or identified using various analytical techniques well known in the art, including, for example, the methods of Chou-Fasman, Garnier-Robson, Eisenberg, Karplus-Schultz or Jameson-Wolf analysis, Hopp T.P. and Woods K.R., Kyte J. and Doolittle R.F., Janin J., Bhaskaran R., and Ponnuswamy P.K., Deleage, G. and Roux B., or on the basis of immunogenicity. Fragments that contain such structures are particularly useful in generating subunit-specific anti-101P3A11 antibodies, or T cells or in identifying cellular factors that bind to 101P3A11.

CTL epitopes can be determined using specific algorithms to identify peptides within an 101P3A11 protein that are capable of optimally binding to specified HLA alleles (e.g., by using the SYFPEITHI site at World Wide Web URL syfpeithi.bmi-heidelberg.com/; the listings in Table IV(A)-(E); Epimatrix™ and Epimer™, Brown University, URL (www.brown.edu/Research/TB-HIV Lab/epimatrix/epimatrix.html); and BIMAS, URL bimas.dcrt.nih.gov/). Illustrating this, peptide epitopes from 101P3A11 that are presented in the context of human MHC class I molecules HLA-A1, A2, A3, A11, A24, B7 and B35 were predicted (Tables V-XVIII). Specifically, the complete amino acid sequence of the 101P3A11 protein was entered into the HLA Peptide Motif Search algorithm found in the Bioinformatics and Molecular Analysis Section (BIMAS) web site listed above. The HLA peptide motif search algorithm was developed by Dr. Ken Parker based on binding of specific peptide sequences in the groove of HLA Class I molecules, in particular HLA-A2 (see, e.g., Falk et al., Nature 351: 290-6 (1991); Hunt et al., Science 255:1261-3 (1992); Parker et al., J. Immunol. 149:3580-7 (1992); Parker et al., J. Immunol. 152:163-75 (1994)). This algorithm allows location and ranking of 8mer, 9-mer, and 10-mer peptides from a complete protein sequence for predicted binding to HLA-A2 as well as numerous other HLA Class I molecules. Many HLA class I binding peptides are 8-, 9-, 10 or 11-mers. For example, for class I HLA-A2, the epitopes preferably contain a leucine (L) or methionine (M) at position 2 and a valine (V) or leucine (L) at the C-terminus (see, e.g., Parker et al., J. Immunol. 149:3580-7 (1992)). Selected results of 101P3A11 predicted binding peptides are shown in Tables V-XVIII or XXVI-XXVIII herein. In Tables V-XVIII, the top 50 ranking candidates, 9-mers and 10-mers, for each family member are shown along with their location, the amino acid sequence of each specific peptide, and an estimated binding score. The binding score corresponds to the estimated half

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time of dissociation of complexes containing the peptide at 37°C at pH 6.5. Peptides with the highest binding score are predicted to be the most tightly bound to HLA Class I on the cell surface for the greatest period of time and thus represent the best immunogenic targets for T-cell recognition.

In the BIMAS site, the algorithm used to score each 8-mer, 9-mer, or 10-mer peptide subsequence is as follows: The initial (running) score is set to 1.0. For each residue position, the program examines which amino acid appears at that position. The running score is then multiplied by the coefficient for that amino acid type, at that position, for the chosen HLA molecule. These coefficients were pre-calculated and were stored for use by the scoring algorithm in a separate directory as a collection of HLA coefficient files. Using 9-mers, nine multiplications are performed. Using 10-mers, nine multiplications are again performed, because the residue lying at the fifth position in the subsequence is skipped. The resulting running score is multiplied by a final constant to yield an estimate of the half time of disassociation. (This constant is stored at the end of the coefficient file for the HLA molecule. It can have a different value for each HLA molecule.) The final multiplication yields the score reported in the output table. Using 8mers, eight multiplications are performed instead of nine, with a different coefficient matrix employed (20-by-8 rather than 20-by-9). For each HLA molecule evaluated by the BIMAS site, the coefficient values discussed above are stored in a file in a separate directory of coefficient files. When the user selects the HLA molecule type (or, on the restricted Web page for advanced users, selects the actual filename from the coefficient filename menu) and selects "9-mer" or "10-mer" as the length of the subsequence, the "standard" file containing the 20-by-9 coefficient matrix for the HLA molecule is read on-the-fly, with the 181 values (180 coefficient values plus one final constant) being read into an internal array for use by the scoring program. If the user selected "8-mer" as the subsequence length, then the program proceeds in the same fashion, but reads in a different coefficient file appropriate for 8-mer searches on that selected HLA molecule type.

In addition epitopes were identified from the SYFPEITHI site (SEQ ID NO: 2875) listed above. The scoring system for the SYFPEITHI site evaluates every amino acid within a given peptide. Individual amino acids may be given the arbitrary value 1 for amino acids that are only slightly preferred in the respective position, optimal anchor residues are given the value 15; any value between these two is possible. Negative values are also possible for amino acids which are disadvantageous for the peptide's binding capacity at a certain sequence position. The allocation of values is based on the frequency of the respective amino acid in natural ligands, T-cell epitopes, or binding peptides. Further explanations on the algorithm can be found in Rammensee, HG, et al. (1999) SYFPEITHI: database for MHC ligands and peptide motifs. Immunogenetics 50: 213-219. In order to have an extremely reliable SYFPEITHI epitope prediction algorithm, only those MHC class I alleles for which a large amount of data is available are included in the "epitope prediction" section of SYFPEITHI. A reliability of at least 80% in retrieving the most apt epitope can be expected. Thus the naturally presented epitope should be among the top-scoring 2 % of all peptides predicted in 80 % of all predictions. For example, if a protein sequence is as long as 400 amino acids, it can be split into 392 possible nonameric peptides. Therefore, the correct epitope should then be among the 8 top-scoring peptides. For epitope predictions using MHC class II motifs high reliabilities usually cannot be achieved because of the more variable pocket binding behavior. A reliability of approximately 50% only is expected for HLA class II epitopes.

Also, different MHC class I molecules prefer a different length of ligands. For example, SYFPEITHI offers predictions for H2-Kb octamers, HLA-A*0201 nonamers and decamers, or HLA-B8 octamers and nonamers. The

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maximal scores vary between different MHC alleles. Therefore, one can include known ligands/epitopes in order to have an approximation of the scoring. For example, the maximal score for HLA-A*0201 peptides is 36. The well-known epitope GILGFVFTL derived from the influenza A matrix protein scores 30. All predicted MHC class II ligands are 15mers, consisting of three N-terminal flanking residues, the nonamer core sequence located within the binding groove, and three C-terminal flanking residues. Thus, anchor residue P1 appears in position 4 of the peptides predicted with "SYFPEITHI".

Accordingly, identification of HLA Class I and Class II immunogenic peptides for 101P3A11 was carried out using the SYFPEITHI database for MHC ligands and the accompanying prediction algorithm (Hans-Georg Rammensee, Jutta Bachmann, Niels Nikolaus Emmerich, Oskar Alexander Bachor, Stefan Stevanovic: SYFPEITHI: database for MHC ligands and peptide motifs. Immunogenetics 1999, 50: 213-219) accessed through the website with a URL: syfpeithi.bmi-heidelberg.com/scripts/MHCServer.dll/home.htm. (Tables XXVI-XXVIII) For HLA Class I alleles nonameric (Table XXVI) and decameric peptides (Table XXVII) were identified. Given the nature of the HLA Class II binding pocket, for HLA Class II alleles sample 15-mer peptides were identified (Table XXVIII). It is appreciated by those in the art that additional amino acids can be present on the termini of the 15-mer epitopes, or that less than 15 amino acids can be present, and that the epitope will still be bound in the open-ended HLA Class II binding pocket.

For each given HLA allele, listed in Tables XXVI-XXVIII are the peptides whose score ranks within the top 50% of the highest scoring peptide(s) predicted for that allele. For example, if the highest scoring value for nonamers of the HLA-A1 allele is 30, all peptides that are given a score of 15 or higher are listed. Listed in the tables is the amino acid position within the 101P3A11 protein sequence for the first amino acid of each listed peptide (Pos), the peptide sequence, and the relative ranked score (Score).

Actual binding of peptides to an HLA allele can be evaluated by stabilization of HLA expression on the antigen-processing defective cell line T2 (see, e.g., Xue et al., Prostate 30:73-8 (1997) and Peshwa et al., Prostate 36:129-38 (1998)). Immunogenicity of specific peptides can be evaluated *in vitro* by stimulation of CD8+ cytotoxic T lymphocytes (CTL) in the presence of antigen presenting cells such as dendritic cells.

It is to be appreciated that every epitope predicted by the BIMAS site, Epimer™ and Epimatrix™ sites, or specified by the HLA class I or class II motifs available in the art or which become part of the art such as set forth in Table IV (or determined using World Wide Web site URL syfpeithi.bmi-heidelberg.com/) are to be "applied" to the 101P3A11 protein. As used in this context "applied" means that the 101P3A11 protein is evaluated, e.g., visually or by computer-based patterns finding methods, as appreciated by those of skill in the relevant art. Every subsequence of the 101P3A11 of 8, 9, 10, or 11 amino acid residues that bears an HLA Class I motif, or a subsequence of 9 or more amino acid residues that bear an HLA Class II motif are within the scope of the invention.

III.B.) Expression of 101P3A11-related Proteins

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In an embodiment described in the examples that follow, 101P3A11 can be conveniently expressed in cells (such as 293T cells) transfected with a commercially available expression vector such as a CMV-driven expression vector encoding 101P3A11 with a C-terminal 6XHis and MYC tag (pcDNA3.1/mycHIS, Invitrogen or Tag5, GenHunter Corporation, Nashville TN). The Tag5 vector provides an IgGK secretion signal that can be used to facilitate the production of a secreted 101P3A11 protein in transfected cells. The secreted HIS-tagged 101P3A11 in the culture media can be purified, e.g., using a nickel column using standard techniques.

III.C.) Modifications of 101P3A11-related Proteins

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Modifications of 101P3A11-related proteins such as covalent modifications are included within the scope of this invention. One type of covalent modification includes reacting targeted amino acid residues of a 101P3A11 polypeptide with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C-terminal residues of the 101P3A11. Another type of covalent modification of the 101P3A11 polypeptide included within the scope of this invention comprises altering the native glycosylation pattern of a protein of the invention. Another type of covalent modification of 101P3A11 comprises linking the 101P3A11 polypeptide to one of a variety of nonproteinaceous polymers, e.g., polyethylene glycol (PEG), polypropylene glycol, or polyoxyalkylenes, in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337.

The 101P3A11-related proteins of the present invention can also be modified to form a chimeric molecule comprising 101P3A11 fused to another, heterologous polypeptide or amino acid sequence. Such a chimeric molecule can be synthesized chemically or recombinantly. A chimeric molecule can have a protein of the invention fused to another tumor-associated antigen or fragment thereof. Alternatively, a protein in accordance with the invention can comprise a fusion of fragments of the 101P3A11 sequence (amino or nucleic acid) such that a molecule is created that is not, through its length, directly homologous to the amino or nucleic acid sequences shown in Figure 2 or Figure 3. Such a chimeric molecule can comprise multiples of the same subsequence of 101P3A11. A chimeric molecule can comprise a fusion of a 101P3A11-related protein with a polyhistidine epitope tag, which provides an epitope to which immobilized nickel can selectively bind, with cytokines or with growth factors. The epitope tag is generally placed at the amino- or carboxyl- terminus of the 101P3A11. In an alternative embodiment, the chimeric molecule can comprise a fusion of a 101P3A11-related protein with an immunoglobulin or a particular region of an immunoglobulin. For a bivalent form of the chimeric molecule (also referred to as an "immunoadhesin"), such a fusion could be to the Fc region of an IgG molecule. The Ig fusions preferably include the substitution of a soluble (transmembrane domain deleted or inactivated) form of a 101P3A11 polypeptide in place of at least one variable region within an Ig molecule. In a preferred embodiment, the immunoglobulin fusion includes the hinge, CH2 and CH3, or the hinge, CHI, CH2 and CH3 regions of an IgGI molecule. For the production of immunoglobulin fusions see, e.g., U.S. Patent No. 5,428,130 issued June 27, 1995.

III.D.) Uses of 101P3A11-related Proteins

The proteins of the invention have a number of different specific uses. As 101P3A11 is highly expressed in prostate and other cancers, 101P3A11-related proteins are used in methods that assess the status of 101P3A11 gene products in normal versus cancerous tissues, thereby elucidating the malignant phenotype. Typically, polypeptides from specific regions of the 101P3A11 protein are used to assess the presence of perturbations (such as deletions, insertions, point mutations etc.) in those regions (such as regions containing one or more motifs). Exemplary assays utilize antibodies or T cells targeting 101P3A11-related proteins comprising the amino acid residues of one or more of the biological motifs contained within the 101P3A11 polypeptide sequence in order to evaluate the characteristics of this region in normal versus cancerous tissues or to elicit an immune response to the epitope. Alternatively, 101P3A11-related proteins that contain the amino acid residues of one or more of the biological motifs in the 101P3A11 protein are used to screen for factors that interact with that region of 101P3A11.

101P3A11 protein fragments/subsequences are particularly useful in generating and characterizing domain-specific antibodies (e.g., antibodies recognizing an extracellular or intracellular epitope of an 101P3A11 protein), for identifying agents or cellular factors that bind to 101P3A11 or a particular structural domain thereof, and in various therapeutic and diagnostic contexts, including but not limited to diagnostic assays, cancer vaccines and methods of preparing such vaccines.

Proteins encoded by the 101P3A11 genes, or by analogs, homologs or fragments thereof, have a variety of uses, including but not limited to generating antibodies and in methods for identifying ligands and other agents and cellular constituents that bind to an 101P3A11 gene product. Antibodies raised against an 101P3A11 protein or fragment thereof are useful in diagnostic and prognostic assays, and imaging methodologies in the management of human cancers characterized by expression of 101P3A11 protein, such as those listed in Table I. Such antibodies can be expressed intracellularly and used in methods of treating patients with such cancers. 101P3A11-related nucleic acids or proteins are also used in generating HTL or CTL responses.

Various immunological assays useful for the detection of 101P3A11 proteins are used, including but not limited to various types of radioimmunoassays, enzyme-linked immunosorbent assays (ELISA), enzyme-linked immunofluorescent assays (ELIFA), immunocytochemical methods, and the like. Antibodies can be labeled and used as immunological imaging reagents capable of detecting 101P3A11-expressing cells (e.g., in radioscintigraphic imaging methods). 101P3A11 proteins are also particularly useful in generating cancer vaccines, as further described herein.

IV.) 101P3A11 Antibodies

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Another aspect of the invention provides antibodies that bind to 101P3A11-related proteins. Preferred antibodies specifically bind to a 101P3A11-related protein and do not bind (or bind weakly) to peptides or proteins that are not 101P3A11-related proteins. For example, antibodies bind 101P3A11 can bind 101P3A11-related proteins such as the homologs or analogs thereof.

101P3A11 antibodies of the invention are particularly useful in cancer diagnostic, diagnostic and prognostic assays, and imaging methodologies. Similarly, such antibodies are useful in the treatment, diagnosis, and/or prognosis of other cancers, to the extent 101P3A11 is also expressed or overexpressed in these other cancers. Moreover, intracellularly expressed antibodies (e.g., single chain antibodies) are therapeutically useful in treating cancers in which the expression of 101P3A11 is involved, such as advanced or metastatic prostate cancers.

The invention also provides various immunological assays useful for the detection and quantification of 101P3A11 and mutant 101P3A11-related proteins. Such assays can comprise one or more 101P3A11 antibodies capable of recognizing and binding a 101P3A11-related protein, as appropriate. These assays are performed within various immunological assay formats well known in the art, including but not limited to various types of radioimmunoassays, enzyme-linked immunosorbent assays (ELISA), enzyme-linked immunofluorescent assays (ELIFA), and the like.

Immunological non-antibody assays of the invention also comprise T cell immunogenicity assays (inhibitory or stimulatory) as well as major histocompatibility complex (MHC) binding assays.

In addition, immunological imaging methods capable of detecting prostate cancer and other cancers expressing 101P3A11 are also provided by the invention, including but not limited to radioscintigraphic imaging methods using labeled 101P3A11 antibodies. Such assays are clinically useful in the detection, monitoring, and prognosis of 101P3A11 expressing cancers such as prostate cancer.

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101P3A11 antibodies are also used in methods for purifying a 101P3A11-related protein and for isolating 101P3A11 homologues and related molecules. For example, a method of purifying a 101P3A11-related protein comprises incubating an 101P3A11 antibody, which has been coupled to a solid matrix, with a lysate or other solution containing a 101P3A11-related protein under conditions that permit the 101P3A11 antibody to bind to the 101P3A11-related protein; washing the solid matrix to eliminate impurities; and eluting the 101P3A11-related protein from the coupled antibody. Other uses of the 101P3A11 antibodies of the invention include generating anti-idiotypic antibodies that mimic the 101P3A11 protein.

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Various methods for the preparation of antibodies are well known in the art. For example, antibodies can be prepared by immunizing a suitable mammalian host using a 101P3A11-related protein, peptide, or fragment, in isolated or immunoconjugated form (Antibodies: A Laboratory Manual, CSH Press, Eds., Harlow, and Lane (1988); Harlow, Antibodies, Cold Spring Harbor Press, NY (1989)). In addition, fusion proteins of 101P3A11 can also be used, such as a 101P3A11 GST-fusion protein. In a particular embodiment, a GST fusion protein comprising all or most of the amino acid sequence of Figure 2 or Figure 3 is produced, then used as an immunogen to generate appropriate antibodies. In another embodiment, a 101P3A11-related protein is synthesized and used as an immunogen.

In addition, naked DNA immunization techniques known in the art are used (with or without purified 101P3A11-related protein or 101P3A11 expressing cells) to generate an immune response to the encoded immunogen (for review, see Donnelly et al., 1997, Ann. Rev. Immunol. 15: 617-648).

The amino acid sequence of 101P3A11 as shown in Figure 2 or Figure 3 can be analyzed to select specific regions of the 101P3A11 protein for generating antibodies. For example, hydrophobicity and hydrophilicity analyses of the 101P3A11 amino acid sequence are used to identify hydrophilic regions in the 101P3A11 structure. Regions of the 101P3A11 protein that show immunogenic structure, as well as other regions and domains, can readily be identified using various other methods known in the art, such as Chou-Fasman, Garnier-Robson, Eisenberg, Karplus-Schultz or Jameson-Wolf analysis, Hopp T.P. and Woods K.R., Kyte J. and Doolittle R.F., Janin J., Bhaskaran R., and Ponnuswamy P.K., or, Deleage, G. and Roux B. Thus, each region identified by any of these programs or methods is within the scope of the present invention. Methods for the generation of 101P3A11 antibodies are further illustrated by way of the examples provided herein. Methods for preparing a protein or polypeptide for use as an immunogen are well known in the art. Also well known in the art are methods for preparing immunogenic conjugates of a protein with a carrier, such as BSA, KLH or other carrier protein. In some circumstances, direct conjugation using, for example, carbodiimide reagents are used; in other instances linking reagents such as those supplied by Pierce Chemical Co., Rockford, IL, are effective. Administration of a 101P3A11 immunogen is often conducted by injection over a suitable time period and with use of a suitable adjuvant, as is understood in the art. During the immunization schedule, titers of antibodies can be taken to determine adequacy of antibody formation.

101P3A11 monoclonal antibodies can be produced by various means well known in the art. For example, immortalized cell lines that secrete a desired monoclonal antibody are prepared using the standard hybridoma technology of Kohler and Milstein or modifications that immortalize antibody-producing B cells, as is generally known. Immortalized cell lines that secrete the desired antibodies are screened by immunoassay in which the antigen is a 101P3A11-related protein. When the appropriate immortalized cell culture is identified, the cells can be expanded and antibodies produced either from *in vitro* cultures or from ascites fluid.

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The antibodies or fragments of the invention can also be produced, by recombinant means. Regions that bind specifically to the desired regions of the 101P3A11 protein can also be produced in the context of chimeric or complementarity determining region (CDR) grafted antibodies of multiple species origin. Humanized or human 101P3A11 antibodies can also be produced, and are preferred for use in therapeutic contexts. Methods for humanizing murine and other non-human antibodies, by substituting one or more of the non-human antibody CDRs for corresponding human antibody sequences, are well known (see for example, Jones et al., 1986, Nature 321: 522-525; Riechmann et al., 1988, Nature 332: 323-327; Verhoeyen et al., 1988, Science 239: 1534-1536). See also, Carter et al., 1993, Proc. Natl. Acad. Sci. USA 89: 4285 and Sims et al., 1993, J. Immunol. 151: 2296.

Methods for producing fully human monoclonal antibodies include phage display and transgenic methods (for review, see Vaughan et al., 1998, Nature Biotechnology 16: 535-539). Fully human 101P3A11 monoclonal antibodies can be generated using cloning technologies employing large human Ig gene combinatorial libraries (i.e., phage display) (Griffiths and Hoogenboom, Building an *in vitro* immune system: human antibodies from phage display libraries. In: Protein Engineering of Antibody Molecules for Prophylactic and Therapeutic Applications in Man, Clark, M. (Ed.), Nottingham Academic, pp 45-64 (1993); Burton and Barbas, Human Antibodies from combinatorial libraries. <u>Id.</u>, pp 65-82). Fully human 101P3A11 monoclonal antibodies can also be produced using transgenic mice engineered to contain human immunoglobulin gene loci as described in PCT Patent Application WO98/24893, Kucherlapati and Jakobovits et al., published December 3, 1997 (see also, Jakobovits, 1998, Exp. Opin. Invest. Drugs 7(4): 607-614; U.S. patents 6,162,963 issued 19 December 2000; 6,150,584 issued 12 November 2000; and, 6,114598 issued 5 September 2000). This method avoids the *in vitro* manipulation required with phage display technology and efficiently produces high affinity authentic human antibodies.

Reactivity of 101P3A11 antibodies with an 101P3A11-related protein can be established by a number of well known means, including Western blot, immunoprecipitation, ELISA, and FACS analyses using, as appropriate, 101P3A11-related proteins, 101P3A11-expressing cells or extracts thereof. A 101P3A11 antibody or fragment thereof can be labeled with a detectable marker or conjugated to a second molecule. Suitable detectable markers include, but are not limited to, a radioisotope, a fluorescent compound, a bioluminescent compound, chemiluminescent compound, a metal chelator or an enzyme. Further, bi-specific antibodies specific for two or more 101P3A11 epitopes are generated using methods generally known in the art. Homodimeric antibodies can also be generated by cross-linking techniques known in the art (e.g., Wolff et al., Cancer Res. 53: 2560-2565).

V.) 101P3A11 Cellular Immune Responses

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The mechanism by which T cells recognize antigens has been delineated. Efficacious peptide epitope vaccine compositions of the invention induce a therapeutic or prophylactic immune responses in very broad segments of the world-wide population. For an understanding of the value and efficacy of compositions of the invention that induce cellular immune responses, a brief review of immunology-related technology is provided.

A complex of an HLA molecule and a peptidic antigen acts as the ligand recognized by HLA-restricted T cells (Buus, S. et al., Cell 47:1071, 1986; Babbitt, B. P. et al., Nature 317:359, 1985; Townsend, A. and Bodmer, H., Annu. Rev. Immunol. 7:601, 1989; Germain, R. N., Annu. Rev. Immunol. 11:403, 1993). Through the study of single amino acid substituted antigen analogs and the sequencing of endogenously bound, naturally processed peptides, critical residues that correspond to motifs required for specific binding to HLA antigen molecules have been identified and are

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set forth in Table IV (see also, e.g., Southwood, et al., J. Immunol. 160:3363, 1998; Rammensee, et al., Immunogenetics 41:178, 1995; Rammensee et al., SYFPEITHI, access via World Wide Web at URL syfpeithi.bmi-heidelberg.com/; Sette, A. and Sidney, J. Curr. Opin. Immunol. 10:478, 1998; Engelhard, V. H., Curr. Opin. Immunol. 6:13, 1994; Sette, A. and Grey, H. M., Curr. Opin. Immunol. 4:79, 1992; Sinigaglia, F. and Hammer, J. Curr. Biol. 6:52, 1994; Ruppert et al., Cell 74:929-937, 1993; Kondo et al., J. Immunol. 155:4307-4312, 1995; Sidney et al., J. Immunol. 157:3480-3490, 1996; Sidney et al., Human Immunol. 45:79-93, 1996; Sette, A. and Sidney, J. Immunogenetics 1999 Nov; 50(3-4):201-12, Review).

Furthermore, x-ray crystallographic analyses of HLA-peptide complexes have revealed pockets within the peptide binding cleft/groove of HLA molecules which accommodate, in an allele-specific mode, residues borne by peptide ligands; these residues in turn determine the HLA binding capacity of the peptides in which they are present. (See, e.g., Madden, D.R. Annu. Rev. Immunol. 13:587, 1995; Smith, et al., Immunity 4:203, 1996; Fremont et al., Immunity 8:305, 1998; Stern et al., Structure 2:245, 1994; Jones, E.Y. Curr. Opin. Immunol. 9:75, 1997; Brown, J. H. et al., Nature 364:33, 1993; Guo, H. C. et al., Proc. Natl. Acad. Sci. USA 90:8053, 1993; Guo, H. C. et al., Nature 360:364, 1992; Silver, M. L. et al., Nature 360:367, 1992; Matsumura, M. et al., Science 257:927, 1992; Madden et al., Cell 70:1035, 1992; Fremont, D. H. et al., Science 257:919, 1992; Saper, M. A., Bjorkman, P. J. and Wiley, D. C., J. Mol. Biol. 219:277, 1991.)

Accordingly, the definition of class I and class II allele-specific HLA binding motifs, or class I or class II supermotifs allows identification of regions within a protein that are correlated with binding to particular HLA antigen(s).

Thus, by a process of HLA motif identification, candidates for epitope-based vaccines have been identified; such candidates can be further evaluated by HLA-peptide binding assays to determine binding affinity and/or the time period of association of the epitope and its corresponding HLA molecule. Additional confirmatory work can be performed to select, amongst these vaccine candidates, epitopes with preferred characteristics in terms of population coverage, and/or immunogenicity.

Various strategies can be utilized to evaluate cellular immunogenicity, including:

- 1) Evaluation of primary T cell cultures from normal individuals (see, e.g., Wentworth, P. A. et al., Mol. Immunol. 32:603, 1995; Celis, E. et al., Proc. Natl. Acad. Sci. USA 91:2105, 1994; Tsai, V. et al., J. Immunol. 158:1796, 1997; Kawashima, I. et al., Human Immunol. 59:1, 1998). This procedure involves the stimulation of peripheral blood lymphocytes (PBL) from normal subjects with a test peptide in the presence of antigen presenting cells in vitro over a period of several weeks. T cells specific for the peptide become activated during this time and are detected using, e.g., a lymphokine- or 51Cr-release assay involving peptide sensitized target cells.
- 2) Immunization of HLA transgenic mice (see, e.g., Wentworth, P. A. et al., J. Immunol. 26:97, 1996; Wentworth, P. A. et al., Int. Immunol. 8:651, 1996; Alexander, J. et al., J. Immunol. 159:4753, 1997). For example, in such methods peptides in incomplete Freund's adjuvant are administered subcutaneously to HLA transgenic mice. Several weeks following immunization, splenocytes are removed and cultured in vitro in the presence of test peptide for approximately one week. Peptide-specific T cells are detected using, e.g., a ⁵¹Cr-release assay involving peptide sensitized target cells and target cells expressing endogenously generated antigen.

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3) Demonstration of recall T cell responses from immune individuals who have been either effectively vaccinated and/or from chronically ill patients (see, e.g., Rehermann, B. et al., J. Exp. Med. 181:1047, 1995; Doolan, D. L. et al., Immunity 7:97, 1997; Bertoni, R. et al., J. Clin. Invest. 100:503, 1997; Threlkeld, S. C. et al., J. Immunol. 159:1648, 1997; Diepolder, H. M. et al., J. Virol. 71:6011, 1997). Accordingly, recall responses are detected by culturing PBL from subjects that have been exposed to the antigen due to disease and thus have generated an immune response "naturally", or from patients who were vaccinated against the antigen. PBL from subjects are cultured in vitro for 1-2 weeks in the presence of test peptide plus antigen presenting cells (APC) to allow activation of "memory" T cells, as compared to "naive" T cells. At the end of the culture period, T cell activity is detected using assays including 51Cr release involving peptide-sensitized targets, T cell proliferation, or lymphokine release.

VI.) 101P3A11 Transgenic Animals

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Nucleic acids that encode a 101P3A11-related protein can also be used to generate either transgenic animals or "knock out" animals which, in turn, are useful in the development and screening of therapeutically useful reagents. In accordance with established techniques, cDNA encoding 101P3A11 can be used to clone genomic DNA that encodes 101P3A11. The cloned genomic sequences can then be used to generate transgenic animals containing cells that express DNA that encode 101P3A11. Methods for generating transgenic animals, particularly animals such as mice or rats, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 issued 12 April 1988, and 4,870,009 issued 26 September 1989. Typically, particular cells would be targeted for 101P3A11 transgene incorporation with tissue-specific enhancers.

Transgenic animals that include a copy of a transgene encoding 101P3A11 can be used to examine the effect of increased expression of DNA that encodes 101P3A11. Such animals can be used as tester animals for reagents thought to confer protection from, for example, pathological conditions associated with its overexpression. In accordance with this aspect of the invention, an animal is treated with a reagent and a reduced incidence of a pathological condition, compared to untreated animals that bear the transgene, would indicate a potential therapeutic intervention for the pathological condition.

Alternatively, non-human homologues of 101P3A11 can be used to construct a 101P3A11 "knock out" animal that has a defective or altered gene encoding 101P3A11 as a result of homologous recombination between the endogenous gene encoding 101P3A11 and altered genomic DNA encoding 101P3A11 introduced into an embryonic cell of the animal. For example, cDNA that encodes 101P3A11 can be used to clone genomic DNA encoding 101P3A11 can be deleted or replaced with established techniques. A portion of the genomic DNA encoding 101P3A11 can be deleted or replaced with another gene, such as a gene encoding a selectable marker that can be used to monitor integration. Typically, several kilobases of unaltered flanking DNA (both at the 5' and 3' ends) are included in the vector (see, e.g., Thomas and Capecchi, Cell, 51:503 (1987) for a description of homologous recombination vectors). The vector is introduced into an embryonic stem cell line (e.g., by electroporation) and cells in which the introduced DNA has homologously recombined with the endogenous DNA are selected (see, e.g.,, Li et al., Cell, 69:915 (1992)). The selected cells are then injected into a blastocyst of an animal (e.g., a mouse or rat) to form aggregation chimeras (see, e.g.,, Bradley, in *Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, E. J. Robertson, ed. (IRL, Oxford, 1987), pp. 113-152). A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal, and the embryo brought to term to create a "knock out" animal. Progeny harboring the homologously

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recombined DNA in their germ cells can be identified by standard techniques and used to breed animals in which all cells of the animal contain the homologously recombined DNA. Knock out animals can be characterized, for example, for their ability to defend against certain pathological conditions or for their development of pathological conditions due to absence of the 101P3A11 polypeptide.

VII.) Methods for the Detection of 101P3A11

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Another aspect of the present invention relates to methods for detecting 101P3A11 polynucleotides and 101P3A11-related proteins, as well as methods for identifying a cell that expresses 101P3A11. The expression profile of 101P3A11 makes it a diagnostic marker for metastasized disease. Accordingly, the status of 101P3A11 gene products provides information useful for predicting a variety of factors including susceptibility to advanced stage disease, rate of progression, and/or tumor aggressiveness. As discussed in detail herein, the status of 101P3A11 gene products in patient samples can be analyzed by a variety protocols that are well known in the art including immunohistochemical analysis, the variety of Northern blotting techniques including *in situ* hybridization, RT-PCR analysis (for example on laser capture micro-dissected samples), Western blot analysis and tissue array analysis.

More particularly, the invention provides assays for the detection of 101P3A11 polynucleotides in a biological sample, such as serum, bone, prostate, and other tissues, urine, semen, cell preparations, and the like. Detectable 101P3A11 polynucleotides include, for example, a 101P3A11 gene or fragment thereof, 101P3A11 mRNA, alternative splice variant 101P3A11 mRNAs, and recombinant DNA or RNA molecules that contain a 101P3A11 polynucleotide. A number of methods for amplifying and/or detecting the presence of 101P3A11 polynucleotides are well known in the art and can be employed in the practice of this aspect of the invention.

In one embodiment, a method for detecting an 101P3A11 mRNA in a biological sample comprises producing cDNA from the sample by reverse transcription using at least one primer; amplifying the cDNA so produced using an 101P3A11 polynucleotides as sense and antisense primers to amplify 101P3A11 cDNAs therein; and detecting the presence of the amplified 101P3A11 cDNA. Optionally, the sequence of the amplified 101P3A11 cDNA can be determined.

In another embodiment, a method of detecting a 101P3A11 gene in a biological sample comprises first isolating genomic DNA from the sample; amplifying the isolated genomic DNA using 101P3A11 polynucleotides as sense and antisense primers; and detecting the presence of the amplified 101P3A11 gene. Any number of appropriate sense and antisense probe combinations can be designed from the nucleotide sequence provided for the 101P3A11 (Figure 2) and used for this purpose.

The invention also provides assays for detecting the presence of an 101P3A11 protein in a tissue or other biological sample such as serum, semen, bone, prostate, urine, cell preparations, and the like. Methods for detecting a 101P3A11-related protein are also well known and include, for example, immunoprecipitation, immunohistochemical analysis, Western blot analysis, molecular binding assays, ELISA, ELIFA and the like. For example, a method of detecting the presence of a 101P3A11-related protein in a biological sample comprises first contacting the sample with a 101P3A11 antibody, a 101P3A11-reactive fragment thereof, or a recombinant protein containing an antigen binding region of a 101P3A11 antibody; and then detecting the binding of 101P3A11-related protein in the sample.

Methods for identifying a cell that expresses 101P3A11 are also within the scope of the invention. In one embodiment, an assay for identifying a cell that expresses a 101P3A11 gene comprises detecting the presence of 101P3A11

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mRNA in the cell. Methods for the detection of particular mRNAs in cells are well known and include, for example, hybridization assays using complementary DNA probes (such as *in situ* hybridization using labeled 101P3A11 riboprobes, Northern blot and related techniques) and various nucleic acid amplification assays (such as RT-PCR using complementary primers specific for 101P3A11, and other amplification type detection methods, such as, for example, branched DNA, SISBA, TMA and the like). Alternatively, an assay for identifying a cell that expresses a 101P3A11 gene comprises detecting the presence of 101P3A11-related protein in the cell or secreted by the cell. Various methods for the detection of proteins are well known in the art and are employed for the detection of 101P3A11-related proteins and cells that express 101P3A11-related proteins.

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101P3A11 expression analysis is also useful as a tool for identifying and evaluating agents that modulate 101P3A11 gene expression. For example, 101P3A11 expression is significantly upregulated in prostate cancer, and is expressed in cancers of the tissues listed in Table I. Identification of a molecule or biological agent that inhibits 101P3A11 expression or over-expression in cancer cells is of therapeutic value. For example, such an agent can be identified by using a screen that quantifies 101P3A11 expression by RT-PCR, nucleic acid hybridization or antibody binding.

VIII.) Methods for Monitoring the Status of 101P3A11-related Genes and Their Products

Oncogenesis is known to be a multistep process where cellular growth becomes progressively dysregulated and cells progress from a normal physiological state to precancerous and then cancerous states (see, e.g., Alers et al., Lab Invest. 77(5): 437-438 (1997) and Isaacs et al., Cancer Surv. 23: 19-32 (1995)). In this context, examining a biological sample for evidence of dysregulated cell growth (such as aberrant 101P3A11 expression in cancers) allows for early detection of such aberrant physiology, before a pathologic state such as cancer has progressed to a stage that therapeutic options are more limited and or the prognosis is worse. In such examinations, the status of 101P3A11 in a biological sample of interest can be compared, for example, to the status of 101P3A11 in a corresponding normal sample (e.g. a sample from that individual or alternatively another individual that is not affected by a pathology). An alteration in the status of 101P3A11 in the biological sample (as compared to the normal sample) provides evidence of dysregulated cellular growth. In addition to using a biological sample that is not affected by a pathology as a normal sample, one can also use a predetermined normative value such as a predetermined normal level of mRNA expression (see, e.g., Grever et al., J. Comp. Neurol. 1996 Dec 9;376(2):306-14 and U.S. Patent No. 5,837,501) to compare 101P3A11 status in a sample.

The term "status" in this context is used according to its art accepted meaning and refers to the condition or state of a gene and its products. Typically, skilled artisans use a number of parameters to evaluate the condition or state of a gene and its products. These include, but are not limited to the location of expressed gene products (including the location of 101P3A11 expressing cells) as well as the level, and biological activity of expressed gene products (such as 101P3A11 mRNA, polynucleotides and polypeptides). Typically, an alteration in the status of 101P3A11 comprises a change in the location of 101P3A11 and/or 101P3A11 expressing cells and/or an increase in 101P3A11 mRNA and/or protein expression.

101P3A11 status in a sample can be analyzed by a number of means well known in the art, including without limitation, immunohistochemical analysis, *in situ* hybridization, RT-PCR analysis on laser capture micro-dissected samples, Western blot analysis, and tissue array analysis. Typical protocols for evaluating the status of the 101P3A11 gene and gene

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products are found, for example in Ausubel et al. eds., 1995, Current Protocols In Molecular Biology, Units 2 (Northern Blotting), 4 (Southern Blotting), 15 (Immunoblotting) and 18 (PCR Analysis). Thus, the status of 101P3A11 in a biological sample is evaluated by various methods utilized by skilled artisans including, but not limited to genomic Southern analysis (to examine, for example perturbations in the 101P3A11 gene), Northern analysis and/or PCR analysis of 101P3A11 mRNA (to examine, for example alterations in the polynucleotide sequences or expression levels of 101P3A11 mRNAs), and, Western and/or immunohistochemical analysis (to examine, for example alterations in polypeptide sequences, alterations in polypeptide localization within a sample, alterations in expression levels of 101P3A11 proteins and/or associations of 101P3A11 proteins with polypeptide binding partners). Detectable 101P3A11 polynucleotides include, for example, a 101P3A11 gene or fragment thereof, 101P3A11 mRNA, alternative splice variants, 101P3A11 mRNAs, and recombinant DNA or RNA molecules containing a 101P3A11 polynucleotide.

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The expression profile of 101P3A11 makes it a diagnostic marker for local and/or metastasized disease, and provides information on the growth or oncogenic potential of a biological sample. In particular, the status of 101P3A11 provides information useful for predicting susceptibility to particular disease stages, progression, and/or tumor aggressiveness. The invention provides methods and assays for determining 101P3A11 status and diagnosing cancers that express 101P3A11, such as cancers of the tissues listed in Table I. For example, because 101P3A11 mRNA is so highly expressed in prostate and other cancers relative to normal prostate tissue, assays that evaluate the levels of 101P3A11 mRNA transcripts or proteins in a biological sample can be used to diagnose a disease associated with 101P3A11 dysregulation, and can provide prognostic information useful in defining appropriate therapeutic options.

The expression status of 101P3A11 provides information including the presence, stage and location of dysplastic, precancerous and cancerous cells, predicting susceptibility to various stages of disease, and/or for gauging tumor aggressiveness. Moreover, the expression profile makes it useful as an imaging reagent for metastasized disease. Consequently, an aspect of the invention is directed to the various molecular prognostic and diagnostic methods for examining the status of 101P3A11 in biological samples such as those from individuals suffering from, or suspected of suffering from a pathology characterized by dysregulated cellular growth, such as cancer.

As described above, the status of 101P3A11 in a biological sample can be examined by a number of well-known procedures in the art. For example, the status of 101P3A11 in a biological sample taken from a specific location in the body can be examined by evaluating the sample for the presence or absence of 101P3A11 expressing cells (e.g. those that express 101P3A11 mRNAs or proteins). This examination can provide evidence of dysregulated cellular growth, for example, when 101P3A11-expressing cells are found in a biological sample that does not normally contain such cells (such as a lymph node), because such alterations in the status of 101P3A11 in a biological sample are often associated with dysregulated cellular growth. Specifically, one indicator of dysregulated cellular growth is the metastases of cancer cells from an organ of origin (such as the prostate) to a different area of the body (such as a lymph node). In this context, evidence of dysregulated cellular growth is important for example because occult lymph node metastases can be detected in a substantial proportion of patients with prostate cancer, and such metastases are associated with known predictors of disease progression (see, e.g., Murphy et al., Prostate 42(4): 315-317 (2000);Su et al., Semin. Surg. Oncol. 18(1): 17-28 (2000) and Freeman et al., J Urol 1995 Aug 154(2 Pt 1):474-8).

In one aspect, the invention provides methods for monitoring 101P3A11 gene products by determining the status of 101P3A11 gene products expressed by cells from an individual suspected of having a disease associated with

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dysregulated cell growth (such as hyperplasia or cancer) and then comparing the status so determined to the status of 101P3A11 gene products in a corresponding normal sample. The presence of aberrant 101P3A11 gene products in the test sample relative to the normal sample provides an indication of the presence of dysregulated cell growth within the cells of the individual.

In another aspect, the invention provides assays useful in determining the presence of cancer in an individual, comprising detecting a significant increase in 101P3A11 mRNA or protein expression in a test cell or tissue sample relative to expression levels in the corresponding normal cell or tissue. The presence of 101P3A11 mRNA can, for example, be evaluated in tissue samples including but not limited to those listed in Table I. The presence of significant 101P3A11 expression in any of these tissues is useful to indicate the emergence, presence and/or severity of a cancer, since the corresponding normal tissues do not express 101P3A11 mRNA or express it at lower levels.

In a related embodiment, 101P3A11 status is determined at the protein level rather than at the nucleic acid level. For example, such a method comprises determining the level of 101P3A11 protein expressed by cells in a test tissue sample and comparing the level so determined to the level of 101P3A11 expressed in a corresponding normal sample. In one embodiment, the presence of 101P3A11 protein is evaluated, for example, using immunohistochemical methods. 101P3A11 antibodies or binding partners capable of detecting 101P3A11 protein expression are used in a variety of assay formats well known in the art for this purpose.

In a further embodiment, one can evaluate the status of 101P3A11 nucleotide and amino acid sequences in a biological sample in order to identify perturbations in the structure of these molecules. These perturbations can include insertions, deletions, substitutions and the like. Such evaluations are useful because perturbations in the nucleotide and amino acid sequences are observed in a large number of proteins associated with a growth dysregulated phenotype (see, e.g., Marrogi et al., 1999, J. Cutan. Pathol. 26(8):369-378). For example, a mutation in the sequence of 101P3A11 may be indicative of the presence or promotion of a tumor. Such assays therefore have diagnostic and predictive value where a mutation in 101P3A11 indicates a potential loss of function or increase in tumor growth.

A wide variety of assays for observing perturbations in nucleotide and amino acid sequences are well known in the art. For example, the size and structure of nucleic acid or amino acid sequences of 101P3A11 gene products are observed by the Northern, Southern, Western, PCR and DNA sequencing protocols discussed herein. In addition, other methods for observing perturbations in nucleotide and amino acid sequences such as single strand conformation polymorphism analysis are well known in the art (see, e.g., U.S. Patent Nos. 5,382,510 issued 7 September 1999, and 5,952,170 issued 17 January 1995).

Additionally, one can examine the methylation status of the 101P3A11 gene in a biological sample. Aberrant demethylation and/or hypermethylation of CpG islands in gene 5' regulatory regions frequently occurs in immortalized and transformed cells, and can result in altered expression of various genes. For example, promoter hypermethylation of the piclass glutathione S-transferase (a protein expressed in normal prostate but not expressed in >90% of prostate carcinomas) appears to permanently silence transcription of this gene and is the most frequently detected genomic alteration in prostate carcinomas (De Marzo et al., Am. J. Pathol. 155(6): 1985-1992 (1999)). In addition, this alteration is present in at least 70% of cases of high-grade prostatic intraepithelial neoplasia (PIN) (Brooks et al, Cancer Epidemiol. Biomarkers Prev., 1998, 7:531-536). In another example, expression of the LAGE-I tumor specific gene (which is not expressed in normal prostate but is expressed in 25-50% of prostate cancers) is induced by deoxy-

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azacytidine in lymphoblastoid cells, suggesting that tumoral expression is due to demethylation (Lethe et al., Int. J. Cancer 76(6): 903-908 (1998)). A variety of assays for examining methylation status of a gene are well known in the art. For example, one can utilize, in Southern hybridization approaches, methylation-sensitive restriction enzymes which cannot cleave sequences that contain methylated CpG sites to assess the methylation status of CpG islands. In addition, MSP (methylation specific PCR) can rapidly profile the methylation status of all the CpG sites present in a CpG island of a given gene. This procedure involves initial modification of DNA by sodium bisulfite (which will convert all unmethylated cytosines to uracil) followed by amplification using primers specific for methylated versus unmethylated DNA. Protocols involving methylation interference can also be found for example in Current Protocols In Molecular Biology, Unit 12, Frederick M. Ausubel et al. eds., 1995.

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Gene amplification is an additional method for assessing the status of 101P3A11. Gene amplification is measured in a sample directly, for example, by conventional Southern blotting or Northern blotting to quantitate the transcription of mRNA (Thomas, 1980, Proc. Natl. Acad. Sci. USA, 77:5201-5205), dot blotting (DNA analysis), or *in situ* hybridization, using an appropriately labeled probe, based on the sequences provided herein. Alternatively, antibodies are employed that recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn are labeled and the assay carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Biopsied tissue or peripheral blood can be conveniently assayed for the presence of cancer cells using for example, Northern, dot blot or RT-PCR analysis to detect 101P3A11 expression. The presence of RT-PCR amplifiable 101P3A11 mRNA provides an indication of the presence of cancer. RT-PCR assays are well known in the art. RT-PCR detection assays for tumor cells in peripheral blood are currently being evaluated for use in the diagnosis and management of a number of human solid tumors. In the prostate cancer field, these include RT-PCR assays for the detection of cells expressing PSA and PSM (Verkaik et al., 1997, Urol. Res. 25:373-384; Ghossein et al., 1995, J. Clin. Oncol. 13:1195-2000; Heston et al., 1995, Clin. Chem. 41:1687-1688).

A further aspect of the invention is an assessment of the susceptibility that an individual has for developing cancer. In one embodiment, a method for predicting susceptibility to cancer comprises detecting 101P3A11 mRNA or 101P3A11 protein in a tissue sample, its presence indicating susceptibility to cancer, wherein the degree of 101P3A11 mRNA expression correlates to the degree of susceptibility. In a specific embodiment, the presence of 101P3A11 in prostate or other tissue is examined, with the presence of 101P3A11 in the sample providing an indication of prostate cancer susceptibility (or the emergence or existence of a prostate tumor). Similarly, one can evaluate the integrity 101P3A11 nucleotide and amino acid sequences in a biological sample, in order to identify perturbations in the structure of these molecules such as insertions, deletions, substitutions and the like. The presence of one or more perturbations in 101P3A11 gene products in the sample is an indication of cancer susceptibility (or the emergence or existence of a tumor).

The invention also comprises methods for gauging tumor aggressiveness. In one embodiment, a method for gauging aggressiveness of a tumor comprises determining the level of 101P3A11 mRNA or 101P3A11 protein expressed by tumor cells, comparing the level so determined to the level of 101P3A11 mRNA or 101P3A11 protein expressed in a corresponding normal tissue taken from the same individual or a normal tissue reference sample, wherein the degree of 101P3A11 mRNA or 101P3A11 protein expression in the tumor sample relative to the normal sample indicates the degree

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of aggressiveness. In a specific embodiment, aggressiveness of a tumor is evaluated by determining the extent to which 101P3A11 is expressed in the tumor cells, with higher expression levels indicating more aggressive tumors. Another embodiment is the evaluation of the integrity of 101P3A11 nucleotide and amino acid sequences in a biological sample, in order to identify perturbations in the structure of these molecules such as insertions, deletions, substitutions and the like. The presence of one or more perturbations indicates more aggressive tumors.

Another embodiment of the invention is directed to methods for observing the progression of a malignancy in an individual over time. In one embodiment, methods for observing the progression of a malignancy in an individual over time comprise determining the level of 101P3A11 mRNA or 101P3A11 protein expressed by cells in a sample of the tumor, comparing the level so determined to the level of 101P3A11 mRNA or 101P3A11 protein expressed in an equivalent tissue sample taken from the same individual at a different time, wherein the degree of 101P3A11 mRNA or 101P3A11 protein expression in the tumor sample over time provides information on the progression of the cancer. In a specific embodiment, the progression of a cancer is evaluated by determining 101P3A11 expression in the tumor cells over time, where increased expression over time indicates a progression of the cancer. Also, one can evaluate the integrity 101P3A11 nucleotide and amino acid sequences in a biological sample in order to identify perturbations in the structure of these molecules such as insertions, deletions, substitutions and the like, where the presence of one or more perturbations indicates a progression of the cancer.

The above diagnostic approaches can be combined with any one of a wide variety of prognostic and diagnostic protocols known in the art. For example, another embodiment of the invention is directed to methods for observing a coincidence between the expression of 101P3A11 gene and 101P3A11 gene products (or perturbations in 101P3A11 gene and 101P3A11 gene products) and a factor that is associated with malignancy, as a means for diagnosing and prognosticating the status of a tissue sample. A wide variety of factors associated with malignancy can be utilized, such as the expression of genes associated with malignancy (e.g. PSA, PSCA and PSM expression for prostate cancer etc.) as well as gross cytological observations (see, e.g., Bocking et al., 1984, Anal. Quant. Cytol. 6(2):74-88; Epstein, 1995, Hum. Pathol. 26(2):223-9; Thorson et al., 1998, Mod. Pathol. 11(6):543-51; Baisden et al., 1999, Am. J. Surg. Pathol. 23(8):918-24). Methods for observing a coincidence between the expression of 101P3A11 gene and 101P3A11 gene products (or perturbations in 101P3A11 gene and 101P3A11 gene products) and another factor that is associated with malignancy are useful, for example, because the presence of a set of specific factors that coincide with disease provides information crucial for diagnosing and prognosticating the status of a tissue sample.

In one embodiment, methods for observing a coincidence between the expression of 101P3A11 gene and 101P3A11 gene products (or perturbations in 101P3A11 gene and 101P3A11 gene products) and another factor associated with malignancy entails detecting the overexpression of 101P3A11 mRNA or protein in a tissue sample, detecting the overexpression of PSA mRNA or protein in a tissue sample (or PSCA or PSM expression), and observing a coincidence of 101P3A11 mRNA or protein and PSA mRNA or protein overexpression (or PSCA or PSM expression). In a specific embodiment, the expression of 101P3A11 and PSA mRNA in prostate tissue is examined, where the coincidence of 101P3A11 and PSA mRNA overexpression in the sample indicates the existence of prostate cancer, prostate cancer susceptibility or the emergence or status of a prostate tumor.

Methods for detecting and quantifying the expression of 101P3A11 mRNA or protein are described herein, and standard nucleic acid and protein detection and quantification technologies are well known in the art. Standard methods for

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the detection and quantification of 101P3A11 mRNA include *in situ* hybridization using labeled 101P3A11 riboprobes, Northern blot and related techniques using 101P3A11 polynucleotide probes, RT-PCR analysis using primers specific for 101P3A11, and other amplification type detection methods, such as, for example, branched DNA, SISBA, TMA and the like. In a specific embodiment, semi-quantitative RT-PCR is used to detect and quantify 101P3A11 mRNA expression. Any number of primers capable of amplifying 101P3A11 can be used for this purpose, including but not limited to the various primer sets specifically described herein. In a specific embodiment, polyclonal or monoclonal antibodies specifically reactive with the wild-type 101P3A11 protein can be used in an immunohistochemical assay of biopsied tissue.

IX.) Identification of Molecules that Interact with 101P3A11

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The 101P3A11 protein and nucleic acid sequences disclosed herein allow a skilled artisan to identify proteins, small molecules and other agents that interact with 101P3A11, as well as pathways activated by 101P3A11 via any one of a variety of art accepted protocols. For example, one can utilize one of the so-called interaction trap systems (also referred to as the "two-hybrid assay"). In such systems, molecules interact and reconstitute a transcription factor which directs expression of a reporter gene, whereupon the expression of the reporter gene is assayed. Other systems identify protein-protein interactions *in vivo* through reconstitution of a eukaryotic transcriptional activator, see, e.g., U.S. Patent Nos. 5,955,280 issued 21 September 1999, 5,925,523 issued 20 July 1999, 5,846,722 issued 8 December 1998 and 6,004,746 issued 21 December 1999. Algorithms are also available in the art for genome-based predictions of protein function (see, e.g., Marcotte, et al., Nature 402: 4 November 1999, 83-86).

Alternatively one can screen peptide libraries to identify molecules that interact with 101P3A11 protein sequences. In such methods, peptides that bind to 101P3A11 are identified by screening libraries that encode a random or controlled collection of amino acids. Peptides encoded by the libraries are expressed as fusion proteins of bacteriophage coat proteins, the bacteriophage particles are then screened against the 101P3A11 protein.

Accordingly, peptides having a wide variety of uses, such as therapeutic, prognostic or diagnostic reagents, are thus identified without any prior information on the structure of the expected ligand or receptor molecule. Typical peptide libraries and screening methods that can be used to identify molecules that interact with 101P3A11 protein sequences are disclosed for example in U.S. Patent Nos. 5,723,286 issued 3 March 1998 and 5,733,731 issued 31 March 1998.

Alternatively, cell lines that express 101P3A11 are used to identify protein-protein interactions mediated by 101P3A11. Such interactions can be examined using immunoprecipitation techniques (see, e.g., Hamilton B.J., et al. Biochem. Biophys. Res. Commun. 1999, 261:646-51). 101P3A11 protein can be immunoprecipitated from 101P3A11-expressing cell lines using anti-101P3A11 antibodies. Alternatively, antibodies against His-tag can be used in a cell line engineered to express fusions of 101P3A11 and a His-tag (vectors mentioned above). The immunoprecipitated complex can be examined for protein association by procedures such as Western blotting, ³⁵S-methionine labeling of proteins, protein microsequencing, silver staining and two-dimensional gel electrophoresis.

Small molecules and ligands that interact with 101P3A11 can be identified through related embodiments of such screening assays. For example, small molecules can be identified that interfere with protein function, including molecules that interfere with 101P3A11's ability to mediate phosphorylation and de-phosphorylation, interaction with DNA or RNA molecules as an indication of regulation of cell cycles, second messenger signaling or tumorigenesis. Similarly, small molecules that modulate 101P3A11-related ion channel, protein pump, or cell communication functions

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are identified and used to treat patients that have a cancer that expresses 101P3A11 (see, e.g., Hille, B., Ionic Channels of Excitable Membranes 2nd Ed., Sinauer Assoc., Sunderland, MA, 1992). Moreover, ligands that regulate 101P3A11 function can be identified based on their ability to bind 101P3A11 and activate a reporter construct. Typical methods are discussed for example in U.S. Patent No. 5,928,868 issued 27 July 1999, and include methods for forming hybrid ligands in which at least one ligand is a small molecule. In an illustrative embodiment, cells engineered to express a fusion protein of 101P3A11 and a DNA-binding protein are used to co-express a fusion protein of a hybrid ligand/small molecule and a cDNA library transcriptional activator protein. The cells further contain a reporter gene, the expression of which is conditioned on the proximity of the first and second fusion proteins to each other, an event that occurs only if the hybrid ligand binds to target sites on both hybrid proteins. Those cells that express the reporter gene are selected and the unknown small molecule or the unknown ligand is identified. This method provides a means of identifying modulators which activate or inhibit 101P3A11.

An embodiment of this invention comprises a method of screening for a molecule that interacts with an 101P3A11 amino acid sequence shown in Figure 2 or Figure 3, comprising the steps of contacting a population of molecules with the 101P3A11 amino acid sequence, allowing the population of molecules and the 101P3A11 amino acid sequence to interact under conditions that facilitate an interaction, determining the presence of a molecule that interacts with the 101P3A11 amino acid sequence, and then separating molecules that do not interact with the 101P3A11 amino acid sequence from molecules that do. In a specific embodiment, the method further comprises purifying, characterizing and identifying a molecule that interacts with the 101P3A11 amino acid sequence. The identified molecule can be used to modulate a function performed by 101P3A11. In a preferred embodiment, the 101P3A11 amino acid sequence is contacted with a library of peptides.

Non-limiting examples of small molecules include compounds that bind members of the olfactory receptor family of GPCR., ligands including hormones, neuropeptides, chemokines, odorants, phospholipids, and functional equivalents thereof that bind and preferably inhibit 101P3A11 protein function. Such non-limiting small molecules preferably have a molecular weight of less than about 10 kDa, more preferably below about 9, about 8, about 7, about 6, about 5 or about 4 kDa; physically associates with, or binds, 101P3A11 protein; is not found in naturally occurring metabolic pathways; and/or are more soluble in aqueous than non-aqueous solutions.

X.) Therapeutic Methods and Compositions

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The identification of 101P3A11 as a protein that is normally expressed in a restricted set of tissues, but which is also expressed in prostate and other cancers, opens a number of therapeutic approaches to the treatment of such cancers. As contemplated herein, 101P3A11 functions as a transcription factor involved in activating tumor-promoting genes or repressing genes that block tumorigenesis.

Accordingly, therapeutic approaches that inhibit the activity of the 101P3A11 protein are useful for patients suffering from a cancer that expresses 101P3A11. These therapeutic approaches generally fall into two classes. One class comprises various methods for inhibiting the binding or association of the 101P3A11 protein with its binding partner or with other proteins. Another class comprises a variety of methods for inhibiting the transcription of the 101P3A11 gene or translation of 101P3A11 mRNA.

X.A.) Anti-Cancer Vaccines

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The invention provides cancer vaccines comprising a 101P3A11-related protein or 101P3A11-related nucleic acid. In view of the expression of 101P3A11, cancer vaccines prevent and/or treat 101P3A11-expressing cancers with minimal or no effects on non-target tissues. The use of a tumor antigen in a vaccine that generates humoral and/or cell-mediated immune responses as anti-cancer therapy is well known in the art and has been employed in prostate cancer using human PSMA and rodent PAP immunogens (Hodge et al., 1995, Int. J. Cancer 63:231-237; Fong et al., 1997, J. Immunol. 159:3113-3117).

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Such methods can be readily practiced by employing a 101P3A11-related protein, or an 101P3A11-encoding nucleic acid molecule and recombinant vectors capable of expressing and presenting the 101P3A11 immunogen (which typically comprises a number of antibody or T cell epitopes). Skilled artisans understand that a wide variety of vaccine systems for delivery of immunoreactive epitopes are known in the art (see, e.g., Heryln et al., Ann Med 1999 Feb 31(1):66-78; Maruyama et al., Cancer Immunol Immunother 2000 Jun 49(3):123-32) Briefly, such methods of generating an immune response (e.g. humoral and/or cell-mediated) in a mammal, comprise the steps of: exposing the mammal's immune system to an immunoreactive epitope (e.g. an epitope present in the 101P3A11 protein shown in SEQ ID NO: 703 or analog or homolog thereof) so that the mammal generates an immune response that is specific for that epitope (e.g. generates antibodies that specifically recognize that epitope). In a preferred method, the 101P3A11 immunogen contains a biological motif, see e.g., Tables V-XVIII or XXVI-XXVIII, or a peptide of a size range from 101P3A11 indicated in Figure 5, Figure 6, Figure 7, Figure 8, and Figure 9.

The entire 101P3A11 protein, immunogenic regions or epitopes thereof can be combined and delivered by various means. Such vaccine compositions can include, for example, lipopeptides (e.g., Vitiello, A. et al., J. Clin. Invest. 95:341, 1995), peptide compositions encapsulated in poly(DL-lactide-co-glycolide) ("PLG") microspheres (see, e.g., Eldridge, et al., Molec. Immunol. 28:287-294, 1991: Alonso et al., Vaccine 12:299-306, 1994; Jones et al., Vaccine 13:675-681, 1995), peptide compositions contained in immune stimulating complexes (ISCOMS) (see, e.g., Takahashi et al., Nature 344:873-875, 1990; Hu et al., Clin Exp Immunol. 113:235-243, 1998), multiple antigen peptide systems (MAPs) (see e.g., Tam, J. P., Proc. Natl. Acad. Sci. U.S.A. 85;5409-5413, 1988; Tam, J.P., J. Immunol. Methods 196:17-32, 1996), peptides formulated as multivalent peptides; peptides for use in ballistic delivery systems, typically crystallized peptides, viral delivery vectors (Perkus, M. E. et al., In: Concepts in vaccine development, Kaufmann, S. H. E., ed., p. 379, 1996; Chakrabarti, S. et al., Nature 320:535, 1986; Hu, S. L. et al., Nature 320:537, 1986; Kieny, M.-P. et al., AIDS Bio/Technology 4:790, 1986; Top, F. H. et al., J. Infect. Dis. 124:148, 1971; Chanda, P. K. et al., Virology 175:535, 1990), particles of viral or synthetic origin (e.g., Kofler, N. et al., J. Immunol. Methods. 192:25, 1996; Eldridge, J. H. et al., Sem. Hematol. 30:16, 1993; Falo, L. D., Jr. et al., Nature Med. 7:649, 1995), adjuvants (Warren, H. S., Vogel, F. R., and Chedid, L. A. Annu. Rev. Immunol. 4:369, 1986; Gupta, R. K. et al., Vaccine 11:293, 1993), liposomes (Reddy, R. et al., J. Immunol. 148:1585, 1992; Rock, K. L., Immunol. Today 17:131, 1996), or, naked or particle absorbed cDNA (Ulmer, J. B. et al., Science 259:1745, 1993; Robinson, H. L., Hunt, L. A., and Webster, R. G., Vaccine 11:957, 1993; Shiver, J. W. et al., In: Concepts in vaccine development, Kaufmann, S. H. E., ed., p. 423, 1996; Cease, K. B., and Berzofsky, J. A., Annu. Rev. Immunol. 12:923, 1994 and Eldridge, J. H. et al., Sem. Hematol. 30:16, 1993). Toxin-targeted delivery technologies, also known as receptor mediated targeting, such as those of Avant Immunotherapeutics, Inc. (Needham, Massachusetts) may also be used.

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In patients with 101P3A11-associated cancer, the vaccine compositions of the invention can also be used in conjunction with other treatments used for cancer, *e.g.*, surgery, chemotherapy, drug therapies, radiation therapies, *etc.* including use in combination with immune adjuvants such as IL-2, IL-12, GM-CSF, and the like.

Cellular Vaccines:

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CTL epitopes can be determined using specific algorithms to identify peptides within 101P3A11 protein that bind corresponding HLA alleles (see e.g., Table IV; Epimer™ and Epimatrix™, Brown University (URL www.brown.edu/Research/TB-HIV Lab/epimatrix/epimatrix.html); and, BIMAS, (URL bimas.dcrt.nih.gov/; SYFPEITHI at URL syfpeithi.bmi-heidelberg.com/). In a preferred embodiment, the 101P3A11 immunogen contains one or more amino acid sequences identified using techniques well known in the art, such as the sequences shown in Tables V-XVIII or XXVI-XXVIII or a peptide of 8, 9, 10 or 11 amino acids specified by an HLA Class I motif/supermotif (e.g., Table IV (A), Table IV (D), or Table IV (E)) and/or a peptide of at least 9 amino acids that comprises an HLA Class II motif/supermotif (e.g., Table IV (B) or Table IV (C)). As is appreciated in the art, the HLA Class I binding groove is essentially closed ended so that peptides of only a particular size range can fit into the groove and be bound, generally HLA Class I epitopes are 8, 9, 10, or 11 amino acids long. In contrast, the HLA Class II binding groove is essentially open ended; therefore a peptide of about 9 or more amino acids can be bound by an HLA Class II molecule. Due to the binding groove differences between HLA Class I and II, HLA Class I motifs are length specific, i.e., position two of a Class I motif is the second amino acid in an amino to carboxyl direction of the peptide. The amino acid positions in a Class II motif are relative only to each other, not the overall peptide, i.e., additional amino acids can be attached to the amino and/or carboxyl termini of a motif-bearing sequence. HLA Class II epitopes are often 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 amino acids long, or longer than 25 amino acids.

Antibody-based Vaccines

A wide variety of methods for generating an immune response in a mammal are known in the art (for example as the first step in the generation of hybridomas). Methods of generating an immune response in a mammal comprise exposing the mammal's immune system to an immunogenic epitope on a protein (e.g. the 101P3A11 protein) so that an immune response is generated. In one embodiment the protein can be an anti idiotypic antibody. A typical embodiment consists of a method for generating an immune response to 101P3A11 in a host, by contacting the host with a sufficient amount of at least one 101P3A11 B cell or cytotoxic T-cell epitope or analog thereof; and at least one periodic interval thereafter re-contacting the host with the 101P3A11 B cell or cytotoxic T-cell epitope or analog thereof. A specific embodiment consists of a method of generating an immune response against a 101P3A11-related protein or a man-made multiepitopic peptide comprising: administering 101P3A11 immunogen (e.g. the 101P3A11 protein or a peptide fragment thereof, an 101P3A11 fusion protein or analog etc.) in a vaccine preparation to a human or another mammal. Typically, such vaccine preparations further contain a suitable adjuvant (see, e.g., U.S. Patent No. 6,146,635) or a universal helper epitope such as a PADRETM peptide (Epimmune Inc., San Diego, CA; see, e.g., Alexander et al., J. Immunol. 2000 164(3); 164(3): 1625-1633; Alexander et al., Immunity 1994 1(9): 751-761 and Alexander et al., Immunol. Res. 1998 18(2): 79-92). An alternative method comprises generating an immune response in an individual against a 101P3A11 immunogen by: administering in vivo to muscle or skin of the individual's body a DNA molecule that comprises a DNA sequence that encodes an 101P3A11 immunogen, the DNA sequence operatively linked to regulatory sequences which control the expression of the DNA sequence; wherein the DNA molecule is taken up by

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cells, the DNA sequence is expressed in the cells and an immune response is generated against the immunogen (see, e.g., U.S. Patent No. 5,962,428). Optionally a genetic vaccine facilitator such as anionic lipids; saponins; lectins; estrogenic compounds; hydroxylated lower alkyls; dimethyl sulfoxide; and urea is also administered.

Nucleic Acid Vaccines:

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Vaccine compositions of the invention include nucleic acid-mediated modalities. DNA or RNA that encode protein(s) of the invention can be administered to a patient. Genetic immunization methods can be employed to generate prophylactic or therapeutic humoral and cellular immune responses directed against cancer cells expressing 101P3A11. Constructs comprising DNA encoding a 101P3A11-related protein/immunogen and appropriate regulatory sequences can be injected directly into muscle or skin of an individual, such that the cells of the muscle or skin take-up the construct and express the encoded 101P3A11 protein/immunogen. Alternatively, a vaccine comprises a 101P3A11-related protein. Expression of the 101P3A11-related protein immunogen results in the generation of prophylactic or therapeutic humoral and cellular immunity against cells that bear 101P3A11 protein. Various prophylactic and therapeutic genetic immunization techniques known in the art can be used (for review, see information and references published at Internet address www.genweb.com). Nucleic acid-based delivery is described, for instance, in Wolff et. al., Science 247:1465 (1990) as well as U.S. Patent Nos. 5,580,859; 5,589,466; 5,804,566; 5,739,118; 5,736,524; 5,679,647; WO 98/04720. Examples of DNA-based delivery technologies include "naked DNA", facilitated (bupivicaine, polymers, peptide-mediated) delivery, cationic lipid complexes, and particle-mediated ("gene gun") or pressure-mediated delivery (see, e.g., U.S. Patent Nos. 5,922,687).

For therapeutic or prophylactic immunization purposes, proteins of the invention can be expressed via viral or bacterial vectors. Various viral gene delivery systems that can be used in the practice of the invention include, but are not limited to, vaccinia, fowlpox, canarypox, adenovirus, influenza, poliovirus, adeno-associated virus, lentivirus, and sindbis virus (see, e.g., Restifo, 1996, Curr. Opin. Immunol. 8:658-663; Tsang et al. <u>J. Natl. Cancer Inst.</u> 87:982-990 (1995)). Non-viral delivery systems can also be employed by introducing naked DNA encoding a 101P3A11-related protein into the patient (e.g., intramuscularly or intradermally) to induce an anti-tumor response.

Vaccinia virus is used, for example, as a vector to express nucleotide sequences that encode the peptides of the invention. Upon introduction into a host, the recombinant vaccinia virus expresses the protein immunogenic peptide, and thereby elicits a host immune response. Vaccinia vectors and methods useful in immunization protocols are described in, e.g., U.S. Patent No. 4,722,848. Another vector is BCG (Bacille Calmette Guerin). BCG vectors are described in Stover et al., Nature 351:456-460 (1991). A wide variety of other vectors useful for therapeutic administration or immunization of the peptides of the invention, e.g. adeno and adeno-associated virus vectors, retroviral vectors, Salmonella typhi vectors, detoxified anthrax toxin vectors, and the like, will be apparent to those skilled in the art from the description herein.

Thus, gene delivery systems are used to deliver a 101P3A11-related nucleic acid molecule. In one embodiment, the full-length human 101P3A11 cDNA is employed. In another embodiment, 101P3A11 nucleic acid molecules encoding specific cytotoxic T lymphocyte (CTL) and/or antibody epitopes are employed.

Ex Vivo Vaccines

Various ex vivo strategies can also be employed to generate an immune response. One approach involves the use of antigen presenting cells (APCs) such as dendritic cells (DC) to present 101P3A11 antigen to a patient's immune system.

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Dendritic cells express MHC class I and II molecules, B7 co-stimulator, and IL-12, and are thus highly specialized antigen presenting cells. In prostate cancer, autologous dendritic cells pulsed with peptides of the prostate-specific membrane antigen (PSMA) are being used in a Phase I clinical trial to stimulate prostate cancer patients' immune systems (Tjoa et al., 1996, Prostate 28:65-69; Murphy et al., 1996, Prostate 29:371-380). Thus, dendritic cells can be used to present 101P3A11 peptides to T cells in the context of MHC class I or II molecules. In one embodiment, autologous dendritic cells are pulsed with 101P3A11 peptides capable of binding to MHC class I and/or class II molecules. In another embodiment, dendritic cells are pulsed with the complete 101P3A11 protein. Yet another embodiment involves engineering the overexpression of the 101P3A11 gene in dendritic cells using various implementing vectors known in the art, such as adenovirus (Arthur et al., 1997, Cancer Gene Ther. 4:17-25), retrovirus (Henderson et al., 1996, Cancer Res. 56:3763-3770), lentivirus, adeno-associated virus, DNA transfection (Ribas et al., 1997, Cancer Res. 57:2865-2869), or tumor-derived RNA transfection (Ashley et al., 1997, J. Exp. Med. 186:1177-1182). Cells that express 101P3A11 can also be engineered to express immune modulators, such as GM-CSF, and used as immunizing agents.

X.B.) 101P3A11 as a Target for Antibody-based Therapy

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101P3A11 is an attractive target for antibody-based therapeutic strategies. A number of antibody strategies are known in the art for targeting both extracellular and intracellular molecules (e.g., complement and ADCC mediated killing, the use of intrabodies, and conjugated antibodies) 101P3A11 is expressed by cancer cells of various lineages relative to corresponding normal cells, systemic administration of 101P3A11-immunoreactive compositions are prepared that exhibit excellent sensitivity without toxic, non-specific and/or non-target effects caused by binding of the immunoreactive composition to non-target organs and tissues. Antibodies specifically reactive with domains of 101P3A11 are useful to treat 101P3A11-expressing cancers systemically, either as conjugates with a toxin or therapeutic agent, or as naked antibodies capable of inhibiting cell proliferation or function.

101P3A11 antibodies can be introduced into a patient such that the antibody binds to 101P3A11 and modulates a function, such as an interaction with a binding partner, and consequently mediates destruction of the tumor cells and/or inhibits the growth of the tumor cells. Mechanisms by which such antibodies exert a therapeutic effect can include complement-mediated cytolysis, antibody-dependent cellular cytotoxicity, modulation of the physiological function of 101P3A11, inhibition of ligand binding or signal transduction pathways, modulation of tumor cell differentiation, alteration of tumor angiogenesis factor profiles, and/or apoptosis.

Those skilled in the art understand that antibodies can be used to specifically target and bind immunogenic molecules such as an immunogenic region of the 101P3A11 sequence shown in Figure 2 or Figure 3. In addition, skilled artisans understand that it is routine to conjugate antibodies to cytotoxic agents (see, e.g., Slevers et al. <u>Blood</u> 93:11 3678-3684 (June 1, 1999)). When cytotoxic and/or therapeutic agents are delivered directly to cells, such as by conjugating them to antibodies specific for a molecule expressed by that cell (e.g. 101P3A11), the cytotoxic agent will exert its known biological effect (i.e. cytotoxicity) on those cells.

A wide variety of compositions and methods for using antibody-cytotoxic agent conjugates to kill cells are known in the art. In the context of cancers, typical methods entail administering to an animal having a tumor a biologically effective amount of a conjugate comprising a selected cytotoxic and/or therapeutic agent linked to a targeting agent (e.g. an anti-101P3A11 antibody) that binds to a marker (e.g. 101P3A11) expressed, accessible to

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binding or localized on the cell surfaces. A typical embodiment is a method of delivering a cytotoxic and/or therapeutic agent to a cell expressing 101P3A11, comprising conjugating the cytotoxic agent to an antibody that immunospecifically binds to a 101P3A11 epitope, and, exposing the cell to the antibody-agent conjugate. Another illustrative embodiment is a method of treating an individual suspected of suffering from metastasized cancer, comprising a step of administering parenterally to said individual a pharmaceutical composition comprising a therapeutically effective amount of an antibody conjugated to a cytotoxic and/or therapeutic agent.

Cancer immunotherapy using anti-101P3A11 antibodies can be done in accordance with various approaches that have been successfully employed in the treatment of other types of cancer, including but not limited to colon cancer (Arlen et al., 1998, Crit. Rev. Immunol. 18:133-138), multiple myeloma (Ozaki et al., 1997, Blood 90:3179-3186, Tsunenari et al., 1997, Blood 90:2437-2444), gastric cancer (Kasprzyk et al., 1992, Cancer Res. 52:2771-2776), B-cell lymphoma (Funakoshi et al., 1996, J. Immunother. Emphasis Tumor Immunol. 19:93-101), leukemia (Zhong et al., 1996, Leuk. Res. 20:581-589), colorectal cancer (Moun et al., 1994, Cancer Res. 54:6160-6166; Velders et al., 1995, Cancer Res. 55:4398-4403), and breast cancer (Shepard et al., 1991, J. Clin. Immunol. 11:117-127). Some therapeutic approaches involve conjugation of naked antibody to a toxin or radioisotope, such as the conjugation of Y⁹¹ or I¹³¹ to anti-CD20 antibodies (e.g., ZevalinTM, IDEC Pharmaceuticals Corp. or BexxarTM, Coulter Pharmaceuticals), while others involve co-administration of antibodies and other therapeutic agents, such as HerceptinTM (trastuzumab) with paclitaxel (Genentech, Inc.). To treat prostate cancer, for example, 101P3A11 antibodies can be administered in conjunction with radiation, chemotherapy or hormone ablation.

Although 101P3A11 antibody therapy is useful for all stages of cancer, antibody therapy can be particularly appropriate in advanced or metastatic cancers. Treatment with the antibody therapy of the invention is indicated for patients who have received one or more rounds of chemotherapy. Alternatively, antibody therapy of the invention is combined with a chemotherapeutic or radiation regimen for patients who have not received chemotherapeutic treatment. Additionally, antibody therapy can enable the use of reduced dosages of concomitant chemotherapy, particularly for patients who do not tolerate the toxicity of the chemotherapeutic agent very well. Fan et al. (Cancer Res. 53:4637-4642, 1993), Prewett et al. (International J. of Onco. 9:217-224, 1996), and Hancock et al. (Cancer Res. 51:4575-4580, 1991) describe the use of various antibodies and chemotherapeutic agents *in vitro*.

Cancer patients can be evaluated for the presence and level of 101P3A11 expression, preferably using immunohistochemical assessments of tumor tissue, quantitative 101P3A11 imaging, or other techniques that reliably indicate the presence and degree of 101P3A11 expression. Immunohistochemical analysis of tumor biopsies or surgical specimens is preferred for this purpose. Methods for immunohistochemical analysis of tumor tissues are well known in the art.

Anti-101P3A11 monoclonal antibodies that treat prostate and other cancers include those that initiate a potent immune response against the tumor or those that are directly cytotoxic. In this regard, anti-101P3A11 monoclonal antibodies (mAbs) can elicit tumor cell lysis by either complement-mediated or antibody-dependent cell cytotoxicity (ADCC) mechanisms, both of which require an intact Fc portion of the immunoglobulin molecule for interaction with effector cell Fc receptor sites on complement proteins. In addition, anti-101P3A11 mAbs that exert a direct biological effect on tumor growth are useful to treat cancers that express 101P3A11. Mechanisms by which directly cytotoxic mAbs act include: inhibition of cell growth, modulation of cellular differentiation, modulation of tumor angiogenesis

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factor profiles, and the induction of apoptosis. The mechanism(s) by which a particular anti-101P3A11 mAb exerts an anti-tumor effect is evaluated using any number of *in vitro* assays that evaluate cell death such as ADCC, ADMMC, complement-mediated cell lysis, and so forth, as is generally known in the art.

In some patients, the use of murine or other non-human monoclonal antibodies, or human/mouse chimeric mAbs can induce moderate to strong immune responses against the non-human antibody. This can result in clearance of the antibody from circulation and reduced efficacy. In the most severe cases, such an immune response can lead to the extensive formation of immune complexes which, potentially, can cause renal failure. Accordingly, preferred monoclonal antibodies used in the therapeutic methods of the invention are those that are either fully human or humanized and that bind specifically to the target 101P3A11 antigen with high affinity but exhibit low or no antigenicity in the patient.

Therapeutic methods of the invention contemplate the administration of single anti-101P3A11 mAbs as well as combinations, or cocktails, of different mAbs. Such mAb cocktails can have certain advantages inasmuch as they contain mAbs that target different epitopes, exploit different effector mechanisms or combine directly cytotoxic mAbs with mAbs that rely on immune effector functionality. Such mAbs in combination can exhibit synergistic therapeutic effects. In addition, anti-101P3A11 mAbs can be administered concomitantly with other therapeutic modalities, including but not limited to various chemotherapeutic agents, androgen-blockers, immune modulators (e.g., IL-2, GM-CSF), surgery or radiation. The anti-101P3A11 mAbs are administered in their "naked" or unconjugated form, or can have a therapeutic agent(s) conjugated to them.

Anti-101P3A11 antibody formulations are administered via any route capable of delivering the antibodies to a tumor cell. Routes of administration include, but are not limited to, intravenous, intraperitoneal, intramuscular, intratumor, intradermal, and the like. Treatment generally involves repeated administration of the anti-101P3A11 antibody preparation, via an acceptable route of administration such as intravenous injection (IV), typically at a dose in the range of about 0.1, .2, .3, .4, .5, .6, .7, .8, .9., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, or 25 mg/kg body weight. In general, doses in the range of 10-1000 mg mAb per week are effective and well tolerated.

Based on clinical experience with the Herceptin mAb in the treatment of metastatic breast cancer, an initial loading dose of approximately 4 mg/kg patient body weight IV, followed by weekly doses of about 2 mg/kg IV of the anti- 101P3A11 mAb preparation represents an acceptable dosing regimen. Preferably, the initial loading dose is administered as a 90 minute or longer infusion. The periodic maintenance dose is administered as a 30 minute or longer infusion, provided the initial dose was well tolerated. As appreciated by those of skill in the art, various factors can influence the ideal dose regimen in a particular case. Such factors include, for example, the binding affinity and half life of the Ab or mAbs used, the degree of 101P3A11 expression in the patient, the extent of circulating shed 101P3A11 antigen, the desired steady-state antibody concentration level, frequency of treatment, and the influence of chemotherapeutic or other agents used in combination with the treatment method of the invention, as well as the health status of a particular patient.

Optionally, patients should be evaluated for the levels of 101P3A11 in a given sample (e.g. the levels of circulating 101P3A11 antigen and/or 101P3A11 expressing cells) in order to assist in the determination of the most effective dosing regimen, etc. Such evaluations are also used for monitoring purposes throughout therapy, and are

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useful to gauge therapeutic success in combination with the evaluation of other parameters (for example, urine cytology and/or ImmunoCyt levels in bladder cancer therapy, or by analogy, serum PSA levels in prostate cancer therapy).

Anti-idiotypic anti-101P3A11 antibodies can also be used in anti-cancer therapy as a vaccine for inducing an immune response to cells expressing a 101P3A11-related protein. In particular, the generation of anti-idiotypic antibodies is well known in the art; this methodology can readily be adapted to generate anti-idiotypic anti-101P3A11 antibodies that mimic an epitope on a 101P3A11-related protein (see, for example, Wagner et al., 1997, Hybridoma 16: 33-40; Foon et al., 1995, J. Clin. Invest. 96:334-342; Herlyn et al., 1996, Cancer Immunol. Immunother. 43:65-76). Such an anti-idiotypic antibody can be used in cancer vaccine strategies.

X.C.) 101P3A11 as a Target for Cellular Immune Responses

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Vaccines and methods of preparing vaccines that contain an immunogenically effective amount of one or more HLA-binding peptides as described herein are further embodiments of the invention. Furthermore, vaccines in accordance with the invention encompass compositions of one or more of the claimed peptides. A peptide can be present in a vaccine individually. Alternatively, the peptide can exist as a homopolymer comprising multiple copies of the same peptide, or as a heteropolymer of various peptides. Polymers have the advantage of increased immunological reaction and, where different peptide epitopes are used to make up the polymer, the additional ability to induce antibodies and/or CTLs that react with different antigenic determinants of the pathogenic organism or tumor-related peptide targeted for an immune response. The composition can be a naturally occurring region of an antigen or can be prepared, e.g., recombinantly or by chemical synthesis.

Carriers that can be used with vaccines of the invention are well known in the art, and include, *e.g.*, thyroglobulin, albumins such as human serum albumin, tetanus toxoid, polyamino acids such as poly L-lysine, poly L-glutamic acid, influenza, hepatitis B virus core protein, and the like. The vaccines can contain a physiologically tolerable (*i.e.*, acceptable) diluent such as water, or saline, preferably phosphate buffered saline. The vaccines also typically include an adjuvant. Adjuvants such as incomplete Freund's adjuvant, aluminum phosphate, aluminum hydroxide, or alum are examples of materials well known in the art. Additionally, as disclosed herein, CTL responses can be primed by conjugating peptides of the invention to lipids, such as tripalmitoyl-S-glycerylcysteinlyseryl- serine (P₃CSS). Moreover, an adjuvant such as a synthetic cytosine-phosphorothiolated-guanine-containing (CpG) oligonucleotides has been found to increase CTL responses 10- to 100-fold. (see, e.g. Davila and Celis J. Immunol. 165:539-547 (2000))

Upon immunization with a peptide composition in accordance with the invention, via injection, aerosol, oral, transdermal, transmucosal, intrapleural, intrathecal, or other suitable routes, the immune system of the host responds to the vaccine by producing large amounts of CTLs and/or HTLs specific for the desired antigen. Consequently, the host becomes at least partially immune to later development of cells that express or overexpress 101P3A11 antigen, or derives at least some therapeutic benefit when the antigen was tumor-associated.

In some embodiments, it may be desirable to combine the class I peptide components with components that induce or facilitate neutralizing antibody and or helper T cell responses directed to the target antigen. A preferred embodiment of such a composition comprises class I and class II epitopes in accordance with the invention. An alternative embodiment of such a composition comprises a class I and/or class II epitope in accordance with the

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invention, along with a cross reactive HTL epitope such as PADRE™ (Epimmune, San Diego, CA) molecule (described e.g., in U.S. Patent Number 5,736,142).

A vaccine of the invention can also include antigen-presenting cells (APC), such as dendritic cells (DC), as a vehicle to present peptides of the invention. Vaccine compositions can be created *in vitro*, following dendritic cell mobilization and harvesting, whereby loading of dendritic cells occurs *in vitro*. For example, dendritic cells are transfected, *e.g.*, with a minigene in accordance with the invention, or are pulsed with peptides. The dendritic cell can then be administered to a patient to elicit immune responses *in vivo*. Vaccine compositions, either DNA- or peptidebased, can also be administered *in vivo* in combination with dendritic cell mobilization whereby loading of dendritic cells occurs *in vivo*.

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Preferably, the following principles are utilized when selecting an array of epitopes for inclusion in a polyepitopic composition for use in a vaccine, or for selecting discrete epitopes to be included in a vaccine and/or to be encoded by nucleic acids such as a minigene. It is preferred that each of the following principles be balanced in order to make the selection. The multiple epitopes to be incorporated in a given vaccine composition may be, but need not be, contiguous in sequence in the native antigen from which the epitopes are derived.

- 1.) Epitopes are selected which, upon administration, mimic immune responses that have been observed to be correlated with tumor clearance. For HLA Class I this includes 3-4 epitopes that come from at least one tumor associated antigen (TAA). For HLA Class II a similar rationale is employed; again 3-4 epitopes are selected from at least one TAA (see, e.g., Rosenberg et al., Science 278:1447-1450). Epitopes from one TAA may be used in combination with epitopes from one or more additional TAAs to produce a vaccine that targets tumors with varying expression patterns of frequently-expressed TAAs.
- 2.) Epitopes are selected that have the requisite binding affinity established to be correlated with immunogenicity: for HLA Class I an IC₅₀ of 500 nM or less, often 200 nM or less; and for Class II an IC₅₀ of 1000 nM or less.
- 3.) Sufficient supermotif bearing-peptides, or a sufficient array of allele-specific motif-bearing peptides, are selected to give broad population coverage. For example, it is preferable to have at least 80% population coverage. A Monte Carlo analysis, a statistical evaluation known in the art, can be employed to assess the breadth, or redundancy of, population coverage.
- 4.) When selecting epitopes from cancer-related antigens it is often useful to select analogs because the patient may have developed tolerance to the native epitope.
- 5.) Of particular relevance are epitopes referred to as "nested epitopes." Nested epitopes occur where at least two epitopes overlap in a given peptide sequence. A nested peptide sequence can comprise B cell, HLA class I and/or HLA class II epitopes. When providing nested epitopes, a general objective is to provide the greatest number of epitopes per sequence. Thus, an aspect is to avoid providing a peptide that is any longer than the amino terminus of the amino terminal epitope and the carboxyl terminus of the carboxyl terminal epitope in the peptide. When providing a multi-epitopic sequence, such as a sequence comprising nested epitopes, it is generally important to screen the sequence in order to insure that it does not have pathological or other deleterious biological properties.
- 6.) If a polyepitopic protein is created, or when creating a minigene, an objective is to generate the smallest peptide that encompasses the epitopes of interest. This principle is similar, if not the same as that employed

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when selecting a peptide comprising nested epitopes. However, with an artificial polyepitopic peptide, the size minimization objective is balanced against the need to integrate any spacer sequences between epitopes in the polyepitopic protein. Spacer amino acid residues can, for example, be introduced to avoid junctional epitopes (an epitope recognized by the immune system, not present in the target antigen, and only created by the man-made juxtaposition of epitopes), or to facilitate cleavage between epitopes and thereby enhance epitope presentation. Junctional epitopes are generally to be avoided because the recipient may generate an immune response to that non-native epitope. Of particular concern is a junctional epitope that is a "dominant epitope." A dominant epitope may lead to such a zealous response that immune responses to other epitopes are diminished or suppressed.

7.) Where the sequences of multiple variants of the same target protein are present, potential peptide epitopes can also be selected on the basis of their conservancy. For example, a criterion for conservancy may define that the entire sequence of an HLA class I binding peptide or the entire 9-mer core of a class II binding peptide be conserved in a designated percentage of the sequences evaluated for a specific protein antigen.

X.C.1. Minigene Vaccines

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A number of different approaches are available which allow simultaneous delivery of multiple epitopes.

Nucleic acids encoding the peptides of the invention are a particularly useful embodiment of the invention. Epitopes for inclusion in a minigene are preferably selected according to the guidelines set forth in the previous section. A preferred means of administering nucleic acids encoding the peptides of the invention uses minigene constructs encoding a peptide comprising one or multiple epitopes of the invention.

The use of multi-epitope minigenes is described below and in, Ishioka et al., J. Immunol. 162:3915-3925, 1999; An, L. and Whitton, J. L., J. Virol. 71:2292, 1997; Thomson, S. A. et al., J. Immunol. 157:822, 1996; Whitton, J. L. et al., J. Virol. 67:348, 1993; Hanke, R. et al., Vaccine 16:426, 1998. For example, a multi-epitope DNA plasmid encoding supermotif- and/or motif-bearing epitopes derived 101P3A11, the PADRE® universal helper T cell epitope (or multiple HTL epitopes from 101P3A11), and an endoplasmic reticulum-translocating signal sequence can be engineered. A vaccine may also comprise epitopes that are derived from other TAAs.

The immunogenicity of a multi-epitopic minigene can be confirmed in transgenic mice to evaluate the magnitude of CTL induction responses against the epitopes tested. Further, the immunogenicity of DNA-encoded epitopes *in vivo* can be correlated with the *in vitro* responses of specific CTL lines against target cells transfected with the DNA plasmid. Thus, these experiments can show that the minigene serves to both: 1.) generate a CTL response and 2.) that the induced CTLs recognized cells expressing the encoded epitopes.

For example, to create a DNA sequence encoding the selected epitopes (minigene) for expression in human cells, the amino acid sequences of the epitopes may be reverse translated. A human codon usage table can be used to guide the codon choice for each amino acid. These epitope-encoding DNA sequences may be directly adjoined, so that when translated, a continuous polypeptide sequence is created. To optimize expression and/or immunogenicity, additional elements can be incorporated into the minigene design. Examples of amino acid sequences that can be reverse translated and included in the minigene sequence include: HLA class I epitopes, HLA class II epitopes, antibody epitopes, a ubiquitination signal sequence, and/or an endoplasmic reticulum targeting signal. In addition, HLA presentation of CTL and HTL epitopes may be improved by including synthetic (e.g. poly-alanine) or naturally-

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occurring flanking sequences adjacent to the CTL or HTL epitopes; these larger peptides comprising the epitope(s) are within the scope of the invention.

The minigene sequence may be converted to DNA by assembling oligonucleotides that encode the plus and minus strands of the minigene. Overlapping oligonucleotides (30-100 bases long) may be synthesized, phosphorylated, purified and annealed under appropriate conditions using well known techniques. The ends of the oligonucleotides can be joined, for example, using T4 DNA ligase. This synthetic minigene, encoding the epitope polypeptide, can then be cloned into a desired expression vector.

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Standard regulatory sequences well known to those of skill in the art are preferably included in the vector to ensure expression in the target cells. Several vector elements are desirable: a promoter with a down-stream cloning site for minigene insertion; a polyadenylation signal for efficient transcription termination; an *E. coli* origin of replication; and an *E. coli* selectable marker (*e.g.* ampicillin or kanamycin resistance). Numerous promoters can be used for this purpose, *e.g.*, the human cytomegalovirus (hCMV) promoter. See, *e.g.*, U.S. Patent Nos. 5,580,859 and 5,589,466 for other suitable promoter sequences.

Additional vector modifications may be desired to optimize minigene expression and immunogenicity. In some cases, introns are required for efficient gene expression, and one or more synthetic or naturally-occurring introns could be incorporated into the transcribed region of the minigene. The inclusion of mRNA stabilization sequences and sequences for replication in mammalian cells may also be considered for increasing minigene expression.

Once an expression vector is selected, the minigene is cloned into the polylinker region downstream of the promoter. This plasmid is transformed into an appropriate *E. coli* strain, and DNA is prepared using standard techniques. The orientation and DNA sequence of the minigene, as well as all other elements included in the vector, are confirmed using restriction mapping and DNA sequence analysis. Bacterial cells harboring the correct plasmid can be stored as a master cell bank and a working cell bank.

In addition, immunostimulatory sequences (ISSs or CpGs) appear to play a role in the immunogenicity of DNA vaccines. These sequences may be included in the vector, outside the minigene coding sequence, if desired to enhance immunogenicity.

In some embodiments, a bi-cistronic expression vector which allows production of both the minigene-encoded epitopes and a second protein (included to enhance or decrease immunogenicity) can be used. Examples of proteins or polypeptides that could beneficially enhance the immune response if co-expressed include cytokines (e.g., IL-2, IL-12, GM-CSF), cytokine-inducing molecules (e.g., LeIF), costimulatory molecules, or for HTL responses, pan-DR binding proteins (PADRE™, Epimmune, San Diego, CA). Helper (HTL) epitopes can be joined to intracellular targeting signals and expressed separately from expressed CTL epitopes; this allows direction of the HTL epitopes to a cell compartment different than that of the CTL epitopes. If required, this could facilitate more efficient entry of HTL epitopes into the HLA class II pathway, thereby improving HTL induction. In contrast to HTL or CTL induction, specifically decreasing the immune response by co-expression of immunosuppressive molecules (e.g. TGF-β) may be beneficial in certain diseases.

Therapeutic quantities of plasmid DNA can be produced for example, by fermentation in *E. coli*, followed by purification. Aliquots from the working cell bank are used to inoculate growth medium, and grown to saturation in shaker flasks or a bioreactor according to well-known techniques. Plasmid DNA can be purified using standard

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bioseparation technologies such as solid phase anion-exchange resins supplied by QIAGEN, Inc. (Valencia, California). If required, supercoiled DNA can be isolated from the open circular and linear forms using gel electrophoresis or other methods.

Purified plasmid DNA can be prepared for injection using a variety of formulations. The simplest of these is reconstitution of lyophilized DNA in sterile phosphate-buffer saline (PBS). This approach, known as "naked DNA," is currently being used for intramuscular (IM) administration in clinical trials. To maximize the immunotherapeutic effects of minigene DNA vaccines, an alternative method for formulating purified plasmid DNA may be desirable. A variety of methods have been described, and new techniques may become available. Cationic lipids, glycolipids, and fusogenic liposomes can also be used in the formulation (see, e.g., as described by WO 93/24640; Mannino & Gould-Fogerite, *BioTechniques* 6(7): 682 (1988); U.S. Pat No. 5,279,833; WO 91/06309; and Felgner, et al., *Proc. Nat'l Acad. Sci. USA* 84:7413 (1987). In addition, peptides and compounds referred to collectively as protective, interactive, noncondensing compounds (PINC) could also be complexed to purified plasmid DNA to influence variables such as stability, intramuscular dispersion, or trafficking to specific organs or cell types.

Target cell sensitization can be used as a functional assay for expression and HLA class I presentation of minigene-encoded CTL epitopes. For example, the plasmid DNA is introduced into a mammalian cell line that is suitable as a target for standard CTL chromium release assays. The transfection method used will be dependent on the final formulation. Electroporation can be used for "naked" DNA, whereas cationic lipids allow direct *in vitro* transfection. A plasmid expressing green fluorescent protein (GFP) can be co-transfected to allow enrichment of transfected cells using fluorescence activated cell sorting (FACS). These cells are then chromium-51 (⁵¹Cr) labeled and used as target cells for epitope-specific CTL lines; cytolysis, detected by ⁵¹Cr release, indicates both production of, and HLA presentation of, minigene-encoded CTL epitopes. Expression of HTL epitopes may be evaluated in an analogous manner using assays to assess HTL activity.

In vivo immunogenicity is a second approach for functional testing of minigene DNA formulations. Transgenic mice expressing appropriate human HLA proteins are immunized with the DNA product. The dose and route of administration are formulation dependent (e.g., IM for DNA in PBS, intraperitoneal (i.p.) for lipid-complexed DNA). Twenty-one days after immunization, splenocytes are harvested and restimulated for one week in the presence of peptides encoding each epitope being tested. Thereafter, for CTL effector cells, assays are conducted for cytolysis of peptide-loaded, ⁵¹Cr-labeled target cells using standard techniques. Lysis of target cells that were sensitized by HLA loaded with peptide epitopes, corresponding to minigene-encoded epitopes, demonstrates DNA vaccine function for in vivo induction of CTLs. Immunogenicity of HTL epitopes is confirmed in transgenic mice in an analogous manner.

Alternatively, the nucleic acids can be administered using ballistic delivery as described, for instance, in U.S. Patent No. 5,204,253. Using this technique, particles comprised solely of DNA are administered. In a further alternative embodiment, DNA can be adhered to particles, such as gold particles.

Minigenes can also be delivered using other bacterial or viral delivery systems well known in the art, e.g., an expression construct encoding epitopes of the invention can be incorporated into a viral vector such as vaccinia.

X.C.2. Combinations of CTL Peptides with Helper Peptides

Vaccine compositions comprising CTL peptides of the invention can be modified, e.g., analoged, to provide desired attributes, such as improved serum half life, broadened population coverage or enhanced immunogenicity.

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For instance, the ability of a peptide to induce CTL activity can be enhanced by linking the peptide to a sequence which contains at least one epitope that is capable of inducing a T helper cell response. Although a CTL peptide can be directly linked to a T helper peptide, often CTL epitope/HTL epitope conjugates are linked by a spacer molecule. The spacer is typically comprised of relatively small, neutral molecules, such as amino acids or amino acid mimetics, which are substantially uncharged under physiological conditions. The spacers are typically selected from, e.g., Ala, Gly, or other neutral spacers of nonpolar amino acids or neutral polar amino acids. It will be understood that the optionally present spacer need not be comprised of the same residues and thus may be a hetero- or homo-oligomer. When present, the spacer will usually be at least one or two residues, more usually three to six residues and sometimes 10 or more residues. The CTL peptide epitope can be linked to the T helper peptide epitope either directly or via a spacer either at the amino or carboxy terminus of the CTL peptide. The amino terminus of either the immunogenic peptide or the T helper peptide may be acylated.

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In certain embodiments, the T helper peptide is one that is recognized by T helper cells present in a majority of a genetically diverse population. This can be accomplished by selecting peptides that bind to many, most, or all of the HLA class II molecules. Examples of such amino acid bind many HLA Class II molecules include sequences from antigens such as tetanus toxoid at positions 830-843 (QYIKANSKFIGITE; SEQ ID NO: 7102876), Plasmodium falciparum circumsporozoite (CS) protein at positions 378-398 (DIEKKIAKMEKASSVFNVVNS; SEQ ID NO: 7112877), and Streptococcus 18kD protein at positions 116-131 (GAVDSILGGVATYGAA; SEQ ID NO: 7122878). Other examples include peptides bearing a DR 1-4-7 supermotif, or either of the DR3 motifs.

Alternatively, it is possible to prepare synthetic peptides capable of stimulating T helper lymphocytes, in a loosely HLA-restricted fashion, using amino acid sequences not found in nature (see, e.g., PCT publication WO 95/07707). These synthetic compounds called Pan-DR-binding epitopes (e.g., PADRETM, Epimmune, Inc., San Diego, CA) are designed to most preferably bind most HLA-DR (human HLA class II) molecules. For instance, a pan-DR-binding epitope peptide having the formula: aKXVAAWTLKAAa (SEQ ID NO: 7132879), where "X" is either cyclohexylalanine, phenylalanine, or tyrosine, and a is either D-alanine or L-alanine, has been found to bind to most HLA-DR alleles, and to stimulate the response of T helper lymphocytes from most individuals, regardless of their HLA type. An alternative of a pan-DR binding epitope comprises all "L" natural amino acids and can be provided in the form of nucleic acids that encode the epitope.

HTL peptide epitopes can also be modified to alter their biological properties. For example, they can be modified to include D-amino acids to increase their resistance to proteases and thus extend their serum half life, or they can be conjugated to other molecules such as lipids, proteins, carbohydrates, and the like to increase their biological activity. For example, a T helper peptide can be conjugated to one or more palmitic acid chains at either the amino or carboxyl termini.

X.C.3. Combinations of CTL Peptides with T Cell Priming Agents

In some embodiments it may be desirable to include in the pharmaceutical compositions of the invention at least one component which primes B lymphocytes or T lymphocytes. Lipids have been identified as agents capable of priming CTL *in vivo*. For example, palmitic acid residues can be attached to the ε -and α - amino groups of a lysine residue and then linked, *e.g.*, via one or more linking residues such as Gly, Gly-Gly-, Ser, Ser-Ser, or the like, to an immunogenic peptide. The lipidated peptide can then be administered either directly in a micelle or particle,

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incorporated into a liposome, or emulsified in an adjuvant, e.g., incomplete Freund's adjuvant. In a preferred embodiment, a particularly effective immunogenic composition comprises palmitic acid attached to ε - and α - amino groups of Lys, which is attached via linkage, e.g., Ser-Ser, to the amino terminus of the immunogenic peptide.

As another example of lipid priming of CTL responses, *E. coli* lipoproteins, such as tripalmitoyl-S-glycerylcysteinlyseryl- serine (P₃CSS) can be used to prime virus specific CTL when covalently attached to an appropriate peptide (*see*, *e.g.*, Deres, *et al.*, *Nature* 342:561, 1989). Peptides of the invention can be coupled to P₃CSS, for example, and the lipopeptide administered to an individual to specifically prime an immune response to the target antigen. Moreover, because the induction of neutralizing antibodies can also be primed with P₃CSS-conjugated epitopes, two such compositions can be combined to more effectively elicit both humoral and cell-mediated responses.

X.C.4. Vaccine Compositions Comprising DC Pulsed with CTL and/or HTL Peptides

An embodiment of a vaccine composition in accordance with the invention comprises *ex vivo* administration of a cocktail of epitope-bearing peptides to PBMC, or isolated DC therefrom, from the patient's blood. A pharmaceutical to facilitate harvesting of DC can be used, such as ProgenipoietinTM (Pharmacia-Monsanto, St. Louis, MO) or GM-CSF/IL-4. After pulsing the DC with peptides and prior to reinfusion into patients, the DC are washed to remove unbound peptides. In this embodiment, a vaccine comprises peptide-pulsed DCs which present the pulsed peptide epitopes complexed with HLA molecules on their surfaces.

The DC can be pulsed *ex vivo* with a cocktail of peptides, some of which stimulate CTL responses to 101P3A11. Optionally, a helper T cell (HTL) peptide, such as a natural or artificial loosely restricted HLA Class II peptide, can be included to facilitate the CTL response. Thus, a vaccine in accordance with the invention is used to treat a cancer which expresses or overexpresses 101P3A11.

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Antigenic 101P3A11-related peptides are used to elicit a CTL and/or HTL response ex vivo, as well. The resulting CTL or HTL cells, can be used to treat tumors in patients that do not respond to other conventional forms of therapy, or will not respond to a therapeutic vaccine peptide or nucleic acid in accordance with the invention. Ex vivo CTL or HTL responses to a particular antigen are induced by incubating in tissue culture the patient's, or genetically compatible, CTL or HTL precursor cells together with a source of antigen-presenting cells (APC), such as dendritic cells, and the appropriate immunogenic peptide. After an appropriate incubation time (typically about 7-28 days), in which the precursor cells are activated and expanded into effector cells, the cells are infused back into the patient, where they will destroy (CTL) or facilitate destruction (HTL) of their specific target cell (e.g., a tumor cell). Transfected dendritic cells may also be used as antigen presenting cells.

X.E. Administration of Vaccines for Therapeutic or Prophylactic Purposes

Pharmaceutical and vaccine compositions of the invention are typically used to treat and/or prevent a cancer that expresses or overexpresses 101P3A11. In therapeutic applications, peptide and/or nucleic acid compositions are administered to a patient in an amount sufficient to elicit an effective B cell, CTL and/or HTL response to the antigen and to cure or at least partially arrest or slow symptoms and/or complications. An amount adequate to accomplish this is defined as "therapeutically effective dose." Amounts effective for this use will depend on, e.g., the particular composition administered, the manner of administration, the stage and severity of the disease being treated, the weight and general state of health of the patient, and the judgment of the prescribing physician.

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For pharmaceutical compositions, the immunogenic peptides of the invention, or DNA encoding them, are generally administered to an individual already bearing a tumor that expresses 101P3A11. The peptides or DNA encoding them can be administered individually or as fusions of one or more peptide sequences. Patients can be treated with the immunogenic peptides separately or in conjunction with other treatments, such as surgery, as appropriate.

For therapeutic use, administration should generally begin at the first diagnosis of 101P3A11-associated cancer. This is followed by boosting doses until at least symptoms are substantially abated and for a period thereafter. The embodiment of the vaccine composition (*i.e.*, including, but not limited to embodiments such as peptide cocktails, polyepitopic polypeptides, minigenes, or TAA-specific CTLs or pulsed dendritic cells) delivered to the patient may vary according to the stage of the disease or the patient's health status. For example, in a patient with a tumor that expresses 101P3A11, a vaccine comprising 101P3A11-specific CTL may be more efficacious in killing tumor cells in patient with advanced disease than alternative embodiments.

It is generally important to provide an amount of the peptide epitope delivered by a mode of administration sufficient to effectively stimulate a cytotoxic T cell response; compositions which stimulate helper T cell responses can also be given in accordance with this embodiment of the invention.

The dosage for an initial therapeutic immunization generally occurs in a unit dosage range where the lower value is about 1, 5, 50, 500, or 1,000 µg and the higher value is about 10,000; 20,000; 30,000; or 50,000 µg. Dosage values for a human typically range from about 500 µg to about 50,000 µg per 70 kilogram patient. Boosting dosages of between about 1.0 µg to about 50,000 µg of peptide pursuant to a boosting regimen over weeks to months may be administered depending upon the patient's response and condition as determined by measuring the specific activity of CTL and HTL obtained from the patient's blood. Administration should continue until at least clinical symptoms or laboratory tests indicate that the neoplasia, has been eliminated or reduced and for a period thereafter. The dosages, routes of administration, and dose schedules are adjusted in accordance with methodologies known in the art.

In certain embodiments, the peptides and compositions of the present invention are employed in serious disease states, that is, life-threatening or potentially life threatening situations. In such cases, as a result of the minimal amounts of extraneous substances and the relative nontoxic nature of the peptides in preferred compositions of the invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these peptide compositions relative to these stated dosage amounts.

The vaccine compositions of the invention can also be used purely as prophylactic agents. Generally the dosage for an initial prophylactic immunization generally occurs in a unit dosage range where the lower value is about $1, 5, 50, 500, \text{ or } 1000 \text{ }\mu\text{g}$ and the higher value is about $10,000; 20,000; 30,000; \text{ or } 50,000 \text{ }\mu\text{g}$. Dosage values for a human typically range from about $500 \text{ }\mu\text{g}$ to about $50,000 \text{ }\mu\text{g}$ per 70 kilogram patient. This is followed by boosting dosages of between about $1.0 \text{ }\mu\text{g}$ to about $50,000 \text{ }\mu\text{g}$ of peptide administered at defined intervals from about four weeks to six months after the initial administration of vaccine. The immunogenicity of the vaccine can be assessed by measuring the specific activity of CTL and HTL obtained from a sample of the patient's blood.

The pharmaceutical compositions for therapeutic treatment are intended for parenteral, topical, oral, nasal, intrathecal, or local (e.g. as a cream or topical ointment) administration. Preferably, the pharmaceutical compositions are administered parentally, e.g., intravenously, subcutaneously, intradermally, or intramuscularly. Thus, the invention

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provides compositions for parenteral administration which comprise a solution of the immunogenic peptides dissolved or suspended in an acceptable carrier, preferably an aqueous carrier.

A variety of aqueous carriers may be used, e.g., water, buffered water, 0.8% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be sterilized by conventional, well-known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile solution prior to administration.

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The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions, such as pH-adjusting and buffering agents, tonicity adjusting agents, wetting agents, preservatives, and the like, for example, sodium acetate, sodium lactate, sodium chloride, potassium chloride, calcium chloride, sorbitan monolaurate, triethanolamine oleate. *etc.*

The concentration of peptides of the invention in the pharmaceutical formulations can vary widely, *i.e.*, from less than about 0.1%, usually at or at least about 2% to as much as 20% to 50% or more by weight, and will be selected primarily by fluid volumes, viscosities, *etc.*, in accordance with the particular mode of administration selected.

A human unit dose form of a composition is typically included in a pharmaceutical composition that comprises a human unit dose of an acceptable carrier, in one embodiment an aqueous carrier, and is administered in a volume/quantity that is known by those of skill in the art to be used for administration of such compositions to humans (*see, e.g.*, Remington's Pharmaceutical Sciences, 17th Edition, A. Gennaro, Editor, Mack Publishing Co., Easton, Pennsylvania, 1985). For example a peptide dose for initial immunization can be from about 1 to about 50,000 μg, generally 100-5,000 μg, for a 70 kg patient. For example, for nucleic acids an initial immunization may be performed using an expression vector in the form of naked nucleic acid administered IM (or SC or ID) in the amounts of 0.5-5 mg at multiple sites. The nucleic acid (0.1 to 1000 μg) can also be administered using a gene gun (i.e., ballistic delivery) in accordance with regimens known in the art. Following an incubation period of 3-4 weeks, a booster dose is then administered. The booster can be recombinant fowlpox virus administered at a dose of 5-10⁷ to 5x10⁹ pfu.

For antibodies, a treatment generally involves repeated administration of the anti-101P3A11 antibody preparation, via an acceptable route of administration such as intravenous injection (IV), typically at a dose in the range of about 0.1 to about 10 mg/kg body weight. In general, doses in the range of 10-500 mg mAb per week are effective and well tolerated. Moreover, an initial loading dose of approximately 4 mg/kg patient body weight IV, followed by weekly doses of about 2 mg/kg IV of the anti- 101P3A11 mAb preparation represents an acceptable dosing regimen. As appreciated by those of skill in the art, various factors can influence the ideal dose in a particular case. Such factors include, for example, half life of a composition, the binding affinity of an Ab, the immunogenicity of a substance, the degree of 101P3A11 expression in the patient, the extent of circulating shed 101P3A11 antigen, the desired steady-state concentration level, frequency of treatment, and the influence of chemotherapeutic or other agents used in combination with the treatment method of the invention, as well as the health status of a particular patient. Non-limiting preferred human unit doses are, for example, 500µg - 1mg, 1mg - 50mg, 50mg-100mg, 100mg - 200 mg, 200mg - 300mg, 400mg - 500mg, 500mg - 600mg, 600mg - 700mg, or 1mg - 700mg. In certain embodiments, the dose is in a range of 2-5 mg/kg body weight, e.g., with follow on weekly doses of 1-3 mg/kg; 0.5mg, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10mg/kg body weight followed, e.g., in two, three or four weeks by weekly doses; 0.5 - 10mg/kg body weight, e.g., followed in two, three or four weeks by weekly doses; 225, 250, 275, 300, 325, 350, 375, 400mg m² of body area weekly; 1-600mg m² of

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body area weekly; 225-400mg m² of body area weekly; these does can be followed by weekly doses for 2, 3, 4, 5, 6, 7, 8, 9, 19, 11, 12 or more weeks.

In one embodiment, human unit dose forms of polynucleotides comprise a suitable dosage range or effective amount that provides any therapeutic effect. As appreciated by one of ordinary skill in the art a therapeutic effect depends on a number of factors, including the sequence of the polynucleotide, molecular weight of the polynucleotide and route of administration. Dosages are generally selected by the physician or other health care professional in accordance with a variety of parameters known in the art, such as severity of symptoms, history of the patient and the like. Generally, for a polynucleotide of about 20 bases, a dosage range may be selected from, for example, an independently selected lower limit such as about 0.1, 0.25, 0.5, 1, 2, 5, 10, 20, 30, 40, 50, 60, 80, 100, 200, 300, 400 or 500 mg/kg up to an independently selected upper limit, greater than the lower limit, of about 60, 80, 100, 200, 300, 400, 500, 750, 1000, 1500, 2000, 3000, 4000, 5000, 6000, 7000, 8000, 9000 or 10,000 mg/kg. For example, a dose may be about any of the following: 0.1 to 100 mg/kg, 0.1 to 50 mg/kg, 0.1 to 25 mg/kg, 0.1 to 10 mg/kg, 1 to 500 mg/kg, 100 to 400 mg/kg, 200 to 300 mg/kg, 1 to 100 mg/kg, 100 to 200 mg/kg, 300 to 400 mg/kg, 400 to 500 mg/kg, 500 to 1000 mg/kg, 500 to 5000 mg/kg, or 500 to 10,000 mg/kg. Generally, parenteral routes of administration may require higher doses of polynucleotide compared to more direct application to the nucleotide to diseased tissue, as do polynucleotides of increasing length.

In one embodiment, human unit dose forms of T-cells comprise a suitable dosage range or effective amount that provides any therapeutic effect. As appreciated by one of ordinary skill in the art, a therapeutic effect depends on a number of factors. Dosages are generally selected by the physician or other health care professional in accordance with a variety of parameters known in the art, such as severity of symptoms, history of the patient and the like. A dose may be about 10^4 cells to about 10^6 cells, about 10^6 cells to about 10^8 to about 10^8 to about 10^{10} cells. A dose may also about 10^6 cells/m² to about 10^{10} cells/m², or about 10^6 cells/m² to about 10^8 cells/m².

Proteins(s) of the invention, and/or nucleic acids encoding the protein(s), can also be administered via liposomes, which may also serve to: 1) target the proteins(s) to a particular tissue, such as lymphoid tissue; 2) to target selectively to diseases cells; or, 3) to increase the half-life of the peptide composition. Liposomes include emulsions, foams, micelles, insoluble monolayers, liquid crystals, phospholipid dispersions, lamellar layers and the like. In these preparations, the peptide to be delivered is incorporated as part of a liposome, alone or in conjunction with a molecule which binds to a receptor prevalent among lymphoid cells, such as monoclonal antibodies which bind to the CD45 antigen, or with other therapeutic or immunogenic compositions. Thus, liposomes either filled or decorated with a desired peptide of the invention can be directed to the site of lymphoid cells, where the liposomes then deliver the peptide compositions. Liposomes for use in accordance with the invention are formed from standard vesicle-forming lipids, which generally include neutral and negatively charged phospholipids and a sterol, such as cholesterol. The selection of lipids is generally guided by consideration of, e.g., liposome size, acid lability and stability of the liposomes in the blood stream. A variety of methods are available for preparing liposomes, as described in, e.g., Szoka, et al., Ann. Rev. Biophys. Bioeng. 9:467 (1980), and U.S. Patent Nos. 4,235,871, 4,501,728, 4,837,028, and 5,019,369.

For targeting cells of the immune system, a ligand to be incorporated into the liposome can include, e.g., antibodies or fragments thereof specific for cell surface determinants of the desired immune system cells. A liposome

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suspension containing a peptide may be administered intravenously, locally, topically, *etc.* in a dose which varies according to, *inter alia*, the manner of administration, the peptide being delivered, and the stage of the disease being treated.

For solid compositions, conventional nontoxic solid carriers may be used which include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talcum, cellulose, glucose, sucrose, magnesium carbonate, and the like. For oral administration, a pharmaceutically acceptable nontoxic composition is formed by incorporating any of the normally employed excipients, such as those carriers previously listed, and generally 10-95% of active ingredient, that is, one or more peptides of the invention, and more preferably at a concentration of 25%-75%.

For aerosol administration, immunogenic peptides are preferably supplied in finely divided form along with a surfactant and propellant. Typical percentages of peptides are about 0.01%-20% by weight, preferably about 1%-10%. The surfactant must, of course, be nontoxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from about 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed. The surfactant may constitute about 0.1%-20% by weight of the composition, preferably about 0.25-5%. The balance of the composition is ordinarily propellant. A carrier can also be included, as desired, as with, e.g., lecithin for intranasal delivery.

XI.) Diagnostic and Prognostic Embodiments of 101P3A11.

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As disclosed herein, 101P3A11 polynucleotides, polypeptides, reactive cytotoxic T cells (CTL), reactive helper T cells (HTL) and anti-polypeptide antibodies are used in well known diagnostic, prognostic and therapeutic assays that examine conditions associated with dysregulated cell growth such as cancer, in particular the cancers listed in Table I (see, e.g., both its specific pattern of tissue expression as well as its overexpression in certain cancers as described, e.g., in the Example entitled "Expression analysis of 101P3A11 in normal tissues and patient specimens").

101P3A11 can be analogized to a prostate associated antigen PSA, the archetypal marker that has been used by medical practitioners for years to identify and monitor the presence of prostate cancer (see, e.g., Merrill et al., J. Urol. 163(2): 503-5120 (2000); Polascik et al., J. Urol. Aug; 162(2):293-306 (1999) and Fortier et al., J. Nat. Cancer Inst. 91(19): 1635-1640(1999)). A variety of other diagnostic markers are also used in similar contexts including p53 and K-ras (see, e.g., Tulchinsky et al., Int J Mol Med 1999 Jul 4(1):99-102 and Minimoto et al., Cancer Detect Prev 2000;24(1):1-12). Therefore, this disclosure of the 101P3A11 polynucleotides and polypeptides (as well as the 101P3A11 polynucleotide probes and anti-101P3A11 antibodies used to identify the presence of these molecules) and their properties allows skilled artisans to utilize these molecules in methods that are analogous to those used, for example, in a variety of diagnostic assays directed to examining conditions associated with cancer.

Typical embodiments of diagnostic methods which utilize the 101P3A11 polynucleotides, polypeptides, reactive T cells and antibodies are analogous to those methods from well-established diagnostic assays which employ, e.g., PSA polynucleotides, polypeptides, reactive T cells and antibodies. For example, just as PSA polynucleotides are used as probes (for example in Northern analysis, see, e.g., Sharief et al., Biochem. Mol. Biol. Int. 33(3):567-74(1994)) and primers (for example in PCR analysis, see, e.g., Okegawa et al., J. Urol. 163(4): 1189-1190 (2000)) to observe the

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presence and/or the level of PSA mRNAs in methods of monitoring PSA overexpression or the metastasis of prostate cancers, the 101P3A11 polynucleotides described herein can be utilized in the same way to detect 101P3A11 overexpression or the metastasis of prostate and other cancers expressing this gene. Alternatively, just as PSA polypeptides are used to generate antibodies specific for PSA which can then be used to observe the presence and/or the level of PSA proteins in methods to monitor PSA protein overexpression (see, e.g., Stephan et al., Urology 55(4):560-3 (2000)) or the metastasis of prostate cells (see, e.g., Alanen et al., Pathol. Res. Pract. 192(3):233-7 (1996)), the 101P3A11 polypeptides described herein can be utilized to generate antibodies for use in detecting 101P3A11 overexpression or the metastasis of prostate cells and cells of other cancers expressing this gene.

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Specifically, because metastases involves the movement of cancer cells from an organ of origin (such as the lung or prostate gland etc.) to a different area of the body (such as a lymph node), assays which examine a biological sample for the presence of cells expressing 101P3A11 polynucleotides and/or polypeptides can be used to provide evidence of metastasis. For example, when a biological sample from tissue that does not normally contain 101P3A11-expressing cells (lymph node) is found to contain 101P3A11-expressing cells such as the 101P3A11 expression seen in LAPC4 and LAPC9, xenografts isolated from lymph node and bone metastasis, respectively, this finding is indicative of metastasis.

Alternatively 101P3A11 polynucleotides and/or polypeptides can be used to provide evidence of cancer, for example, when cells in a biological sample that do not normally express 101P3A11 or express 101P3A11 at a different level are found to express 101P3A11 or have an increased expression of 101P3A11 (see, e.g., the 101P3A11 expression in the cancers listed in Table I and in patient samples etc. shown in the accompanying Figures). In such assays, artisans may further wish to generate supplementary evidence of metastasis by testing the biological sample for the presence of a second tissue restricted marker (in addition to 101P3A11) such as PSA, PSCA etc. (see, e.g., Alanen et al., Pathol. Res. Pract. 192(3): 233-237 (1996)).

Just as PSA polynucleotide fragments and polynucleotide variants are employed by skilled artisans for use in methods of monitoring PSA, 101P3A11 polynucleotide fragments and polynucleotide variants are used in an analogous manner. In particular, typical PSA polynucleotides used in methods of monitoring PSA are probes or primers which consist of fragments of the PSA cDNA sequence. Illustrating this, primers used to PCR amplify a PSA polynucleotide must include less than the whole PSA sequence to function in the polymerase chain reaction. In the context of such PCR reactions, skilled artisans generally create a variety of different polynucleotide fragments that can be used as primers in order to amplify different portions of a polynucleotide of interest or to optimize amplification reactions (see, e.g., Caetano-Anolles, G. Biotechniques 25(3): 472-476, 478-480 (1998); Robertson et al., Methods Mol. Biol. 98:121-154 (1998)). An additional illustration of the use of such fragments is provided in the Example entitled "Expression analysis of 101P3A11 in normal tissues and patient specimens", where a 101P3A11 polynucleotide fragment is used as a probe to show the expression of 101P3A11 RNAs in cancer cells. In addition, variant polynucleotide sequences are typically used as primers and probes for the corresponding mRNAs in PCR and Northern analyses (see, e.g., Sawai et al., Fetal Diagn. Ther. 1996 Nov-Dec 11(6):407-13 and Current Protocols In Molecular Biology, Volume 2, Unit 2, Frederick M. Ausubel et al. eds., 1995)). Polynucleotide fragments and variants are useful in this context where they are capable of binding to a target polynucleotide sequence (e.g. the 101P3A11 polynucleotide shown in SEQ ID NO: 701) under conditions of high stringency.

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Furthermore, PSA polypeptides which contain an epitope that can be recognized by an antibody or T cell that specifically binds to that epitope are used in methods of monitoring PSA. 101P3A11 polypeptide fragments and polypeptide analogs or variants can also be used in an analogous manner. This practice of using polypeptide fragments or polypeptide variants to generate antibodies (such as anti-PSA antibodies or T cells) is typical in the art with a wide variety of systems such as fusion proteins being used by practitioners (see, e.g., Current Protocols In Molecular Biology, Volume 2, Unit 16, Frederick M. Ausubel et al. eds., 1995). In this context, each epitope(s) functions to provide the architecture with which an antibody or T cell is reactive. Typically, skilled artisans create a variety of different polypeptide fragments that can be used in order to generate immune responses specific for different portions of a polypeptide of interest (see, e.g., U.S. Patent No. 5,840,501 and U.S. Patent No. 5,939,533). For example it may be preferable to utilize a polypeptide comprising one of the 101P3A11 biological motifs discussed herein or a motif-bearing subsequence which is readily identified by one of skill in the art based on motifs available in the art. Polypeptide fragments, variants or analogs are typically useful in this context as long as they comprise an epitope capable of generating an antibody or T cell specific for a target polypeptide sequence (e.g. the 101P3A11 polypeptide shown in SEQ ID NO: 703).

As shown herein, the 101P3A11 polynucleotides and polypeptides (as well as the 101P3A11 polynucleotide probes and anti-101P3A11 antibodies or T cells used to identify the presence of these molecules) exhibit specific properties that make them useful in diagnosing cancers such as those listed in Table I. Diagnostic assays that measure the presence of 101P3A11 gene products, in order to evaluate the presence or onset of a disease condition described herein, such as prostate cancer, are used to identify patients for preventive measures or further monitoring, as has been done so successfully with PSA. Moreover, these materials satisfy a need in the art for molecules having similar or complementary characteristics to PSA in situations where, for example, a definite diagnosis of metastasis of prostatic origin cannot be made on the basis of a test for PSA alone (see, e.g., Alanen et al., Pathol. Res. Pract. 192(3): 233-237 (1996)), and consequently, materials such as 101P3A11 polynucleotides and polypeptides (as well as the 101P3A11 polynucleotide probes and anti-101P3A11 antibodies used to identify the presence of these molecules) must be employed to confirm metastases of prostatic origin.

Finally, in addition to their use in diagnostic assays, the 101P3A11 polynucleotides disclosed herein have a number of other utilities such as their use in the identification of oncogenetic associated chromosomal abnormalities in the chromosomal region to which the 101P3A11 gene maps (see Table XXV). Moreover, in addition to their use in diagnostic assays, the 101P3A11-related proteins and polynucleotides disclosed herein have other utilities such as their use in the forensic analysis of tissues of unknown origin (see, e.g., Takahama K Forensic Sci Int 1996 Jun 28;80(1-2): 63-9).

Additionally, 101P3A11-related proteins or polynucleotides of the invention can be used to treat a pathologic condition characterized by the over-expression of 101P3A11. For example, the amino acid or nucleic acid sequence of Figure 2 or Figure 3, or fragments of either, can be used to generate an immune response to the 101P3A11 antigen. Antibodies or other molecules that react with 101P3A11 can be used to modulate the function of this molecule, and thereby provide a therapeutic benefit.

XII.) Inhibition of 101P3A11 Protein Function

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The invention includes various methods and compositions for inhibiting the binding of 101P3A11 to its binding partner or its association with other protein(s) as well as methods for inhibiting 101P3A11 function.

XII.A.) Inhibition of 101P3A11 With Intracellular Antibodies

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In one approach, a recombinant vector that encodes single chain antibodies that specifically bind to 101P3A11 are introduced into 101P3A11 expressing cells via gene transfer technologies. Accordingly, the encoded single chain anti-101P3A11 antibody is expressed intracellularly, binds to 101P3A11 protein, and thereby inhibits its function. Methods for engineering such intracellular single chain antibodies are well known. Such intracellular antibodies, also known as "intrabodies", are specifically targeted to a particular compartment within the cell, providing control over where the inhibitory activity of the treatment is focused. This technology has been successfully applied in the art (for review, see Richardson and Marasco, 1995, TIBTECH vol. 13). Intrabodies have been shown to virtually eliminate the expression of otherwise abundant cell surface receptors (see, e.g., Richardson et al., 1995, Proc. Natl. Acad. Sci. USA 92: 3137-3141; Beerli et al., 1994, J. Biol. Chem. 289: 23931-23936; Deshane et al., 1994, Gene Ther. 1: 332-337).

Single chain antibodies comprise the variable domains of the heavy and light chain joined by a flexible linker polypeptide, and are expressed as a single polypeptide. Optionally, single chain antibodies are expressed as a single chain variable region fragment joined to the light chain constant region. Well-known intracellular trafficking signals are engineered into recombinant polynucleotide vectors encoding such single chain antibodies in order to precisely target the intrabody to the desired intracellular compartment. For example, intrabodies targeted to the endoplasmic reticulum (ER) are engineered to incorporate a leader peptide and, optionally, a C-terminal ER retention signal, such as the KDEL amino acid motif. Intrabodies intended to exert activity in the nucleus are engineered to include a nuclear localization signal. Lipid moieties are joined to intrabodies in order to tether the intrabody to the cytosolic side of the plasma membrane. Intrabodies can also be targeted to exert function in the cytosol. For example, cytosolic intrabodies are used to sequester factors within the cytosol, thereby preventing them from being transported to their natural cellular destination.

In one embodiment, intrabodies are used to capture 101P3A11 in the nucleus, thereby preventing its activity within the nucleus. Nuclear targeting signals are engineered into such 101P3A11 intrabodies in order to achieve the desired targeting. Such 101P3A11 intrabodies are designed to bind specifically to a particular 101P3A11 domain. In another embodiment, cytosolic intrabodies that specifically bind to the 101P3A11 protein are used to prevent 101P3A11 from gaining access to the nucleus, thereby preventing it from exerting any biological activity within the nucleus (e.g., preventing 101P3A11 from forming transcription complexes with other factors).

In order to specifically direct the expression of such intrabodies to particular cells, the transcription of the intrabody is placed under the regulatory control of an appropriate tumor-specific promoter and/or enhancer. In order to target intrabody expression specifically to prostate, for example, the PSA promoter and/or promoter/enhancer can be utilized (See, for example, U.S. Patent No. 5,919,652 issued 6 July 1999).

XII.B.) Inhibition of 101P3A11 with Recombinant Proteins

In another approach, recombinant molecules bind to 101P3A11 and thereby inhibit 101P3A11 function. For example, these recombinant molecules prevent or inhibit 101P3A11 from accessing/binding to its binding partner(s) or associating with other protein(s). Such recombinant molecules can, for example, contain the reactive part(s) of a 101P3A11 specific antibody molecule. In a particular embodiment, the 101P3A11 binding domain of a 101P3A11 binding

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partner is engineered into a dimeric fusion protein, whereby the fusion protein comprises two 101P3A11 ligand binding domains linked to the Fc portion of a human IgG, such as human IgG1. Such IgG portion can contain, for example, the C_H2 and C_H3 domains and the hinge region, but not the C_H1 domain. Such dimeric fusion proteins are administered in soluble form to patients suffering from a cancer associated with the expression of 101P3A11, whereby the dimeric fusion protein specifically binds to 101P3A11 and blocks 101P3A11 interaction with a binding partner. Such dimeric fusion proteins are further combined into multimeric proteins using known antibody linking technologies.

XII.C.) Inhibition of 101P3A11 Transcription or Translation

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The present invention also comprises various methods and compositions for inhibiting the transcription of the 101P3A11 gene. Similarly, the invention also provides methods and compositions for inhibiting the translation of 101P3A11 mRNA into protein.

In one approach, a method of inhibiting the transcription of the 101P3A11 gene comprises contacting the 101P3A11 gene with a 101P3A11 antisense polynucleotide. In another approach, a method of inhibiting 101P3A11 mRNA translation comprises contacting the 101P3A11 mRNA with an antisense polynucleotide. In another approach, a 101P3A11 specific ribozyme is used to cleave the 101P3A11 message, thereby inhibiting translation. Such antisense and ribozyme based methods can also be directed to the regulatory regions of the 101P3A11 gene, such as the 101P3A11 promoter and/or enhancer elements. Similarly, proteins capable of inhibiting a 101P3A11 gene transcription factor are used to inhibit 101P3A11 mRNA transcription. The various polynucleotides and compositions useful in the aforementioned methods have been described above. The use of antisense and ribozyme molecules to inhibit transcription and translation is well known in the art.

Other factors that inhibit the transcription of 101P3A11 by interfering with 101P3A11 transcriptional activation are also useful to treat cancers expressing 101P3A11. Similarly, factors that interfere with 101P3A11 processing are useful to treat cancers that express 101P3A11. Cancer treatment methods utilizing such factors are also within the scope of the invention.

XII.D.) General Considerations for Therapeutic Strategies

Gene transfer and gene therapy technologies can be used to deliver therapeutic polynucleotide molecules to tumor cells synthesizing 101P3A11 (i.e., antisense, ribozyme, polynucleotides encoding intrabodies and other 101P3A11 inhibitory molecules). A number of gene therapy approaches are known in the art. Recombinant vectors encoding 101P3A11 antisense polynucleotides, ribozymes, factors capable of interfering with 101P3A11 transcription, and so forth, can be delivered to target tumor cells using such gene therapy approaches.

The above therapeutic approaches can be combined with any one of a wide variety of surgical, chemotherapy or radiation therapy regimens. The therapeutic approaches of the invention can enable the use of reduced dosages of chemotherapy (or other therapies) and/or less frequent administration, an advantage for all patients and particularly for those that do not tolerate the toxicity of the chemotherapeutic agent well.

The anti-tumor activity of a particular composition (e.g., antisense, ribozyme, intrabody), or a combination of such compositions, can be evaluated using various *in vitro* and *in vivo* assay systems. *In vitro* assays that evaluate therapeutic activity include cell growth assays, soft agar assays and other assays indicative of tumor promoting activity, binding assays capable of determining the extent to which a therapeutic composition will inhibit the binding of 101P3A11 to a binding partner, etc.

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In vivo, the effect of a 101P3A11 therapeutic composition can be evaluated in a suitable animal model. For example, xenogenic prostate cancer models can be used, wherein human prostate cancer explants or passaged xenograft tissues are introduced into immune compromised animals, such as nude or SCID mice (Klein et al., 1997, Nature Medicine 3: 402-408). For example, PCT Patent Application WO98/16628 and U.S. Patent 6,107,540, Sawyers et al., published April 23, 1998, describes various xenograft models of human prostate cancer capable of recapitulating the development of primary tumors, micrometastasis, and the formation of osteoblastic metastases characteristic of late stage disease. Efficacy can be predicted using assays that measure inhibition of tumor formation, tumor regression or metastasis, and the like.

In vivo assays that evaluate the promotion of apoptosis are useful in evaluating therapeutic compositions. In one embodiment, xenografts from tumor bearing mice treated with the therapeutic composition can be examined for the presence of apoptotic foci and compared to untreated control xenograft-bearing mice. The extent to which apoptotic foci are found in the tumors of the treated mice provides an indication of the therapeutic efficacy of the composition.

The therapeutic compositions used in the practice of the foregoing methods can be formulated into pharmaceutical compositions comprising a carrier suitable for the desired delivery method. Suitable carriers include any material that when combined with the therapeutic composition retains the anti-tumor function of the therapeutic composition and is generally non-reactive with the patient's immune system. Examples include, but are not limited to, any of a number of standard pharmaceutical carriers such as sterile phosphate buffered saline solutions, bacteriostatic water, and the like (see, generally, Remington's Pharmaceutical Sciences 16th Edition, A. Osal., Ed., 1980).

Therapeutic formulations can be solubilized and administered via any route capable of delivering the therapeutic composition to the tumor site. Potentially effective routes of administration include, but are not limited to, intravenous, parenteral, intraperitoneal, intramuscular, intratumor, intradermal, intraorgan, orthotopic, and the like. A preferred formulation for intravenous injection comprises the therapeutic composition in a solution of preserved bacteriostatic water, sterile unpreserved water, and/or diluted in polyvinylchloride or polyethylene bags containing 0.9% sterile Sodium Chloride for Injection, USP. Therapeutic protein preparations can be lyophilized and stored as sterile powders, preferably under vacuum, and then reconstituted in bacteriostatic water (containing for example, benzyl alcohol preservative) or in sterile water prior to injection.

Dosages and administration protocols for the treatment of cancers using the foregoing methods will vary with the method and the target cancer, and will generally depend on a number of other factors appreciated in the art.

XIII.) Kits

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For use in the diagnostic and therapeutic applications described herein, kits are also within the scope of the invention. Such kits can comprise a carrier, package or container that is compartmentalized to receive one or more containers such as vials, tubes, and the like, each of the container(s) comprising one of the separate elements to be used in the method. For example, the container(s) can comprise a probe that is or can be detectably labeled. Such probe can be an antibody or polynucleotide specific for a 101P3A11-related protein or a 101P3A11 gene or message, respectively. Where the method utilizes nucleic acid hybridization to detect the target nucleic acid, the kit can also have containers containing nucleotide(s) for amplification of the target nucleic acid sequence and/or a container comprising a reportermeans, such as a biotin-binding protein, such as avidin or streptavidin, bound to a reporter molecule, such as an

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enzymatic, florescent, or radioisotope label. The kit can include all or part of the amino acid sequence of Figure 2 or Figure 3 or analogs thereof, or a nucleic acid molecules that encodes such amino acid sequences or which hybridize to such sequences.

The kit of the invention will typically comprise the container described above and one or more other containers comprising materials desirable from a commercial and user standpoint, including buffers, diluents, filters, needles, syringes, and package inserts with instructions for use. Any physical object in a kit of the invention, e.g. syringe or vial, can have a label affixed thereto. A label or package insert can have instructions for use in human administration printed thereon. Thus, a label can be present on the container to indicate that the composition is used for a specific therapy or non-therapeutic application, and can also indicate directions for either *in vivo* or *in vitro* use, such as those described above. Directions and or other information can also be included on an insert that is included with the kit.

EXAMPLES

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Various aspects of the invention are further described and illustrated by way of the several examples that follow, none of which are intended to limit the scope of the invention.

Example 1: Expression analysis of 101P3A11 in normal tissues and patient specimens

Analysis of 101P3A11 by RT-PCR is shown in Figure 10A. First strand cDNA was prepared from vital pool 1 (VP1: liver, lung and kidney), vital pool 2 (VP2, pancreas, colon and stomach), prostate xenograft pool, prostate cancer pool, kidney cancer pool, colon cancer pool, breast cancer pool, and cancer metastasis pool. Normalization was performed by PCR using primers to actin and GAPDH. Semi-quantitative PCR, using primers to 101P3A11, was performed at 30 cycles of amplification. Expression of 101P3A11 was observed in prostate xenograft pool, prostate cancer pool, kidney cancer pool, colon cancer pool, breast cancer pool, and cancer metastasis pool, but not in VP1 and VP2. Dot blots using patient-derived amplified cDNAs (Clontech, CA) show upregulation of PHOR-1 in 3/3 prostate cancer patients, 6/14 kidney cancer patients, 2/8 uterine cancer, 1/1 cervical cancer, 3/8 stomach cancer, and in 7/7 rectal cancer patients (Figure 10B). Expression of 101P3A11 was assayed in a panel of human patient cancer specimens (Figure 11). RNA was extracted from a pool of three prostate cancer tumors, kidney cancer tumors, colon cancer tumors, breast cancer tumors, and cancer metastasis pool derived from cancer patients, as well as from normal prostate (NP), normal bladder (NB), normal kidney (NK) and normal colon (NC). Northern blots with 10 µg of total RNA/lane were probed with a 101P3A11 sequence fragment. Size standards in kilobases (kb) are indicated on the side. The results show expression of 101P3A11 in prostate cancer tumors, kidney cancer tumors, colon cancer tumors, breast cancer tumors, cancer metastasis pool, bladder cancer pool, and in the normal prostate but not in the other normal tissues.

Northern blot analysis on individual prostate patient tumor specimens is shown in Figure 12A. RNA was extracted from prostate tumors (T) and their normal adjacent tissues (Nat) derived from prostate cancer patients. Northern blots with 10 µg of total RNA/lane were probed with 101P3A11 sequence. Results showed expression of 101P3A11 in all three patient specimens, and expression is especially upregulated in one of the three prostate tumor tissues.

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RNA in situ analysis using anti-sense 101P3A11 riboprobe showed significant glandular epithelial and basal cell expression in normal prostate (4/4), PIN (1/1), and prostate cancer (6/6) patients. 101P3A11 sense riboprobe had little to no staining. The RNA in situ staining in PIN and prostate cancer is shown in Figure 12B and 12C. The staining intensity in the cancer cells was generally higher than that observed in normal glands (Figure 12D and 12E). The RNA in situ results also demonstrate that the expression observed in the prostate tissues is in the glandular epithelia, basal cells, and cancer cells.

Endogenous expression of the 101P3A11 protein is demonstrated in the immunohistochemistry analysis of the anti-101P3A11 (PEPTIDE 1: amino acids 1-14) rabbit polyclonal antibody (Figure 40A-40F). Staining in prostate cancer is greater than the staining observed in normal prostate. The staining is localized apically within the luminal epithelia of the normal prostate (Figures 40E and 40F). The staining observed in prostate cancer is also localized apically in low to intermediate grade cancer (Figures 40B and 40C) and throughout all cells of more advanced prostate cancer (Figure 40A). Staining was observed in 19/20 normal prostate patients and in all of the nineteen prostate cancer patients analyzed. The prostate cancer cell line, LNCaP also shows similar staining (Figures 40D and 40F) in almost all cells.

In addition, the present protocol was used to identify endogenous expression of the 101P3A11 protein in prostate cancer, bladder cancer, kidney cancer, colon cancer, lung cancer, and breast cancer. Immunohistochemical analysis was performed with the anti-101P3A11 (PEPTIDE 1: amino acids 1-14) rabbit polyclonal antibody (prostate cancer, Figure 41A; bladder cancer, Figure 41B; kidney cancer, Figure 41C; colon cancer, lung cancer, Figure 41E; and breast cancer, Figure 41F). Specific staining is observed in tumor cells of the six cancers analyzed.

Expression of 101P3A11 was also detected in the tumors of two colon cancer patients but not in normal colon tissues (Figure 13), and in five out of six kidney tumors isolated from kidney cancer patients (Figure 14). The expression detected in normal adjacent tissues (isolated from diseased tissues) but not in normal tissues of the kidney (isolated from healthy donors) indicates that these tissues are not fully normal and that 101P3A11 is expressed in early stage tumors. In order to assay for androgen regulation of 101P3A11 expression, LAPC-9 cells were grown in charcoal-stripped medium and stimulated with the synthetic androgen mibolerone, for either 14 or 24 hours (Figure 15A, Figure 15B, and Figure 15C). Northern blots with 10 µg of total RNA/lane were probed with the 101P3A11 sequences (Figure 15A). A picture of the ethidium-bromide staining of the RNA gel is also presented (Figure 15C). Results showed expression of 101P3A11 is not regulated by androgen. The experimental samples were confirmed by testing for the expression of the androgen-regulated prostate cancer gene PSA (Figure 15B). This experiment showed that, as expected, PSA levels go down in presence of charcoal-stripped serum, and expression is induced at 14 and 24 hours in presence of mibolerone.

Analysis of androgen regulation of 101P3A11 *in vivo* is shown in Figure 16. Male mice were injected with LAPC-9AD tumor cells. When tumors reached a palpable size, mice were castrated and tumors harvested at different time points following castration. RNA was isolated from the xenograft tissues. Northern blots with 10 µg of total RNA/lane were probed with 101P3A11sequences. Size standards in kilobases (kb) are indicated on the side. A picture of the ethidium-bromide staining of the RNA gel is also presented in Figure 16. The results showed that expression of 101P3A11 was not affected by androgen deprivation, and therefore, is not androgen regulated.

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Example 2: Production of Recombinant 101P3A11 in Prokaryotic and Yeast Systems

To express recombinant 101P3A11 in prokaryotic cells, the full or partial length 101P3A11 cDNA sequences can be cloned into any one of a variety of expression vectors known in the art. One or more of the following regions of 101P3A11 are expressed in these constructs, amino acids 1 to 317; or any 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30 or more contiguous amino acids from 101P3A11, variants, or analogs thereof.

A. *In vitro* transcription and translation constructs:

pCRII: To generate 101P3A11 sense and anti-sense RNA probes for RNA *in situ* investigations, pCRII constructs (Invitrogen, Carlsbad, CA) are generated encoding either all or fragments of the 101P3A11 cDNA. The pCRII vector has Sp6 and T7 promoters flanking the insert to drive the transcription of 101P3A11 RNA for use as probes in RNA *in situ* hybridization experiments. These probes are used to analyze the cell and tissue expression of 101P3A11 at the RNA level. Transcribed 101P3A11 RNA representing the cDNA amino acid coding region of the 101P3A11 gene is used in *in vitro* translation systems such as the TnTTM Coupled Reticulolysate System (Promega, Corp., Madison, WI) to synthesize 101P3A11 protein.

B. Bacterial Constructs:

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pGEX Constructs: To generate recombinant 101P3A11 proteins in bacteria that are fused to the Glutathione Stransferase (GST) protein, all or parts of the 101P3A11 cDNA protein coding sequence are fused to the GST gene by cloning into pGEX-6P-1 or any other GST- fusion vector of the pGEX family (Amersham Pharmacia Biotech, Piscataway, NJ). These constructs allow controlled expression of recombinant 101P3A11 protein sequences with GST fused at the amino-terminus and a six histidine epitope (6X His) at the carboxyl-terminus. The GST and 6X His tags permit purification of the recombinant fusion protein from induced bacteria with the appropriate affinity matrix and allow recognition of the fusion protein with anti-GST and anti-His antibodies. The 6X His tag is generated by adding 6 histidine codons to the cloning primer at the 3' end, e.g., of the open reading frame (ORF). A proteolytic cleavage site, such as the PreScissionTM recognition site in pGEX-6P-1, can be employed that permits cleavage of the GST tag from 101P3A11-related protein. The ampicillin resistance gene and pBR322 origin permit selection and maintenance of the pGEX plasmids in *E. coli*. In one embodiment, amino acids 86-317 are cloned into the pGEX-2T expression vector, the protein is expressed and purified.

pMAL Constructs: To generate, in bacteria, recombinant 101P3A11 proteins that are fused to maltose-binding protein (MBP), all or parts of the 101P3A11 cDNA protein coding sequence are fused to the MBP gene by cloning into the pMAL-c2X and pMAL-p2X vectors (New England Biolabs, Beverly, MA). These constructs allow controlled expression of recombinant 101P3A11 protein sequences with MBP fused at the amino-terminus and a 6X His epitope tag at the carboxyl-terminus. The MBP and 6X His tags permit purification of the recombinant protein from induced bacteria with the appropriate affinity matrix and allow recognition of the fusion protein with anti-MBP and anti-His antibodies. The 6X His epitope tag is generated by adding 6 histidine codons to the 3' cloning primer. A Factor Xa recognition site permits cleavage of the pMAL tag from 101P3A11. The pMAL-c2X and pMAL-p2X vectors are optimized to express the recombinant protein in the cytoplasm or periplasm respectively. Periplasm expression enhances folding of proteins with disulfide bonds. In one embodiment, amino acids 86-310 is cloned into the pMAL-c2X expression vector, the protein is expressed and purified.

pET Constructs: To express 101P3A11 in bacterial cells, all or parts of the 101P3A11 cDNA protein coding sequence are cloned into the pET family of vectors (Novagen, Madison, WI). These vectors allow tightly controlled expression of recombinant 101P3A11 protein in bacteria with and without fusion to proteins that enhance solubility, such as NusA and thioredoxin (Trx), and epitope tags, such as 6X His and S-Tag TM that aid purification and detection of the recombinant protein. For example, constructs are made utilizing pET NusA fusion system 43.1 such that regions of the 101P3A11 protein are expressed as amino-terminal fusions to NusA.

C. Yeast Constructs:

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pESC Constructs: To express 101P3A11 in the yeast species Saccharomyces cerevisiae for generation of recombinant protein and functional studies, all or parts of the 101P3A11 cDNA protein coding sequence are cloned into the pESC family of vectors each of which contain 1 of 4 selectable markers, HIS3, TRP1, LEU2, and URA3 (Stratagene, La Jolla, CA). These vectors allow controlled expression from the same plasmid of up to 2 different genes or cloned sequences containing either FlagTM or Myc epitope tags in the same yeast cell. This system is used to confirm protein-protein interactions of 101P3A11. In addition, expression in yeast yields similar post-translational modifications, such as glycosylations and phosphorylations, that are found when expressed in eukaryotic cells.

pESP Constructs: To express 101P3A11 in the yeast species Saccharomyces pombe, all or parts of the 101P3A11 cDNA protein coding sequence are cloned into the pESP family of vectors. These vectors allow controlled high level expression of a 101P3A11 protein sequence that is fused at either the amino terminus or at the carboxyl terminus to GST which aids purification of the recombinant protein. A FlagTM epitope tag allows detection of the recombinant protein with anti- FlagTM antibody.

Example 3: Production of Recombinant 101P3A11 in Higher Eukaryotic Systems

A. Mammalian Constructs:

To express recombinant 101P3A11 in eukaryotic cells, full or partial length 101P3A11 cDNA sequences can be cloned into any one of a variety of expression vectors known in the art. One or more of the following regions of 101P3A11 are expressed in these constructs, amino acids 1 to 317; or any 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30 or more contiguous amino acids from 101P3A11, variants, or analogs thereof.

The constructs can be transfected into any one of a wide variety of mammalian cells such as 293T cells. Transfected 293T cell lysates can be probed with the anti-101P3A11 polyclonal serum, described herein.

pcDNA4/HisMax Constructs: To express 101P3A11 in mammalian cells, the 101P3A11 ORF was cloned into pcDNA4/HisMax Version A (Invitrogen, Carlsbad, CA). Protein expression is driven from the cytomegalovirus (CMV) promoter and the SP16 translational enhancer. The recombinant protein has XpressTM and six histidine (6X His) epitopes fused to the amino-terminus. The pcDNA4/HisMax vector also contains the bovine growth hormone (BGH) polyadenylation signal and transcription termination sequence to enhance mRNA stability along with the SV40 origin for episomal replication and simple vector rescue in cell lines expressing the large T antigen. The Zeocin resistance gene allows for selection of mammalian cells expressing the protein and the ampicillin resistance gene and ColE1 origin permits selection and maintenance of the plasmid in *E. coli*.

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pcDNA3.1/MycHis Constructs: To express 101P3A11 in mammalian cells, the 101P3A11 ORF, with a consensus Kozak translation initiation site, was cloned into pcDNA3.1/MycHis Version A (Invitrogen, Carlsbad, CA). Protein expression is driven from the cytomegalovirus (CMV) promoter. The recombinant proteins have the myc epitope and 6X His epitope fused to the carboxyl-terminus. The pcDNA3.1/MycHis vector also contains the bovine growth hormone (BGH) polyadenylation signal and transcription termination sequence to enhance mRNA stability, along with the SV40 origin for episomal replication and simple vector rescue in cell lines expressing the large T antigen. The Neomycin resistance gene can be used, as it allows for selection of mammalian cells expressing the protein and the ampicillin resistance gene and ColE1 origin permits selection and maintenance of the plasmid in *E. coli*.

pcDNA3.1/CT-GFP-TOPO Construct: To express 101P3A11 in mammalian cells and to allow detection of the recombinant proteins using fluorescence, the 101P3A11 ORF, with a consensus Kozak translation initiation site, was cloned into pcDNA3.1/CT-GFP-TOPO (Invitrogen, CA). Protein expression is driven from the cytomegalovirus (CMV) promoter. The recombinant proteins have the Green Fluorescent Protein (GFP) fused to the carboxyl-terminus facilitating non-invasive, *in vivo* detection and cell biology studies. The pcDNA3.1CT-GFP-TOPO vector also contains the bovine growth hormone (BGH) polyadenylation signal and transcription termination sequence to enhance mRNA stability along with the SV40 origin for episomal replication and simple vector rescue in cell lines expressing the large T antigen. The Neomycin resistance gene allows for selection of mammalian cells that express the protein, and the ampicillin resistance gene and ColE1 origin permits selection and maintenance of the plasmid in *E. coli*. Additional constructs with an amino-terminal GFP fusion are made in pcDNA3.1/NT-GFP-TOPO spanning the entire length of the 101P3A11 proteins.

PAPtag: The 101P3A11 ORF, or portions thereof, of 101P3A11 are cloned into pAPtag-5 (GenHunter Corp. Nashville, TN). This construct generates an alkaline phosphatase fusion at the carboxyl-terminus of the 101P3A11 proteins while fusing the IgGκ signal sequence to the amino-terminus. Constructs are also generated in which alkaline phosphatase with an amino-terminal IgGκ signal sequence is fused to the amino-terminus of 101P3A11 proteins. The resulting recombinant 101P3A11 proteins are optimized for secretion into the media of transfected mammalian cells and can be used to identify proteins such as ligands or receptors that interact with the 101P3A11 proteins. Protein expression is driven from the CMV promoter and the recombinant proteins also contain myc and 6X His epitopes fused at the carboxyl-terminus that facilitates detection and purification. The Zeocin resistance gene present in the vector allows for selection of mammalian cells expressing the recombinant protein and the ampicillin resistance gene permits selection of the plasmid in *E. coli*.

ptag5: The 101P3A11 ORF, or portions thereof, of 101P3A11 are cloned into pTag-5. This vector is similar to pAPtag but without the alkaline phosphatase fusion. This construct generated 101P3A11 protein with an aminoterminal IgGκ signal sequence and myc and 6X His epitope tags at the carboxyl-terminus that facilitate detection and affinity purification. The resulting recombinant 101P3A11 protein was optimized for secretion into the media of transfected mammalian cells, and was used as immunogen or ligand to identify proteins such as ligands or receptors that interact with the 101P3A11 proteins. Protein expression is driven from the CMV promoter. The Zeocin resistance gene present in the vector allows for selection of mammalian cells expressing the protein, and the ampicillin resistance gene permits selection of the plasmid in *E. coli*.

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PsecFc: The 101P3A11 ORF, or portions thereof, of 101P3A11 are also cloned into psecFc. The psecFc vector was assembled by cloning the human immunoglobulin G1 (IgG) Fc (hinge, CH2, CH3 regions) into pSecTag2 (Invitrogen, California). This construct generates an IgG1 Fc fusion at the carboxyl-terminus of the 101P3A11 proteins, while fusing the IgGκ signal sequence to N-terminus. 101P3A11 fusions utilizing the murine IgG1 Fc region are also used. The resulting recombinant 101P3A11 proteins are optimized for secretion into the media of transfected mammalian cells, and can be used as immunogens or to identify proteins such as ligands or receptors that interact with the 101P3A11 protein. Protein expression is driven from the CMV promoter. The hygromycin resistance gene present in the vector allows for selection of mammalian cells that express the recombinant protein, and the ampicillin resistance gene permits selection of the plasmid in *E. coli*.

The amino acid region 159-202 of the 101P3A11 ORF was cloned into psecFc. The resulting recombinant 101P3A11(159-202)-psecFc construct was transfected into 293T and Cos-7 cells, and the expression of recombinant 101P3A11(159-202)-psecFc protein assayed by Western blotting (Figure 17). Results show that 101P3A11(159-202)-psecFc fusion protein was expressed in the lysates of both 293T and Cos-7 cells. The 101P3A11(159-202)-psecFc fusion protein was also secreted and detected in the culture supernatants of both cell types.

pSRα Constructs: To generate mammalian cell lines that express 101P3A11, constitutively, the ORF of 101P3A11 was cloned into pSRα constructs. Amphotropic and ecotropic retroviruses were generated by transfection of pSRα constructs into the 293T-10A1 packaging line or co-transfection of pSRα and a helper plasmid (containing deleted packaging sequences) into the 293 cells, respectively. The retrovirus was used to infect a variety of mammalian cell lines, resulting in the integration of the cloned gene, 101P3A11, into the host cell lines. Protein expression is driven from a long terminal repeat (LTR). The Neomycin resistance gene present in the vector allows for selection of mammalian cells that express the protein, and the ampicillin resistance gene and ColE1 origin permit selection and maintenance of the plasmid in *E. coli*. Figure 18 shows that 101P3A11 was expressed using the pSRα retroviral vector in the cell line 300.19. The retroviral vectors can thereafter be used for infection and generation of various cell lines using, for example, PC3, NIH 3T3, TsuPr1, 293 or rat-1 cells.

Additional pSR α constructs are made that fuse an epitope tag such as the FLAGTM tag to the carboxyl-terminus of 101P3A11 sequences to allow detection using anti-Flag antibodies. For example, the FLAGTM sequence 5' gat tac aag gat gac gat aag 3' is added to cloning primer at the 3' end of the ORF. Additional pSR α constructs are made to produce both amino-terminal and carboxyl-terminal GFP and myc/6X His fusion proteins of the full-length 101P3A11 proteins.

Additional Viral Vectors: Additional constructs are made for viral-mediated delivery and expression of 101P3A11. High virus titer leading to high level expression of 101P3A11 is achieved in viral delivery systems such as adenoviral vectors and herpes amplicon vectors. The 101P3A11 coding sequences or fragments thereof are amplified by PCR and subcloned into the AdEasy shuttle vector (Stratagene). Recombination and virus packaging are performed according to the manufacturer's instructions to generate adenoviral vectors. Alternatively, 101P3A11 coding sequences or fragments thereof are cloned into the HSV-1 vector (Imgenex) to generate herpes viral vectors. The viral vectors are thereafter used for infection of various cell lines such as SCaBER, NIH 3T3, 293 or rat-1 cells.

Regulated Expression Systems: To control expression of 101P3A11 in mammalian cells, coding sequences of 101P3A11, or portions thereof, are cloned into regulated mammalian expression systems such as the T-Rex System sd-63712

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(Invitrogen), the GeneSwitch System (Invitrogen) and the tightly-regulated Ecdysone System (Sratagene). These systems allow the study of the temporal and concentration dependent effects of recombinant 101P3A11. These vectors are thereafter used to control expression of 101P3A11 in various cell lines such as SCaBER, NIH 3T3, 293 or rat-1 cells.

B. Baculovirus Expression Systems

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To generate recombinant 101P3A11 proteins in a baculovirus expression system, 101P3A11 ORF, or portions thereof, are cloned into the baculovirus transfer vector pBlueBac 4.5 (Invitrogen), which provides a His-tag at the N-terminus. Specifically, pBlueBac-101P3A11 is co-transfected with helper plasmid pBac-N-Blue (Invitrogen) into SF9 (Spodoptera frugiperda) insect cells to generate recombinant baculovirus (see Invitrogen instruction manual for details). Baculovirus is then collected from cell supernatant and purified by plaque assay.

Recombinant 101P3A11 protein is then generated by infection of HighFive insect cells (Invitrogen) with purified baculovirus. Recombinant 101P3A11 protein can be detected using anti-101P3A11 or anti-His-tag antibody. 101P3A11 protein can be purified and used in various cell-based assays or as immunogen to generate polyclonal and monoclonal antibodies specific for 101P3A11.

Example 4 Antigenicity Profiles and Secondary Structure

Figure 5, Figure 6, Figure 7, Figure 8, and Figure 9 depict graphically five amino acid profiles of the 101P3A11 amino acid sequence, each assessment available by accessing the ProtScale website (URL www.expasy.ch/cgi-bin/protscale.pl) on the ExPasy molecular biology server.

These profiles: Figure 5, Hydrophilicity, (Hopp T.P., Woods K.R., 1981. Proc. Natl. Acad. Sci. U.S.A. 78:3824-3828); Figure 6, Hydropathicity, (Kyte J., Doolittle R.F., 1982. J. Mol. Biol. 157:105-132); Figure 7, Percentage Accessible Residues (Janin J., 1979 Nature 277:491-492); Figure 8, Average Flexibility, (Bhaskaran R., and Ponnuswamy P.K., 1988. Int. J. Pept. Protein Res. 32:242-255); Figure 9, Beta-turn (Deleage, G., Roux B. 1987 Protein Engineering 1:289-294); and optionally others available in the art, such as on the ProtScale website, were used to identify antigenic regions of the 101P3A11 protein. Each of the above amino acid profiles of 101P3A11 were generated using the following ProtScale parameters for analysis: 1) A window size of 9; 2) 100% weight of the window edges compared to the window center; and, 3) amino acid profile values normalized to lie between 0 and 1.

Hydrophilicity (Figure 5), Hydropathicity (Figure 6) and Percentage Accessible Residues (Figure 7) profiles were used to determine stretches of hydrophilic amino acids (i.e., values greater than 0.5 on the Hydrophilicity and Percentage Accessible Residues profile, and values less than 0.5 on the Hydropathicity profile). Such regions are likely to be exposed to the aqueous environment, be present on the surface of the protein, and thus are available for immune recognition, such as by antibodies.

Average Flexibility (Figure 8) and Beta-turn (Figure 9) profiles determine stretches of amino acids (i.e., values greater than 0.5 on the Beta-turn profile and the Average Flexibility profile) that are not constrained in secondary structures such as beta sheets and alpha helices. Such regions are also more likely to be exposed portions of the protein and thus are accessible to immune recognition, such as by antibodies.

Antigenic sequences of the 101P3A11 protein indicated, e.g., by the profiles set forth in Figure 5, Figure 6, Figure 7, Figure 8, and/or Figure 9 are used to prepare immunogens, either peptides or nucleic acids that encode them,

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to generate anti-101P3A11 antibodies. The immunogen can be any 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30, 35, 40, 45, 50 or more than 50 contiguous amino acids, or the corresponding nucleic acids that encode them, from the 101P3A11 protein. In particular, peptide immunogens of the invention can comprise, a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 317 that includes an amino acid position having a value greater than 0.5 in the Hydrophilicity profile of Figure 5; a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 317 that includes an amino acid position having a value less than 0.5 in the Hydropathicity profile of Figure 6; a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 317 that includes an amino acid position having a value greater than 0.5 in the Percent Accessible Residues profile of Figure 7; a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 317 that includes an amino acid position having a value greater than 0.5 in the Average Flexibility profile on Figure 8; and, a peptide region of at least 5 amino acids of Figure 2 in any whole number increment up to 317 that includes an amino acid position having a value greater than 0.5 in the Beta-turn profile of Figure 9. Peptide immunogens of the invention can also comprise nucleic acids that encode any of the forgoing.

All immunogens of the invention, peptide or nucleic acid, can be embodied in human unit dose form, or comprised by a composition that includes a pharmaceutical excipient compatible with human physiology.

The secondary structure of 101P3A11, namely the predicted presence and location of alpha helices, extended strands, and random coils, is predicted from the primary amino acid sequence using the HNN - Hierarchical Neural Network method (Guermeur, 1997, http://pbil.ibcp.fr/cgi-bin/npsa_automat.pl?page=npsa_nn.html), accessed from the ExPasy molecular biology server http://www.expasy.ch/tools/. The analysis indicates that 101P3A11 is composed 47.95% alpha helix, 21.45% extended strand, and 30.60% random coil (Figure 19A).

Analysis for the potential presence of transmembrane domains in 101P3A11 was carried out using a variety of transmembrane prediction algorithms accessed from the ExPasy molecular biology server http://www.expasy.ch/tools. The programs predict the presence of 7 transmembrane domains in 101P3A11, consistent with the structure of a G-protein coupled receptor. Shown graphically in Figure 19A are the results of analysis using the TMpred (Figure 19B) and TMHMM (Figure 19C) prediction programs depicting the location of the 7 transmembrane domains. The results of each program, namely the amino acids encoding the transmembrane domains are summarized in Table XXV.

Example 5: Generation of 101P3A11 Polyclonal Antibodies

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Polyclonal antibodies can be raised in a mammal, for example, by one or more injections of an immunizing agent and, if desired, an adjuvant. Typically, the immunizing agent and/or adjuvant will be injected in the mammal by multiple subcutaneous or intraperitoneal injections. In addition to immunizing with the full length 101P3A11 protein, computer algorithms are employed in design of immunogens that, based on amino acid sequence analysis are antigenic and available for recognition by the immune system of the immunized host (see the Example entitled "Antigenicity Profiles and Secondary Structure"). Such regions would generally be hydrophilic, flexible, in beta-turn conformations, and/or exposed on the surface of the protein (see, e.g., Figure 5, Figure 6, Figure 7, Figure 8, or Figure 9 for amino acid profiles that indicate such regions of 101P3A11).

For example, 101P3A11 recombinant bacterial fusion proteins or peptides containing hydrophilic, flexible, beta-turn regions of the 101P3A11 amino acid sequence, such as amino acids 1-23, plus or minus 1-10 amino acids at

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available termini, and amino acids 159-202, plus or minus 1-10 amino acids at available termini, are used as antigens to generate polyclonal antibodies in New Zealand White rabbits. It is useful to conjugate the immunizing agent to a protein known to be immunogenic in the mammal being immunized. Examples of such immunogenic proteins include, but are not limited to, keyhole limpet hemocyanin (KLH), serum albumin, bovine thyroglobulin, and soybean trypsin inhibitor. In one embodiment, a peptide encoding amino acids 1-23 of 101P3A11 is conjugated to KLH and used to immunize the rabbit. Alternatively the immunizing agent may include all or portions of the 101P3A11 protein, analogs or fusion proteins thereof. For example, the 101P3A11 amino acid sequence can be fused using recombinant DNA techniques to any one of a variety of fusion protein partners that are well known in the art, such as glutathione-S-transferase (GST) and HIS tagged fusion proteins. Such fusion proteins are purified from induced bacteria using the appropriate affinity matrix.

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In one embodiment, a GST-fusion protein encoding amino acids 86-317, plus or minus 1-10 amino acids at available termini, is produced and purified and used as immunogen. Other recombinant bacterial fusion proteins that may be employed include maltose binding protein, LacZ, thioredoxin, NusA, or an immunoglobulin constant region (see the section entitled "Production of 101P3A11 in Prokaryotic Systems" and Current Protocols In Molecular Biology, Volume 2, Unit 16, Frederick M. Ausubul et al. eds., 1995; Linsley, P.S., Brady, W., Urnes, M., Grosmaire, L., Damle, N., and Ledbetter, L.(1991) J.Exp. Med. 174, 561-566).

In addition to bacterial derived fusion proteins, mammalian expressed protein antigens are also used. These antigens are expressed from mammalian expression vectors such as the Tag5 and Fc-fusion vectors (see the section entitled "Production of Recombinant 101P3A11 in Eukaryotic Systems"), and retain post-translational modifications such as glycosylations found in native protein. In one embodiment, amino acids 159-202 is cloned into the Tag5 mammalian secretion vector. The recombinant protein is purified by metal chelate chromatography from tissue culture supernatants of 293T cells stably expressing the recombinant vector. The purified Tag5 101P3A11 protein is then used as immunogen.

During the immunization protocol, it is useful to mix or emulsify the antigen in adjuvants that enhance the immune response of the host animal. Examples of adjuvants include, but are not limited to, complete Freund's adjuvant (CFA) and MPL-TDM adjuvant (monophosphoryl Lipid A, synthetic trehalose dicorynomycolate).

In a typical protocol, rabbits are initially immunized subcutaneously with up to 200 μ g, typically 100-200 μ g, of fusion protein or peptide conjugated to KLH mixed in complete Freund's adjuvant (CFA). Rabbits are then injected subcutaneously every two weeks with up to 200 μ g, typically 100-200 μ g, of the immunogen in incomplete Freund's adjuvant (IFA). Test bleeds are taken approximately 7-10 days following each immunization and used to monitor the titer of the antiserum by ELISA.

To test reactivity and specificity of immune serum, such as the rabbit serum derived from immunization with Tag5 101P3A11 encoding amino acids 159-202, the full-length 101P3A11 cDNA is cloned into pCDNA 3.1 myc-his expression vector (Invitrogen, see the Example entitled "Production of Recombinant 101P3A11 in Eukaryotic Systems"). After transfection of the constructs into 293T cells, cell lysates are probed with the anti-101P3A11 serum and with anti-His antibody (Santa Cruz Biotechnologies, Santa Cruz, CA) to determine specific reactivity to denatured 101P3A11 protein using the Western blot technique. Immunoprecipitation and flow cytometric analyses of 293T and other recombinant 101P3A11-expressing cells determine recognition of native protein by the antiserum. In addition,

Western blot, immunoprecipitation, fluorescent microscopy, and flow cytometric techniques using cells that endogenously express 101P3A11 are carried out to test specificity.

The anti-serum from the Tag5 101P3A11 immunized rabbit is affinity purified by passage over a column composed of the Tag5 antigen covalently coupled to Affigel matrix (BioRad, Hercules, Calif.). The serum is then further purified by protein G affinity chromatography to isolate the IgG fraction. Serum from rabbits immunized with fusion proteins, such as GST and MBP fusion proteins, are purified by depletion of antibodies reactive to the fusion partner sequence by passage over an affinity column containing the fusion partner either alone or in the context of an irrelevant fusion protein. Sera from other His-tagged antigens and peptide immunized rabbits as well as fusion partner depleted sera are affinity purified by passage over a column matrix composed of the original protein immunogen or free peptide.

Example 6: Generation of 101P3A11 Monoclonal Antibodies (mAbs)

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In one embodiment, therapeutic mAbs to 101P3A11 comprise those that react with epitopes of the protein that would disrupt or modulate the biological function of 101P3A11, for example those that would disrupt its interaction with ligands or proteins that mediate or are involved in its biological activity. Therapeutic mAbs also comprise those that specifically bind epitopes of 101P3A11 exposed on the cell surface and thus are useful in targeting mAb-toxin conjugates. Immunogens for generation of such mAbs include those designed to encode or contain the entire 101P3A11 protein or regions of the 101P3A11 protein predicted to be exposed to the extracellular environment and/or antigenic from computer analysis of the amino acid sequence (see, e.g., Figure 5, Figure 6, Figure 7, Figure 8, or Figure 9, and the Example entitled "Antigenicity Profiles and Secondary Structure"). Immunogens include peptides, recombinant bacterial proteins, and mammalian expressed Tag 5 proteins and human and murine IgG FC fusion proteins. In addition, cells expressing high levels of 101P3A11, such as 293T-101P3A11 or 300.19-101P3A11 murine Pre-B cells, are used to immunize mice.

To generate mAbs to 101P3A11, mice are first immunized intraperitoneally (IP) with, typically, 10-50 μg of protein immunogen or 10⁷ 101P3A11-expressing cells mixed in complete Freund's adjuvant. Alternatively, mice are immunized intradermally. Mice are then subsequently immunized IP every 2-4 weeks with, typically, 10-50 μg of protein immunogen or 10⁷ cells mixed in incomplete Freund's adjuvant. Alternatively, MPL-TDM adjuvant is used in immunizations. In addition to the above protein and cell-based immunization strategies, a DNA-based immunization protocol is employed in which a mammalian expression vector encoding 101P3A11 sequence is used to immunize mice by direct injection of the plasmid DNA. For example, the predicted first extracellular loop, amino acids 82-104, or second extracellular loop of 101P3A11, amino acids 159-202, or the third extracellular loop, amino acids 258 - 275 (in each instance plus or minus 10 amino acids) is cloned into the Tag5 mammalian secretion vector and the recombinant vector is used as immunogen. In another example the same amino acids are cloned into an Fc-fusion secretion vector in which the 101P3A11 sequence is fused at the amino-terminus to an IgK leader sequence and at the carboxyl-terminus to the coding sequence of the human IgG Fc region. This recombinant vector is then used as immunogen. The plasmid immunization protocols are used in combination with purified proteins expressed from the same vector and with cells expressing 101P3A11.

During the immunization protocol, test bleeds are taken 7-10 days following an injection to monitor titer and specificity of the immune response. Once appropriate reactivity and specificity is obtained as determined by ELISA, Western blotting, immunoprecipitation, fluorescence microscopy, and flow cytometric analyses, fusion and hybridoma generation is then carried out with established procedures well known in the art (see, e.g., Harlow and Lane, 1988).

In one embodiment for generating 101P3A11 monoclonal antibodies, a Tag5-101P3A11 antigen encoding amino acids 159-202 is expressed and purified from stably transfected 293T cells. Balb C mice are initially immunized intraperitoneally with 25 µg of the Tag5-101P3A11 protein mixed in complete Freund's adjuvant. Mice are subsequently immunized every two weeks with 25 µg of the antigen mixed in incomplete Freund's adjuvant for a total of three immunizations. ELISA using the Tag5 antigen determines the titer of serum from immunized mice. Reactivity and specificity of serum to full length 101P3A11 protein is monitored by Western blotting, immunoprecipitation and flow cytometry using 293T cells transfected with an expression vector encoding the 101P3A11 cDNA (see e.g., the Example entitled "Production of Recombinant 101P3A11 in Eukaryotic Systems"). Other recombinant 101P3A11-expressing cells or cells endogenously expressing 101P3A11 are also used. Mice showing the strongest reactivity are rested and given a final injection of Tag5 antigen in PBS and then sacrificed four days later. The spleens of the sacrificed mice are harvested and fused to SPO/2 myeloma cells using standard procedures (Harlow and Lane, 1988). Supernatants from HAT selected growth wells are screened by ELISA, Western blot, immunoprecipitation, fluorescent microscopy, and flow cytometry to identify 101P3A11 specific antibody-producing clones.

The binding affinity of a 101P3A11 monoclonal antibody is determined using standard technologies. Affinity measurements quantify the strength of antibody to epitope binding and are used to help define which 101P3A11 monoclonal antibodies preferred, e.g., for diagnostic or therapeutic use, as appreciated by one of skill in the art. The BIAcore system (Uppsala, Sweden) is a preferred method for determining binding affinity. The BIAcore system uses surface plasmon resonance (SPR, Welford K. 1991, Opt. Quant. Elect. 23:1; Morton and Myszka, 1998, Methods in Enzymology 295: 268) to monitor biomolecular interactions in real time. BIAcore analysis conveniently generates association rate constants, dissociation rate constants, equilibrium dissociation constants, and affinity constants.

Example 7: HLA Class I and Class II Binding Assays

HLA class I and class II binding assays using purified HLA molecules are performed in accordance with disclosed protocols (e.g., PCT publications WO 94/20127 and WO 94/03205; Sidney et al., Current Protocols in Immunology 18.3.1 (1998); Sidney, et al., J. Immunol. 154:247 (1995); Sette, et al., Mol. Immunol. 31:813 (1994)). Briefly, purified MHC molecules (5 to 500 nM) are incubated with various unlabeled peptide inhibitors and 1-10 nM ¹²⁵I-radiolabeled probe peptides as described. Following incubation, MHC-peptide complexes are separated from free peptide by gel filtration and the fraction of peptide bound is determined. Typically, in preliminary experiments, each MHC preparation is titered in the presence of fixed amounts of radiolabeled peptides to determine the concentration of HLA molecules necessary to bind 10-20% of the total radioactivity. All subsequent inhibition and direct binding assays are performed using these HLA concentrations.

Since under these conditions [label]<[HLA] and IC₅₀ \geq [HLA], the measured IC₅₀ values are reasonable approximations of the true K_D values. Peptide inhibitors are typically tested at concentrations ranging from 120 μ g/ml to 1.2 ng/ml, and are tested in two to four completely independent experiments. To allow comparison of the data

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obtained in different experiments, a relative binding figure is calculated for each peptide by dividing the IC_{50} of a positive control for inhibition by the IC_{50} for each tested peptide (typically unlabeled versions of the radiolabeled probe peptide). For database purposes, and inter-experiment comparisons, relative binding values are compiled. These values can subsequently be converted back into IC_{50} nM values by dividing the IC_{50} nM of the positive controls for inhibition by the relative binding of the peptide of interest. This method of data compilation is accurate and consistent for comparing peptides that have been tested on different days, or with different lots of purified MHC.

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Binding assays as outlined above may be used to analyze HLA supermotif and/or HLA motif-bearing peptides.

Example 8: Identification of HLA Supermotif- and Motif-Bearing CTL Candidate Epitopes

HLA vaccine compositions of the invention can include multiple epitopes. The multiple epitopes can comprise multiple HLA supermotifs or motifs to achieve broad population coverage. This example illustrates the identification and confirmation of supermotif- and motif-bearing epitopes for the inclusion in such a vaccine composition.

Calculation of population coverage is performed using the strategy described below.

Computer searches and algorithms for identification of supermotif and/or motif-bearing epitopes

The searches performed to identify the motif-bearing peptide sequences in the Example entitled "Antigenicity Profiles" and Tables V-XVIII or XXVI-XXVIII employ the protein sequence data from the gene product of 101P3A11 set forth in Figures 2 and 3.

Computer searches for epitopes bearing HLA Class I or Class II supermotifs or motifs are performed as follows. All translated 101P3A11 protein sequences are analyzed using a text string search software program to identify potential peptide sequences containing appropriate HLA binding motifs; such programs are readily produced in accordance with information in the art in view of known motif/supermotif disclosures. Furthermore, such calculations can be made mentally.

Identified A2-, A3-, and DR-supermotif sequences are scored using polynomial algorithms to predict their capacity to bind to specific HLA-Class I or Class II molecules. These polynomial algorithms account for the impact of different amino acids at different positions, and are essentially based on the premise that the overall affinity (or ΔG) of peptide-HLA molecule interactions can be approximated as a linear polynomial function of the type:

"
$$\Delta G$$
" = $a_{1i} \times a_{2i} \times a_{3i} \dots \times a_{ni}$

where a_{ji} is a coefficient which represents the effect of the presence of a given amino acid (j) at a given position (i) along the sequence of a peptide of n amino acids. The crucial assumption of this method is that the effects at each position are essentially independent of each other (i.e., independent binding of individual side-chains). When residue j occurs at position i in the peptide, it is assumed to contribute a constant amount j_i to the free energy of binding of the peptide irrespective of the sequence of the rest of the peptide.

The method of derivation of specific algorithm coefficients has been described in Gulukota et al., J. Mol. Biol. 267:1258-126, 1997; (see also Sidney et al., Human Immunol. 45:79-93, 1996; and Southwood et al., J. Immunol. 160:3363-3373, 1998). Briefly, for all i positions, anchor and non-anchor alike, the geometric mean of the average relative binding (ARB) of all peptides carrying j is calculated relative to the remainder of the group, and used as the estimate of j_i . For Class II peptides, if multiple alignments are possible, only the highest scoring alignment is utilized, following an iterative procedure. To calculate an algorithm score of a given peptide in a test set, the ARB values

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corresponding to the sequence of the peptide are multiplied. If this product exceeds a chosen threshold, the peptide is predicted to bind. Appropriate thresholds are chosen as a function of the degree of stringency of prediction desired.

Selection of HLA-A2 supertype cross-reactive peptides

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Complete protein sequences from 101P3A11 are scanned utilizing motif identification software, to identify 8-, 9- 10- and 11-mer sequences containing the HLA-A2-supermotif main anchor specificity. Typically, these sequences are then scored using the protocol described above and the peptides corresponding to the positive-scoring sequences are synthesized and tested for their capacity to bind purified HLA-A*0201 molecules *in vitro* (HLA-A*0201 is considered a prototype A2 supertype molecule).

These peptides are then tested for the capacity to bind to additional A2-supertype molecules (A*0202, A*0203, A*0206, and A*6802). Peptides that bind to at least three of the five A2-supertype alleles tested are typically deemed A2-supertype cross-reactive binders. Preferred peptides bind at an affinity equal to or less than 500 nM to three or more HLA-A2 supertype molecules.

Selection of HLA-A3 supermotif-bearing epitopes

The 101P3A11 protein sequence scanned above is also examined for the presence of peptides with the HLA-A3-supermotif primary anchors. Peptides corresponding to the HLA A3 supermotif-bearing sequences are then synthesized and tested for binding to HLA-A*0301 and HLA-A*1101 molecules, the molecules encoded by the two most prevalent A3-supertype alleles. The peptides that bind at least one of the two alleles with binding affinities of \leq 500 nM, often \leq 200 nM, are then tested for binding cross-reactivity to the other common A3-supertype alleles (e.g., A*3101, A*3301, and A*6801) to identify those that can bind at least three of the five HLA-A3-supertype molecules tested.

Selection of HLA-B7 supermotif bearing epitopes

The 101P3A11 protein is also analyzed for the presence of 8-, 9- 10-, or 11-mer peptides with the HLA-B7-supermotif. Corresponding peptides are synthesized and tested for binding to HLA-B*0702, the molecule encoded by the most common B7-supertype allele (i.e., the prototype B7 supertype allele). Peptides binding B*0702 with IC₅₀ of ≤500 nM are identified using standard methods. These peptides are then tested for binding to other common B7-supertype molecules (e.g., B*3501, B*5101, B*5301, and B*5401). Peptides capable of binding to three or more of the five B7-supertype alleles tested are thereby identified.

Selection of A1 and A24 motif-bearing epitopes

To further increase population coverage, HLA-A1 and -A24 epitopes can also be incorporated into vaccine compositions. An analysis of the 101P3A11 protein can also be performed to identify HLA-A1- and A24-motif-containing sequences.

High affinity and/or cross-reactive binding epitopes that bear other motif and/or supermotifs are identified using analogous methodology.

Example 9: Confirmation of Immunogenicity

Cross-reactive candidate CTL A2-supermotif-bearing peptides that are identified as described herein are selected to confirm *in vitro* immunogenicity. Confirmation is performed using the following methodology:

Target Cell Lines for Cellular Screening:

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The .221A2.1 cell line, produced by transferring the HLA-A2.1 gene into the HLA-A, -B, -C null mutant human B-lymphoblastoid cell line 721.221, is used as the peptide-loaded target to measure activity of HLA-A2.1-restricted CTL. This cell line is grown in RPMI-1640 medium supplemented with antibiotics, sodium pyruvate, nonessential amino acids and 10% (v/v) heat inactivated FCS. Cells that express an antigen of interest, or transfectants comprising the gene encoding the antigen of interest, can be used as target cells to confirm the ability of peptide-specific CTLs to recognize endogenous antigen.

Primary CTL Induction Cultures:

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Generation of Dendritic Cells (DC): PBMCs are thawed in RPMI with 30 μg/ml DNAse, washed twice and resuspended in complete medium (RPMI-1640 plus 5% AB human serum, non-essential amino acids, sodium pyruvate, L-glutamine and penicillin/streptomycin). The monocytes are purified by plating 10 x 10⁶ PBMC/well in a 6-well plate. After 2 hours at 37°C, the non-adherent cells are removed by gently shaking the plates and aspirating the supernatants. The wells are washed a total of three times with 3 ml RPMI to remove most of the non-adherent and loosely adherent cells. Three ml of complete medium containing 50 ng/ml of GM-CSF and 1,000 U/ml of IL-4 are then added to each well. TNFα is added to the DCs on day 6 at 75 ng/ml and the cells are used for CTL induction cultures on day 7.

Induction of CTL with DC and Peptide: CD8+ T-cells are isolated by positive selection with Dynal immunomagnetic beads (Dynabeads® M-450) and the detacha-bead® reagent. Typically about 200-250x10⁶ PBMC are processed to obtain 24x10⁶ CD8⁺ T-cells (enough for a 48-well plate culture). Briefly, the PBMCs are thawed in RPMI with 30μg/ml DNAse, washed once with PBS containing 1% human AB serum and resuspended in PBS/1% AB serum at a concentration of 20x10⁶ cells/ml. The magnetic beads are washed 3 times with PBS/AB serum, added to the cells (140μl beads/20x10⁶ cells) and incubated for 1 hour at 4°C with continuous mixing. The beads and cells are washed 4x with PBS/AB serum to remove the nonadherent cells and resuspended at 100x10⁶ cells/ml (based on the original cell number) in PBS/AB serum containing 100μl/ml detacha-bead® reagent and 30 μg/ml DNAse. The mixture is incubated for 1 hour at room temperature with continuous mixing. The beads are washed again with PBS/AB/DNAse to collect the CD8+ T-cells. The DC are collected and centrifuged at 1300 rpm for 5-7 minutes, washed once with PBS with 1% BSA, counted and pulsed with 40μg/ml of peptide at a cell concentration of 1-2x10⁶/ml in the presence of 3μg/ml β₂- microglobulin for 4 hours at 20°C. The DC are then irradiated (4,200 rads), washed 1 time with medium and counted again.

Setting up induction cultures: 0.25 ml cytokine-generated DC (at 1x10⁵ cells/ml) are co-cultured with 0.25ml of CD8+ T-cells (at 2x10⁶ cell/ml) in each well of a 48-well plate in the presence of 10 ng/ml of IL-7. Recombinant human IL-10 is added the next day at a final concentration of 10 ng/ml and rhuman IL-2 is added 48 hours later at 10 IU/ml.

Restimulation of the induction cultures with peptide-pulsed adherent cells: Seven and fourteen days after the primary induction, the cells are restimulated with peptide-pulsed adherent cells. The PBMCs are thawed and washed twice with RPMI and DNAse. The cells are resuspended at $5x10^6$ cells/ml and irradiated at ~4200 rads. The PBMCs are plated at $2x10^6$ in 0.5 ml complete medium per well and incubated for 2 hours at 37°C. The plates are washed twice with RPMI by tapping the plate gently to remove the nonadherent cells and the adherent cells pulsed with $10\mu g/ml$ of peptide in the presence of $3\mu g/ml$ β_2 microglobulin in 0.25ml RPMI/5%AB per well for 2 hours at 37°C. Peptide

solution from each well is aspirated and the wells are washed once with RPMI. Most of the media is aspirated from the induction cultures (CD8+ cells) and brought to 0.5 ml with fresh media. The cells are then transferred to the wells containing the peptide-pulsed adherent cells. Twenty four hours later recombinant human IL-10 is added at a final concentration of 10 ng/ml and recombinant human IL2 is added the next day and again 2-3 days later at 50IU/ml (Tsai et al., Critical Reviews in Immunology 18(1-2):65-75, 1998). Seven days later, the cultures are assayed for CTL activity in a ⁵¹Cr release assay. In some experiments the cultures are assayed for peptide-specific recognition in the *in situ* IFNy ELISA at the time of the second restimulation followed by assay of endogenous recognition 7 days later. After expansion, activity is measured in both assays for a side-by-side comparison.

Measurement of CTL lytic activity by ⁵¹Cr release.

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Seven days after the second restimulation, cytotoxicity is determined in a standard (5 hr) ⁵¹Cr release assay by assaying individual wells at a single E:T. Peptide-pulsed targets are prepared by incubating the cells with 10µg/ml peptide overnight at 37°C.

Adherent target cells are removed from culture flasks with trypsin-EDTA. Target cells are labeled with 200µCi of ⁵¹Cr sodium chromate (Dupont, Wilmington, DE) for 1 hour at 37°C. Labeled target cells are resuspended at 10⁶ per ml and diluted 1:10 with K562 cells at a concentration of 3.3x10⁶/ml (an NK-sensitive erythroblastoma cell line used to reduce non-specific lysis). Target cells (100 µl) and effectors (100µl) are plated in 96 well round-bottom plates and incubated for 5 hours at 37°C. At that time, 100 µl of supernatant are collected from each well and percent lysis is determined according to the formula:

[(cpm of the test sample- cpm of the spontaneous ⁵¹Cr release sample)/(cpm of the maximal ⁵¹Cr release sample- cpm of the spontaneous ⁵¹Cr release sample)] x 100.

Maximum and spontaneous release are determined by incubating the labeled targets with 1% Trition X-100 and media alone, respectively. A positive culture is defined as one in which the specific lysis (sample-background) is 10% or higher in the case of individual wells and is 15% or more at the two highest E:T ratios when expanded cultures are assayed.

In situ Measurement of Human IFNy Production as an Indicator of Peptide-specific and Endogenous Recognition

Immulon 2 plates are coated with mouse anti-human IFN γ monoclonal antibody (4 µg/ml 0.1M NaHCO₃, pH8.2) overnight at 4°C. The plates are washed with Ca²⁺, Mg²⁺-free PBS/0.05% Tween 20 and blocked with PBS/10% FCS for two hours, after which the CTLs (100 µl/well) and targets (100 µl/well) are added to each well, leaving empty wells for the standards and blanks (which received media only). The target cells, either peptide-pulsed or endogenous targets, are used at a concentration of 1×10^6 cells/ml. The plates are incubated for 48 hours at 37°C with 5% CO₂.

Recombinant human IFN-gamma is added to the standard wells starting at 400 pg or 1200pg/100 microliter/well and the plate incubated for two hours at 37°C. The plates are washed and 100 µl of biotinylated mouse anti-human IFN-gamma monoclonal antibody (2 microgram/ml in PBS/3%FCS/0.05% Tween 20) are added and incubated for 2 hours at room temperature. After washing again, 100 microliter HRP-streptavidin (1:4000) are added and the plates incubated for one hour at room temperature. The plates are then washed 6x with wash buffer, 100 microliter/well developing solution (TMB 1:1) are added, and the plates allowed to develop for 5-15 minutes. The

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reaction is stopped with 50 microliter/well 1M H₃PO₄ and read at OD450. A culture is considered positive if it measured at least 50 pg of IFN-gamma/well above background and is twice the background level of expression.

CTL Expansion.

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Those cultures that demonstrate specific lytic activity against peptide-pulsed targets and/or tumor targets are expanded over a two week period with anti-CD3. Briefly, 5x10⁴ CD8+ cells are added to a T25 flask containing the following: 1x10⁶ irradiated (4,200 rad) PBMC (autologous or allogeneic) per ml, 2x10⁵ irradiated (8,000 rad) EBV-transformed cells per ml, and OKT3 (anti-CD3) at 30ng per ml in RPMI-1640 containing 10% (v/v) human AB serum, non-essential amino acids, sodium pyruvate, 25μM 2-mercaptoethanol, L-glutamine and penicillin/streptomycin. Recombinant human IL2 is added 24 hours later at a final concentration of 2001U/ml and every three days thereafter with fresh media at 50IU/ml. The cells are split if the cell concentration exceeds 1x10⁶/ml and the cultures are assayed between days 13 and 15 at E:T ratios of 30, 10, 3 and 1:1 in the ⁵¹Cr release assay or at 1x10⁶/ml in the *in situ* IFNγ assay using the same targets as before the expansion.

Cultures are expanded in the absence of anti-CD3⁺ as follows. Those cultures that demonstrate specific lytic activity against peptide and endogenous targets are selected and $5x10^4$ CD8⁺ cells are added to a T25 flask containing the following: $1x10^6$ autologous PBMC per ml which have been peptide-pulsed with 10 µg/ml peptide for two hours at 37°C and irradiated (4,200 rad); $2x10^5$ irradiated (8,000 rad) EBV-transformed cells per ml RPMI-1640 containing 10%(v/v) human AB serum, non-essential AA, sodium pyruvate, 25mM 2-ME, L-glutamine and gentamicin.

Immunogenicity of A2 supermotif-bearing peptides

A2-supermotif cross-reactive binding peptides are tested in the cellular assay for the ability to induce peptide-specific CTL in normal individuals. In this analysis, a peptide is typically considered to be an epitope if it induces peptide-specific CTLs in at least individuals, and preferably, also recognizes the endogenously expressed peptide.

Immunogenicity can also be confirmed using PBMCs isolated from patients bearing a tumor that expresses 101P3A11. Briefly, PBMCs are isolated from patients, re-stimulated with peptide-pulsed monocytes and assayed for the ability to recognize peptide-pulsed target cells as well as transfected cells endogenously expressing the antigen.

Evaluation of A*03/A11 immunogenicity

HLA-A3 supermotif-bearing cross-reactive binding peptides are also evaluated for immunogenicity using methodology analogous for that used to evaluate the immunogenicity of the HLA-A2 supermotif peptides.

Evaluation of B7 immunogenicity

Immunogenicity screening of the B7-supertype cross-reactive binding peptides identified as set forth herein are confirmed in a manner analogous to the confirmation of A2-and A3-supermotif-bearing peptides.

Peptides bearing other supermotifs/motifs, e.g., HLA-A1, HLA-A24 etc. are also confirmed using similar methodology

Example 10: Implementation of the Extended Supermotif to Improve the Binding Capacity of Native Epitopes by Creating Analogs

HLA motifs and supermotifs (comprising primary and/or secondary residues) are useful in the identification and preparation of highly cross-reactive native peptides, as demonstrated herein. Moreover, the definition of HLA motifs and supermotifs also allows one to engineer highly cross-reactive epitopes by identifying residues within a native

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peptide sequence which can be analoged to confer upon the peptide certain characteristics, e.g. greater cross-reactivity within the group of HLA molecules that comprise a supertype, and/or greater binding affinity for some or all of those HLA molecules. Examples of analoging peptides to exhibit modulated binding affinity are set forth in this example.

Analoging at Primary Anchor Residues

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Peptide engineering strategies are implemented to further increase the cross-reactivity of the epitopes. For example, the main anchors of A2-supermotif-bearing peptides are altered, for example, to introduce a preferred L, I, V, or M at position 2, and I or V at the C-terminus.

To analyze the cross-reactivity of the analog peptides, each engineered analog is initially tested for binding to the prototype A2 supertype allele A*0201, then, if A*0201 binding capacity is maintained, for A2-supertype cross-reactivity.

Alternatively, a peptide is confirmed as binding one or all supertype members and then analogued to modulate binding affinity to any one (or more) of the supertype members to add population coverage.

The selection of analogs for immunogenicity in a cellular screening analysis is typically further restricted by the capacity of the parent wild type (WT) peptide to bind at least weakly, i.e., bind at an IC₅₀ of 5000nM or less, to three of more A2 supertype alleles. The rationale for this requirement is that the WT peptides must be present endogenously in sufficient quantity to be biologically relevant. Analoged peptides have been shown to have increased immunogenicity and cross-reactivity by T cells specific for the parent epitope (see, e.g., Parkhurst et al., J. Immunol. 157:2539, 1996; and Pogue et al., Proc. Natl. Acad. Sci. USA 92:8166, 1995).

In the cellular screening of these peptide analogs, it is important to confirm that analog-specific CTLs are also able to recognize the wild-type peptide and, when possible, target cells that endogenously express the epitope.

Analoging of HLA-A3 and B7-supermotif-bearing peptides

Analogs of HLA-A3 supermotif-bearing epitopes are generated using strategies similar to those employed in analoging HLA-A2 supermotif-bearing peptides. For example, peptides binding to 3/5 of the A3-supertype molecules are engineered at primary anchor residues to possess a preferred residue (V, S, M, or A) at position 2.

The analog peptides are then tested for the ability to bind A*03 and A*11 (prototype A3 supertype alleles). Those peptides that demonstrate ≤ 500 nM binding capacity are then confirmed as having A3-supertype cross-reactivity.

Similarly to the A2- and A3- motif bearing peptides, peptides binding 3 or more B7-supertype alleles can be improved, where possible, to achieve increased cross-reactive binding or greater binding affinity or binding half life. B7 supermotif-bearing peptides are, for example, engineered to possess a preferred residue (V, I, L, or F) at the C-terminal primary anchor position, as demonstrated by Sidney *et al.* (*J. Immunol.* 157:3480-3490, 1996).

Analoging at primary anchor residues of other motif and/or supermotif-bearing epitopes is performed in a like manner.

The analog peptides are then be confirmed for immunogenicity, typically in a cellular screening assay. Again, it is generally important to demonstrate that analog-specific CTLs are also able to recognize the wild-type peptide and, when possible, targets that endogenously express the epitope.

Analoging at Secondary Anchor Residues

Moreover, HLA supermotifs are of value in engineering highly cross-reactive peptides and/or peptides that bind HLA molecules with increased affinity by identifying particular residues at secondary anchor positions that are

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associated with such properties. For example, the binding capacity of a B7 supermotif-bearing peptide with an F residue at position 1 is analyzed. The peptide is then analoged to, for example, substitute L for F at position 1. The analoged peptide is evaluated for increased binding affinity, binding half life and/or increased cross-reactivity. Such a procedure identifies analoged peptides with enhanced properties.

Engineered analogs with sufficiently improved binding capacity or cross-reactivity can also be tested for immunogenicity in HLA-B7-transgenic mice, following for example, IFA immunization or lipopeptide immunization. Analogued peptides are additionally tested for the ability to stimulate a recall response using PBMC from patients with 101P3A11-expressing tumors.

Other analoguing strategies

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Another form of peptide analoguing, unrelated to anchor positions, involves the substitution of a cysteine with α -amino butyric acid. Due to its chemical nature, cysteine has the propensity to form disulfide bridges and sufficiently alter the peptide structurally so as to reduce binding capacity. Substitution of α -amino butyric acid for cysteine not only alleviates this problem, but has been shown to improve binding and crossbinding capabilities in some instances (*see*, *e.g.*, the review by Sette *et al.*, In: Persistent Viral Infections, Eds. R. Ahmed and I. Chen, John Wiley & Sons, England, 1999).

Thus, by the use of single amino acid substitutions, the binding properties and/or cross-reactivity of peptide ligands for HLA supertype molecules can be modulated.

Example 11. Identification and confirmation of 101P3A11-derived sequences with HLA-DR binding motifs

Peptide epitopes bearing an HLA class II supermotif or motif are identified and confirmed as outlined below using methodology similar to that described for HLA Class I peptides.

Selection of HLA-DR-supermotif-bearing epitopes.

To identify 101P3A11-derived, HLA class II HTL epitopes, the 101P3A11 antigen is analyzed for the presence of sequences bearing an HLA-DR-motif or supermotif. Specifically, 15-mer sequences are selected comprising a DR-supermotif, comprising a 9-mer core, and three-residue N- and C-terminal flanking regions (15 amino acids total).

Protocols for predicting peptide binding to DR molecules have been developed (Southwood *et al.*, *J. Immunol*. 160:3363-3373, 1998). These protocols, specific for individual DR molecules, allow the scoring, and ranking, of 9-mer core regions. Each protocol not only scores peptide sequences for the presence of DR-supermotif primary anchors (i.e., at position 1 and position 6) within a 9-mer core, but additionally evaluates sequences for the presence of secondary anchors. Using allele-specific selection tables (see, *e.g.*, Southwood *et al.*, *ibid.*), it has been found that these protocols efficiently select peptide sequences with a high probability of binding a particular DR molecule. Additionally, it has been found that performing these protocols in tandem, specifically those for DR1, DR4w4, and DR7, can efficiently select DR cross-reactive peptides.

The 101P3A11-derived peptides identified above are tested for their binding capacity for various common HLA-DR molecules. All peptides are initially tested for binding to the DR molecules in the primary panel: DR1, DR4w4, and DR7. Peptides binding at least two of these three DR molecules are then tested for binding to DR2w2 β 1, DR2w2 β 2, DR6w19, and DR9 molecules in secondary assays. Finally, peptides binding at least two of the four

secondary panel DR molecules, and thus cumulatively at least four of seven different DR molecules, are screened for binding to DR4w15, DR5w11, and DR8w2 molecules in tertiary assays. Peptides binding at least seven of the ten DR molecules comprising the primary, secondary, and tertiary screening assays are considered cross-reactive DR binders. 101P3A11-derived peptides found to bind common HLA-DR alleles are of particular interest.

Selection of DR3 motif peptides

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Because HLA-DR3 is an allele that is prevalent in Caucasian, Black, and Hispanic populations, DR3 binding capacity is a relevant criterion in the selection of HTL epitopes. Thus, peptides shown to be candidates may also be assayed for their DR3 binding capacity. However, in view of the binding specificity of the DR3 motif, peptides binding only to DR3 can also be considered as candidates for inclusion in a vaccine formulation.

To efficiently identify peptides that bind DR3, target 101P3A11 antigens are analyzed for sequences carrying one of the two DR3-specific binding motifs reported by Geluk *et al.* (*J. Immunol.* 152:5742-5748, 1994). The corresponding peptides are then synthesized and confirmed as having the ability to bind DR3 with an affinity of 1μM or better, i.e., less than 1 μM. Peptides are found that meet this binding criterion and qualify as HLA class II high affinity binders.

DR3 binding epitopes identified in this manner are included in vaccine compositions with DR supermotifbearing peptide epitopes.

Similarly to the case of HLA class I motif-bearing peptides, the class II motif-bearing peptides are analoged to improve affinity or cross-reactivity. For example, aspartic acid at position 4 of the 9-mer core sequence is an optimal residue for DR3 binding, and substitution for that residue often improves DR 3 binding.

Example 12: Immunogenicity of 101P3A11-derived HTL epitopes

This example determines immunogenic DR supermotif- and DR3 motif-bearing epitopes among those identified using the methodology set forth herein.

Immunogenicity of HTL epitopes are confirmed in a manner analogous to the determination of immunogenicity of CTL epitopes, by assessing the ability to stimulate HTL responses and/or by using appropriate transgenic mouse models. Immunogenicity is determined by screening for: 1.) *in vitro* primary induction using normal PBMC or 2.) recall responses from patients who have 101P3A11-expressing tumors.

<u>Example 13: Calculation of phenotypic frequencies of HLA-supertypes in various ethnic backgrounds</u> <u>to determine breadth of population coverage</u>

This example illustrates the assessment of the breadth of population coverage of a vaccine composition comprised of multiple epitopes comprising multiple supermotifs and/or motifs.

In order to analyze population coverage, gene frequencies of HLA alleles are determined. Gene frequencies for each HLA allele are calculated from antigen or allele frequencies utilizing the binomial distribution formulae gf=1-(SQRT(1-af)) (see, e.g., Sidney et al., Human Immunol. 45:79-93, 1996). To obtain overall phenotypic frequencies, cumulative gene frequencies are calculated, and the cumulative antigen frequencies derived by the use of the inverse formula [af=1-(1-Cgf)²].

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Where frequency data is not available at the level of DNA typing, correspondence to the serologically defined antigen frequencies is assumed. To obtain total potential supertype population coverage no linkage disequilibrium is assumed, and only alleles confirmed to belong to each of the supertypes are included (minimal estimates). Estimates of total potential coverage achieved by inter-loci combinations are made by adding to the A coverage the proportion of the non-A covered population that could be expected to be covered by the B alleles considered (e.g., total=A+B*(1-A)). Confirmed members of the A3-like supertype are A3, A11, A31, A*3301, and A*6801. Although the A3-like supertype may also include A34, A66, and A*7401, these alleles were not included in overall frequency calculations. Likewise, confirmed members of the A2-like supertype family are A*0201, A*0202, A*0203, A*0204, A*0205, A*0206, A*0207, A*6802, and A*6901. Finally, the B7-like supertype-confirmed alleles are: B7, B*3501-03, B51, B*5301, B*5401, B*5501-2, B*5601, B*6701, and B*7801 (potentially also B*1401, B*3504-06, B*4201, and B*5602).

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Population coverage achieved by combining the A2-, A3- and B7-supertypes is approximately 86% in five major ethnic groups. Coverage may be extended by including peptides bearing the A1 and A24 motifs. On average, A1 is present in 12% and A24 in 29% of the population across five different major ethnic groups (Caucasian, North American Black, Chinese, Japanese, and Hispanic). Together, these alleles are represented with an average frequency of 39% in these same ethnic populations. The total coverage across the major ethnicities when A1 and A24 are combined with the coverage of the A2-, A3- and B7-supertype alleles is >95%. An analogous approach can be used to estimate population coverage achieved with combinations of class II motif-bearing epitopes.

Immunogenicity studies in humans (e.g., Bertoni et al., J. Clin. Invest. 100:503, 1997; Doolan et al., Immunity 7:97, 1997; and Threlkeld et al., J. Immunol. 159:1648, 1997) have shown that highly cross-reactive binding peptides are almost always recognized as epitopes. The use of highly cross-reactive binding peptides is an important selection criterion in identifying candidate epitopes for inclusion in a vaccine that is immunogenic in a diverse population.

With a sufficient number of epitopes (as disclosed herein and from the art), an average population coverage is predicted to be greater than 95% in each of five major ethnic populations. The game theory Monte Carlo simulation analysis, which is known in the art (see e.g., Osborne, M.J. and Rubinstein, A. "A course in game theory" MIT Press, 1994), can be used to estimate what percentage of the individuals in a population comprised of the Caucasian, North American Black, Japanese, Chinese, and Hispanic ethnic groups would recognize the vaccine epitopes described herein. A preferred percentage is 90%. A more preferred percentage is 95%.

Recombinant NIH3T3 cells stably expressing 101P3A11 (3T3/101P3A11-pSRa) were derived and were grown in SCID mice (Figure 20). Mice were injected subcutaneously with 3T3/101P3A11-pSRa cells. Tumors were allowed to grow, mice were then sacrificed and tumors harvested. RNA was isolated from LAPC-4AD and LAPC-4AI xenografts, 3T3/neo-pSRa and 3T3/101P3A11-pSRa grown in culture, as well as six different tumors derived from 3T3/101P3A11-pSRa (Tumor #1-3). Northern blots with 10 µg of total RNA/lane were probed with 101P3A11 sequence. Results show that 101P3A11 was expressed in all 3T3-101P3A11 tumors that grew following injection into mice.

Example 14: CTL Recognition Of Endogenously Processed Antigens After Priming

This example confirms that CTL induced by native or analoged peptide epitopes identified and selected as described herein recognize endogenously synthesized, *i.e.*, native antigens.

Effector cells isolated from transgenic mice that are immunized with peptide epitopes, for example HLA-A2 supermotif-bearing epitopes, are re-stimulated *in vitro* using peptide-coated stimulator cells. Six days later, effector cells are assayed for cytotoxicity and the cell lines that contain peptide-specific cytotoxic activity are further restimulated. An additional six days later, these cell lines are tested for cytotoxic activity on ⁵¹Cr labeled Jurkat-A2.1/K^b target cells in the absence or presence of peptide, and also tested on ⁵¹Cr labeled target cells bearing the endogenously synthesized antigen, *i.e.* cells that are stably transfected with 101P3A11 expression vectors.

The results demonstrate that CTL lines obtained from animals primed with peptide epitope recognize endogenously synthesized 101P3A11 antigen. The choice of transgenic mouse model to be used for such an analysis depends upon the epitope(s) that are being evaluated. In addition to HLA-A*0201/K^b transgenic mice, several other transgenic mouse models including mice with human A11, which may also be used to evaluate A3 epitopes, and B7 alleles have been characterized and others (e.g., transgenic mice for HLA-A1 and A24) are being developed. HLA-DR1 and HLA-DR3 mouse models have also been developed, which may be used to evaluate HTL epitopes.

Example 15: Activity Of CTL-HTL Conjugated Epitopes In Transgenic Mice

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This example illustrates the induction of CTLs and HTLs in transgenic mice, by use of a 101P3A11-derived CTL and HTL peptide vaccine compositions. The vaccine composition used herein comprise peptides to be administered to a patient with a 101P3A11-expressing tumor. The peptide composition can comprise multiple CTL and/or HTL epitopes. The epitopes are identified using methodology as described herein. This example also illustrates that enhanced immunogenicity can be achieved by inclusion of one or more HTL epitopes in a CTL vaccine composition; such a peptide composition can comprise an HTL epitope conjugated to a CTL epitope. The CTL epitope can be one that binds to multiple HLA family members at an affinity of 500 nM or less, or analogs of that epitope. The peptides may be lipidated, if desired.

Immunization procedures: Immunization of transgenic mice is performed as described (Alexander et al., J. Immunol. 159:4753-4761, 1997). For example, A2/K^b mice, which are transgenic for the human HLA A2.1 allele and are used to confirm the immunogenicity of HLA-A*0201 motif- or HLA-A2 supermotif-bearing epitopes, and are primed subcutaneously (base of the tail) with a 0.1 ml of peptide in Incomplete Freund's Adjuvant, or if the peptide composition is a lipidated CTL/HTL conjugate, in DMSO/saline, or if the peptide composition is a polypeptide, in PBS or Incomplete Freund's Adjuvant. Seven days after priming, splenocytes obtained from these animals are restimulated with syngenic irradiated LPS-activated lymphoblasts coated with peptide.

Cell lines: Target cells for peptide-specific cytotoxicity assays are Jurkat cells transfected with the HLA-A2.1/Kb chimeric gene (e.g., Vitiello et al., J. Exp. Med. 173:1007, 1991)

<u>In vitro</u> CTL activation: One week after priming, spleen cells (30x10⁶ cells/flask) are co-cultured at 37°C with syngeneic, irradiated (3000 rads), peptide coated lymphoblasts (10x10⁶ cells/flask) in 10 ml of culture medium/T25 flask. After six days, effector cells are harvested and assayed for cytotoxic activity.

Assay for cytotoxic activity: Target cells (1.0 to 1.5×10^6) are incubated at 37°C in the presence of 200 μ l of 51 Cr. After 60 minutes, cells are washed three times and resuspended in R10 medium. Peptide is added where required at a concentration of 1 μ g/ml. For the assay, $10^{4.51}$ Cr-labeled target cells are added to different concentrations of effector cells (final volume of 200 μ l) in U-bottom 96-well plates. After a six hour incubation period at 37°C, a 0.1 ml

aliquot of supernatant is removed from each well and radioactivity is determined in a Micromedic automatic gamma counter. The percent specific lysis is determined by the formula: percent specific release = 100 x (experimental release - spontaneous release)/(maximum release - spontaneous release). To facilitate comparison between separate CTL assays run under the same conditions, % 51 Cr release data is expressed as lytic units/ 10^6 cells. One lytic unit is arbitrarily defined as the number of effector cells required to achieve 30% lysis of 10,000 target cells in a six hour 51 Cr release assay. To obtain specific lytic units/ 10^6 , the lytic units/ 10^6 obtained in the absence of peptide is subtracted from the lytic units/ 10^6 obtained in the presence of peptide. For example, if 30% 51 Cr release is obtained at the effector (E): target (T) ratio of 50:1 (i.e., $5x10^5$ effector cells for 10,000 targets) in the absence of peptide and 5:1 (i.e., $5x10^4$ effector cells for 10,000 targets) in the presence of peptide, the specific lytic units would be: $[(1/50,000)-(1/500,000)] \times 10^6 = 18$ LU.

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The results are analyzed to assess the magnitude of the CTL responses of animals injected with the immunogenic CTL/HTL conjugate vaccine preparation and are compared to the magnitude of the CTL response achieved using, for example, CTL epitopes as outlined above in the Example entitled "Confirmation of Immunogenicity". Analyses similar to this may be performed to confirm the immunogenicity of peptide conjugates containing multiple CTL epitopes and/or multiple HTL epitopes. In accordance with these procedures, it is found that a CTL response is induced, and concomitantly that an HTL response is induced upon administration of such compositions.

Example 16: Selection of CTL and HTL epitopes for inclusion in an 101P3A11-specific vaccine.

This example illustrates a procedure for selecting peptide epitopes for vaccine compositions of the invention. The peptides in the composition can be in the form of a nucleic acid sequence, either single or one or more sequences (i.e., minigene) that encodes peptide(s), or can be single and/or polyepitopic peptides.

The following principles are utilized when selecting a plurality of epitopes for inclusion in a vaccine composition. Each of the following principles is balanced in order to make the selection.

Epitopes are selected which, upon administration, mimic immune responses that are correlated with 101P3A11 clearance. The number of epitopes used depends on observations of patients who spontaneously clear 101P3A11. For example, if it has been observed that patients who spontaneously clear 101P3A11 generate an immune response to at least three (3) from 101P3A11 antigen, then three or four (3-4) epitopes should be included for HLA class I. A similar rationale is used to determine HLA class II epitopes.

Epitopes are often selected that have a binding affinity of an IC_{50} of 500 nM or less for an HLA class I molecule, or for class II, an IC_{50} of 1000 nM or less; or HLA Class I peptides with high binding scores from the BIMAS web site, at URL <u>bimas.dcrt.nih.gov/</u>.

In order to achieve broad coverage of the vaccine through out a diverse population, sufficient supermotif bearing peptides, or a sufficient array of allele-specific motif bearing peptides, are selected to give broad population coverage. In one embodiment, epitopes are selected to provide at least 80% population coverage. A Monte Carlo analysis, a statistical evaluation known in the art, can be employed to assess breadth, or redundancy, of population coverage.

When creating polyepitopic compositions, or a minigene that encodes same, it is typically desirable to generate the smallest peptide possible that encompasses the epitopes of interest. The principles employed are similar, if not the same, as those employed when selecting a peptide comprising nested epitopes. For example, a protein sequence for the vaccine composition is selected because it has maximal number of epitopes contained within the sequence, i.e., it has a high concentration of epitopes. Epitopes may be nested or overlapping (i.e., frame shifted relative to one another). For example, with overlapping epitopes, two 9-mer epitopes and one 10-mer epitope can be present in a 10 amino acid peptide. Each epitope can be exposed and bound by an HLA molecule upon administration of such a peptide. A multiepitopic, peptide can be generated synthetically, recombinantly, or via cleavage from the native source. Alternatively, an analog can be made of this native sequence, whereby one or more of the epitopes comprise substitutions that alter the cross-reactivity and/or binding affinity properties of the polyepitopic peptide. Such a vaccine composition is administered for the apeutic or prophylactic purposes. This embodiment provides for the possibility that an as yet undiscovered aspect of immune system processing will apply to the native nested sequence and thereby facilitate the production of therapeutic or prophylactic immune response-inducing vaccine compositions. Additionally such an embodiment provides for the possibility of motif-bearing epitopes for an HLA makeup that is presently unknown. Furthermore, this embodiment (absent the creating of any analogs) directs the immune response to multiple peptide sequences that are actually present in 101P3A11, thus avoiding the need to evaluate any junctional epitopes. Lastly, the embodiment provides an economy of scale when producing nucleic acid vaccine compositions. Related to this embodiment, computer programs can be derived in accordance with principles in the art, which identify in a target sequence, the greatest number of epitopes per sequence length.

A vaccine composition comprised of selected peptides, when administered, is safe, efficacious, and elicits an immune response similar in magnitude to an immune response that controls or clears cells that bear or overexpress 101P3A11.

Example 17: Construction of "Minigene" Multi-Epitope DNA Plasmids

This example discusses the construction of a minigene expression plasmid. Minigene plasmids may, of course, contain various configurations of B cell, CTL and/or HTL epitopes or epitope analogs as described herein.

A minigene expression plasmid typically includes multiple CTL and HTL peptide epitopes. In the present example, HLA-A2, -A3, -B7 supermotif-bearing peptide epitopes and HLA-A1 and -A24 motif-bearing peptide epitopes are used in conjunction with DR supermotif-bearing epitopes and/or DR3 epitopes. HLA class I supermotif or motif-bearing peptide epitopes derived 101P3A11, are selected such that multiple supermotifs/motifs are represented to ensure broad population coverage. Similarly, HLA class II epitopes are selected from 101P3A11 to provide broad population coverage, *i.e.* both HLA DR-1-4-7 supermotif-bearing epitopes and HLA DR-3 motif-bearing epitopes are selected for inclusion in the minigene construct. The selected CTL and HTL epitopes are then incorporated into a minigene for expression in an expression vector.

Such a construct may additionally include sequences that direct the HTL epitopes to the endoplasmic reticulum. For example, the Ii protein may be fused to one or more HTL epitopes as described in the art, wherein the CLIP sequence of the Ii protein is removed and replaced with an HLA class II epitope sequence so that HLA class II epitope is directed to the endoplasmic reticulum, where the epitope binds to an HLA class II molecules.

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This example illustrates the methods to be used for construction of a minigene-bearing expression plasmid.

Other expression vectors that may be used for minigene compositions are available and known to those of skill in the art.

The minigene DNA plasmid of this example contains a consensus Kozak sequence and a consensus murine kappa Ig-light chain signal sequence followed by CTL and/or HTL epitopes selected in accordance with principles disclosed herein. The sequence encodes an open reading frame fused to the Myc and His antibody epitope tag coded for by the pcDNA 3.1 Myc-His vector.

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Overlapping oligonucleotides that can, for example, average about 70 nucleotides in length with 15 nucleotide overlaps, are synthesized and HPLC-purified. The oligonucleotides encode the selected peptide epitopes as well as appropriate linker nucleotides, Kozak sequence, and signal sequence. The final multiepitope minigene is assembled by extending the overlapping oligonucleotides in three sets of reactions using PCR. A Perkin/Elmer 9600 PCR machine is used and a total of 30 cycles are performed using the following conditions: 95°C for 15 sec, annealing temperature (5° below the lowest calculated Tm of each primer pair) for 30 sec, and 72°C for 1 min.

For example, a minigene is prepared as follows. For a first PCR reaction, 5 µg of each of two oligonucleotides are annealed and extended: In an example using eight oligonucleotides, i.e., four pairs of primers, oligonucleotides 1+2, 3+4, 5+6, and 7+8 are combined in 100 µl reactions containing *Pfu* polymerase buffer (1x= 10 mM KCL, 10 mM (NH4)₂SO₄, 20 mM Tris-chloride, pH 8.75, 2 mM MgSO₄, 0.1% Triton X-100, 100 µg/ml BSA), 0.25 mM each dNTP, and 2.5 U of *Pfu* polymerase. The full-length dimer products are gel-purified, and two reactions containing the product of 1+2 and 3+4, and the product of 5+6 and 7+8 are mixed, annealed, and extended for 10 cycles. Half of the two reactions are then mixed, and 5 cycles of annealing and extension carried out before flanking primers are added to amplify the full length product. The full-length product is gel-purified and cloned into pCR-blunt (Invitrogen) and individual clones are screened by sequencing.

Example 18: The Plasmid Construct and the Degree to Which It Induces Immunogenicity.

The degree to which a plasmid construct, for example a plasmid constructed in accordance with the previous Example, is able to induce immunogenicity is confirmed *in vitro* by determining epitope presentation by APC following transduction or transfection of the APC with an epitope-expressing nucleic acid construct. Such a study determines "antigenicity" and allows the use of human APC. The assay determines the ability of the epitope to be presented by the APC in a context that is recognized by a T cell by quantifying the density of epitope-HLA class I complexes on the cell surface. Quantitation can be performed by directly measuring the amount of peptide eluted from the APC (*see*, *e.g.*, Sijts *et al.*, *J. Immunol.* 156:683-692, 1996; Demotz *et al.*, *Nature* 342:682-684, 1989); or the number of peptide-HLA class I complexes can be estimated by measuring the amount of lysis or lymphokine release induced by diseased or transfected target cells, and then determining the concentration of peptide necessary to obtain equivalent levels of lysis or lymphokine release (*see*, *e.g.*, Kageyama *et al.*, *J. Immunol.* 154:567-576, 1995).

Alternatively, immunogenicity is confirmed through *in vivo* injections into mice and subsequent *in vitro* assessment of CTL and HTL activity, which are analyzed using cytotoxicity and proliferation assays, respectively, as detailed *e.g.*, in Alexander *et al.*, *Immunity* 1:751-761, 1994.

For example, to confirm the capacity of a DNA minigene construct containing at least one HLA-A2 supermotif peptide to induce CTLs *in vivo*, HLA-A2.1/K^b transgenic mice, for example, are immunized intramuscularly with 100 µg of naked cDNA. As a means of comparing the level of CTLs induced by cDNA immunization, a control group of animals is also immunized with an actual peptide composition that comprises multiple epitopes synthesized as a single polypeptide as they would be encoded by the minigene.

Splenocytes from immunized animals are stimulated twice with each of the respective compositions (peptide epitopes encoded in the minigene or the polyepitopic peptide), then assayed for peptide-specific cytotoxic activity in a ⁵¹Cr release assay. The results indicate the magnitude of the CTL response directed against the A2-restricted epitope, thus indicating the *in vivo* immunogenicity of the minigene vaccine and polyepitopic vaccine.

It is, therefore, found that the minigene elicits immune responses directed toward the HLA-A2 supermotif peptide epitopes as does the polyepitopic peptide vaccine. A similar analysis is also performed using other HLA-A3 and HLA-B7 transgenic mouse models to assess CTL induction by HLA-A3 and HLA-B7 motif or supermotif epitopes, whereby it is also found that the minigene elicits appropriate immune responses directed toward the provided epitopes.

To confirm the capacity of a class II epitope-encoding minigene to induce HTLs *in vivo*, DR transgenic mice, or for those epitopes that cross react with the appropriate mouse MHC molecule, I-A^b-restricted mice, for example, are immunized intramuscularly with 100 µg of plasmid DNA. As a means of comparing the level of HTLs induced by DNA immunization, a group of control animals is also immunized with an actual peptide composition emulsified in complete Freund's adjuvant. CD4+ T cells, *i.e.* HTLs, are purified from splenocytes of immunized animals and stimulated with each of the respective compositions (peptides encoded in the minigene). The HTL response is measured using a ³H-thymidine incorporation proliferation assay, (*see*, *e.g.*, Alexander et al. Immunity 1:751-761, 1994). The results indicate the magnitude of the HTL response, thus demonstrating the *in vivo* immunogenicity of the minigene.

DNA minigenes, constructed as described in the previous Example, can also be confirmed as a vaccine in combination with a boosting agent using a prime boost protocol. The boosting agent can consist of recombinant protein (e.g., Barnett et al., Aids Res. and Human Retroviruses 14, Supplement 3:S299-S309, 1998) or recombinant vaccinia, for example, expressing a minigene or DNA encoding the complete protein of interest (see, e.g., Hanke et al., Vaccine 16:439-445, 1998; Sedegah et al., Proc. Natl. Acad. Sci USA 95:7648-53, 1998; Hanke and McMichael, Immunol. Letters 66:177-181, 1999; and Robinson et al., Nature Med. 5:526-34, 1999).

For example, the efficacy of the DNA minigene used in a prime boost protocol is initially evaluated in transgenic mice. In this example, A2.1/K^b transgenic mice are immunized IM with 100 µg of a DNA minigene encoding the immunogenic peptides including at least one HLA-A2 supermotif-bearing peptide. After an incubation period (ranging from 3-9 weeks), the mice are boosted IP with 10⁷ pfu/mouse of a recombinant vaccinia virus expressing the same sequence encoded by the DNA minigene. Control mice are immunized with 100 µg of DNA or recombinant vaccinia without the minigene sequence, or with DNA encoding the minigene, but without the vaccinia boost. After an additional incubation period of two weeks, splenocytes from the mice are immediately assayed for peptide-specific activity in an ELISPOT assay. Additionally, splenocytes are stimulated *in vitro* with the A2-restricted peptide epitopes encoded in the minigene and recombinant vaccinia, then assayed for peptide-specific activity in an alpha, beta and/or gamma IFN ELISA.

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It is found that the minigene utilized in a prime-boost protocol elicits greater immune responses toward the HLA-A2 supermotif peptides than with DNA alone. Such an analysis can also be performed using HLA-A11 or HLA-B7 transgenic mouse models to assess CTL induction by HLA-A3 or HLA-B7 motif or supermotif epitopes. The use of prime boost protocols in humans is described below in the Example entitled "Induction of CTL Responses Using a Prime Boost Protocol."

Example 19: Peptide Composition for Prophylactic Uses

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Vaccine compositions of the present invention can be used to prevent 101P3A11 expression in persons who are at risk for tumors that bear this antigen. For example, a polyepitopic peptide epitope composition (or a nucleic acid comprising the same) containing multiple CTL and HTL epitopes such as those selected in the above Examples, which are also selected to target greater than 80% of the population, is administered to individuals at risk for a 101P3A11-associated tumor.

For example, a peptide-based composition is provided as a single polypeptide that encompasses multiple epitopes. The vaccine is typically administered in a physiological solution that comprises an adjuvant, such as Incomplete Freund's Adjuvant. The dose of peptide for the initial immunization is from about 1 to about 50,000 μg, generally 100-5,000 μg, for a 70 kg patient. The initial administration of vaccine is followed by booster dosages at 4 weeks followed by evaluation of the magnitude of the immune response in the patient, by techniques that determine the presence of epitope-specific CTL populations in a PBMC sample. Additional booster doses are administered as required. The composition is found to be both safe and efficacious as a prophylaxis against 101P3A11-associated disease.

Alternatively, a composition typically comprising transfecting agents is used for the administration of a nucleic acid-based vaccine in accordance with methodologies known in the art and disclosed herein.

Example 20: Polycpitopic Vaccine Compositions Derived from Native 101P3A11 Sequences

A native 101P3A11 polyprotein sequence is analyzed, preferably using computer algorithms defined for each class I and/or class II supermotif or motif, to identify "relatively short" regions of the polyprotein that comprise multiple epitopes. The "relatively short" regions are preferably less in length than an entire native antigen. This relatively short sequence that contains multiple distinct or overlapping, "nested" epitopes is selected; it can be used to generate a minigene construct. The construct is engineered to express the peptide, which corresponds to the native protein sequence. The "relatively short" peptide is generally less than 250 amino acids in length, often less than 100 amino acids in length, preferably less than 75 amino acids in length, and more preferably less than 50 amino acids in length. The protein sequence of the vaccine composition is selected because it has maximal number of epitopes contained within the sequence, *i.e.*, it has a high concentration of epitopes. As noted herein, epitope motifs may be nested or overlapping (*i.e.*, frame shifted relative to one another). For example, with overlapping epitopes, two 9-mer epitopes and one 10-mer epitope can be present in a 10 amino acid peptide. Such a vaccine composition is administered for therapeutic or prophylactic purposes.

The vaccine composition will include, for example, multiple CTL epitopes from 101P3A11 antigen and at least one HTL epitope. This polyepitopic native sequence is administered either as a peptide or as a nucleic acid sequence

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which encodes the peptide. Alternatively, an analog can be made of this native sequence, whereby one or more of the epitopes comprise substitutions that alter the cross-reactivity and/or binding affinity properties of the polyepitopic peptide.

The embodiment of this example provides for the possibility that an as yet undiscovered aspect of immune system processing will apply to the native nested sequence and thereby facilitate the production of therapeutic or prophylactic immune response-inducing vaccine compositions. Additionally such an embodiment provides for the possibility of motif-bearing epitopes for an HLA makeup that is presently unknown. Furthermore, this embodiment (excluding an analoged embodiment) directs the immune response to multiple peptide sequences that are actually present in native 101P3A11, thus avoiding the need to evaluate any junctional epitopes. Lastly, the embodiment provides an economy of scale when producing peptide or nucleic acid vaccine compositions.

Related to this embodiment, computer programs are available in the art which can be used to identify in a target sequence, the greatest number of epitopes per sequence length.

Example 21: Polyepitopic Vaccine Compositions From Multiple Antigens

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The 101P3A11 peptide epitopes of the present invention are used in conjunction with epitopes from other target tumor-associated antigens, to create a vaccine composition that is useful for the prevention or treatment of cancer that expresses 101P3A11 and such other antigens. For example, a vaccine composition can be provided as a single polypeptide that incorporates multiple epitopes from 101P3A11 as well as tumor-associated antigens that are often expressed with a target cancer associated with 101P3A11 expression, or can be administered as a composition comprising a cocktail of one or more discrete epitopes. Alternatively, the vaccine can be administered as a minigene construct or as dendritic cells which have been loaded with the peptide epitopes *in vitro*.

Example 22: Use of peptides to evaluate an immune response

Peptides of the invention may be used to analyze an immune response for the presence of specific antibodies, CTL or HTL directed to 101P3A11. Such an analysis can be performed in a manner described by Ogg *et al.*, *Science* 279:2103-2106, 1998. In this Example, peptides in accordance with the invention are used as a reagent for diagnostic or prognostic purposes, not as an immunogen.

In this example highly sensitive human leukocyte antigen tetrameric complexes ("tetramers") are used for a cross-sectional analysis of, for example, 101P3A11 HLA-A*0201-specific CTL frequencies from HLA A*0201-positive individuals at different stages of disease or following immunization comprising an 101P3A11 peptide containing an A*0201 motif. Tetrameric complexes are synthesized as described (Musey *et al., N. Engl. J. Med.* 337:1267, 1997). Briefly, purified HLA heavy chain (A*0201 in this example) and β2-microglobulin are synthesized by means of a prokaryotic expression system. The heavy chain is modified by deletion of the transmembrane-cytosolic tail and COOH-terminal addition of a sequence containing a BirA enzymatic biotinylation site. The heavy chain, β2-microglobulin, and peptide are refolded by dilution. The 45-kD refolded product is isolated by fast protein liquid chromatography and then biotinylated by BirA in the presence of biotin (Sigma, St. Louis, Missouri), adenosine 5' triphosphate and magnesium. Streptavidin-phycoerythrin conjugate is added in a 1:4 molar ratio, and the tetrameric product is concentrated to 1 mg/ml. The resulting product is referred to as tetramer-phycoerythrin.

For the analysis of patient blood samples, approximately one million PBMCs are centrifuged at 300g for 5 minutes and resuspended in 50 µl of cold phosphate-buffered saline. Tri-color analysis is performed with the tetramer-phycoerythrin, along with anti-CD8-Tricolor, and anti-CD38. The PBMCs are incubated with tetramer and antibodies on ice for 30 to 60 min and then washed twice before formaldehyde fixation. Gates are applied to contain >99.98% of control samples. Controls for the tetramers include both A*0201-negative individuals and A*0201-positive non-diseased donors. The percentage of cells stained with the tetramer is then determined by flow cytometry. The results indicate the number of cells in the PBMC sample that contain epitope-restricted CTLs, thereby readily indicating the extent of immune response to the 101P3A11 epitope, and thus the status of exposure to 101P3A11, or exposure to a vaccine that elicits a protective or therapeutic response.

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Example 23: Use of Peptide Epitopes to Evaluate Recall Responses

The peptide epitopes of the invention are used as reagents to evaluate T cell responses, such as acute or recall responses, in patients. Such an analysis may be performed on patients who have recovered from 101P3A11-associated disease or who have been vaccinated with an 101P3A11 vaccine.

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For example, the class I restricted CTL response of persons who have been vaccinated may be analyzed. The vaccine may be any 101P3A11 vaccine. PBMC are collected from vaccinated individuals and HLA typed. Appropriate peptide epitopes of the invention that, optimally, bear supermotifs to provide cross-reactivity with multiple HLA supertype family members, are then used for analysis of samples derived from individuals who bear that HLA type.

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PBMC from vaccinated individuals are separated on FicoII-Histopaque density gradients (Sigma Chemical Co., St. Louis, MO), washed three times in HBSS (GIBCO Laboratories), resuspended in RPMI-1640 (GIBCO Laboratories) supplemented with L-glutamine (2mM), penicillin (50U/ml), streptomycin (50 μg/ml), and Hepes (10mM) containing 10% heat-inactivated human AB serum (complete RPMI) and plated using microculture formats. A synthetic peptide comprising an epitope of the invention is added at 10 μg/ml to each well and HBV core 128-140 epitope is added at 1 μg/ml to each well as a source of T cell help during the first week of stimulation.

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In the microculture format, 4 x 10⁵ PBMC are stimulated with peptide in 8 replicate cultures in 96-well round bottom plate in 100 µl/well of complete RPMI. On days 3 and 10, 100 µl of complete RPMI and 20 U/ml final concentration of rIL-2 are added to each well. On day 7 the cultures are transferred into a 96-well flat-bottom plate and restimulated with peptide, rIL-2 and 10⁵ irradiated (3,000 rad) autologous feeder cells. The cultures are tested for cytotoxic activity on day 14. A positive CTL response requires two or more of the eight replicate cultures to display greater than 10% specific ⁵¹Cr release, based on comparison with non-diseased control subjects as previously described (Rehermann, et al., Nature Med. 2:1104,1108, 1996; Rehermann et al., J. Clin. Invest. 97:1655-1665, 1996; and Rehermann et al. J. Clin. Invest. 98:1432-1440, 1996).

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Target cell lines are autologous and allogeneic EBV-transformed B-LCL that are either purchased from the American Society for Histocompatibility and Immunogenetics (ASHI, Boston, MA) or established from the pool of patients as described (Guilhot, *et al. J. Virol.* 66:2670-2678, 1992).

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Cytotoxicity assays are performed in the following manner. Target cells consist of either allogeneic HLA-matched or autologous EBV-transformed B lymphoblastoid cell line that are incubated overnight with the synthetic

peptide epitope of the invention at 10 μ M, and labeled with 100 μ Ci of ⁵¹Cr (Amersham Corp., Arlington Heights, IL) for 1 hour after which they are washed four times with HBSS.

Cytolytic activity is determined in a standard 4-h, split well ⁵¹Cr release assay using U-bottomed 96 well plates containing 3,000 targets/well. Stimulated PBMC are tested at effector/target (E/T) ratios of 20-50:1 on day 14. Percent cytotoxicity is determined from the formula: 100 x [(experimental release-spontaneous release)/maximum release-spontaneous release)]. Maximum release is determined by lysis of targets by detergent (2% Triton X-100; Sigma Chemical Co., St. Louis, MO). Spontaneous release is <25% of maximum release for all experiments.

The results of such an analysis indicate the extent to which HLA-restricted CTL populations have been stimulated by previous exposure to 101P3A11 or an 101P3A11 vaccine.

Similarly, Class II restricted HTL responses may also be analyzed. Purified PBMC are cultured in a 96-well flat bottom plate at a density of 1.5×10^5 cells/well and are stimulated with 10 µg/ml synthetic peptide of the invention, whole 101P3A11 antigen, or PHA. Cells are routinely plated in replicates of 4-6 wells for each condition. After seven days of culture, the medium is removed and replaced with fresh medium containing 10U/ml IL-2. Two days later, 1 µCi ³H-thymidine is added to each well and incubation is continued for an additional 18 hours. Cellular DNA is then harvested on glass fiber mats and analyzed for ³H-thymidine incorporation. Antigen-specific T cell proliferation is calculated as the ratio of ³H-thymidine incorporation in the presence of antigen divided by the ³H-thymidine incorporation in the absence of antigen.

Example 24: Induction Of Specific CTL Response In Humans

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A human clinical trial for an immunogenic composition comprising CTL and HTL epitopes of the invention is set up as an IND Phase I, dose escalation study and carried out as a randomized, double-blind, placebo-controlled trial. Such a trial is designed, for example, as follows:

A total of about 27 individuals are enrolled and divided into 3 groups:

Group I: 3 subjects are injected with placebo and 6 subjects are injected with 5 µg of peptide composition;

Group II: 3 subjects are injected with placebo and 6 subjects are injected with 50 µg peptide composition;

Group III: 3 subjects are injected with placebo and 6 subjects are injected with 500 µg of peptide composition.

After 4 weeks following the first injection, all subjects receive a booster inoculation at the same dosage.

The endpoints measured in this study relate to the safety and tolerability of the peptide composition as well as its immunogenicity. Cellular immune responses to the peptide composition are an index of the intrinsic activity of this the peptide composition, and can therefore be viewed as a measure of biological efficacy. The following summarize the clinical and laboratory data that relate to safety and efficacy endpoints.

Safety: The incidence of adverse events is monitored in the placebo and drug treatment group and assessed in terms of degree and reversibility.

Evaluation of Vaccine Efficacy: For evaluation of vaccine efficacy, subjects are bled before and after injection. Peripheral blood mononuclear cells are isolated from fresh heparinized blood by Ficoll-Hypaque density gradient centrifugation, aliquoted in freezing media and stored frozen. Samples are assayed for CTL and HTL activity.

The vaccine is found to be both safe and efficacious.

Example 25: Phase II Trials In Patients Expressing 101P3A11

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Phase II trials are performed to study the effect of administering the CTL-HTL peptide compositions to patients having cancer that expresses 101P3A11. The main objectives of the trial are to determine an effective dose and regimen for inducing CTLs in cancer patients that express 101P3A11, to establish the safety of inducing a CTL and HTL response in these patients, and to see to what extent activation of CTLs improves the clinical picture of these patients, as manifested, e.g., by the reduction and/or shrinking of lesions. Such a study is designed, for example, as follows:

The studies are performed in multiple centers. The trial design is an open-label, uncontrolled, dose escalation protocol wherein the peptide composition is administered as a single dose followed six weeks later by a single booster shot of the same dose. The dosages are 50, 500 and 5,000 micrograms per injection. Drug-associated adverse effects (severity and reversibility) are recorded.

There are three patient groupings. The first group is injected with 50 micrograms of the peptide composition and the second and third groups with 500 and 5,000 micrograms of peptide composition, respectively. The patients within each group range in age from 21-65 and represent diverse ethnic backgrounds. All of them have a tumor that expresses 101P3A11.

Clinical manifestations or antigen-specific T-cell responses are monitored to assess the effects of administering the peptide compositions. The vaccine composition is found to be both safe and efficacious in the treatment of 101P3A11-associated disease.

Example 26: Induction of CTL Responses Using a Prime Boost Protocol

A prime boost protocol similar in its underlying principle to that used to confirm the efficacy of a DNA vaccine in transgenic mice, such as described above in the Example entitled "The Plasmid Construct and the Degree to Which It Induces Immunogenicity," can also be used for the administration of the vaccine to humans. Such a vaccine regimen can include an initial administration of, for example, naked DNA followed by a boost using recombinant virus encoding the vaccine, or recombinant protein/polypeptide or a peptide mixture administered in an adjuvant.

For example, the initial immunization may be performed using an expression vector, such as that constructed in the Example entitled "Construction of 'Minigene' Multi-Epitope DNA Plasmids" in the form of naked nucleic acid administered IM (or SC or ID) in the amounts of 0.5-5 mg at multiple sites. The nucleic acid (0.1 to 1000 μ g) can also be administered using a gene gun. Following an incubation period of 3-4 weeks, a booster dose is then administered. The booster can be recombinant fowlpox virus administered at a dose of 5-10⁷ to $5x10^9$ pfu. An alternative recombinant virus, such as an MVA, canarypox, adenovirus, or adeno-associated virus, can also be used for the booster, or the polyepitopic protein or a mixture of the peptides can be administered. For evaluation of vaccine efficacy, patient blood samples are obtained before immunization as well as at intervals following administration of the initial vaccine and booster doses of the vaccine. Peripheral blood mononuclear cells are isolated from fresh heparinized blood by Ficoll-Hypaque density gradient centrifugation, aliquoted in freezing media and stored frozen. Samples are assayed for CTL and HTL activity.

Analysis of the results indicates that a magnitude of response sufficient to achieve a therapeutic or protective immunity against 101P3A11 is generated.

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Example 27: Administration of Vaccine Compositions Using Dendritic Cells (DC)

Vaccines comprising peptide epitopes of the invention can be administered using APCs, or "professional" APCs such as DC. In this example, peptide-pulsed DC are administered to a patient to stimulate a CTL response *in vivo*. In this method, dendritic cells are isolated, expanded, and pulsed with a vaccine comprising peptide CTL and HTL epitopes of the invention. The dendritic cells are infused back into the patient to elicit CTL and HTL responses *in vivo*. The induced CTL and HTL then destroy or facilitate destruction, respectively, of the target cells that bear the 101P3A11 protein from which the epitopes in the vaccine are derived.

For example, a cocktail of epitope-comprising peptides is administered *ex vivo* to PBMC, or isolated DC therefrom. A pharmaceutical to facilitate harvesting of DC can be used, such as ProgenipoietinTM (Monsanto, St. Louis, MO) or GM-CSF/IL-4. After pulsing the DC with peptides, and prior to reinfusion into patients, the DC are washed to remove unbound peptides.

As appreciated clinically, and readily determined by one of skill based on clinical outcomes, the number of DC reinfused into the patient can vary (see, e.g., Nature Med. 4:328, 1998; Nature Med. 2:52, 1996 and Prostate 32:272, 1997). Although 2-50 x 10⁶ DC per patient are typically administered, larger number of DC, such as 10⁷ or 10⁸ can also be provided. Such cell populations typically contain between 50-90% DC.

In some embodiments, peptide-loaded PBMC are injected into patients without purification of the DC. For example, PBMC generated after treatment with an agent such as ProgenipoietinTM are injected into patients without purification of the DC. The total number of PBMC that are administered often ranges from 10^8 to 10^{10} . Generally, the cell doses injected into patients is based on the percentage of DC in the blood of each patient, as determined, for example, by immunofluorescence analysis with specific anti-DC antibodies. Thus, for example, if ProgenipoietinTM mobilizes 2% DC in the peripheral blood of a given patient, and that patient is to receive 5 x 10^6 DC, then the patient will be injected with a total of 2.5×10^8 peptide-loaded PBMC. The percent DC mobilized by an agent such as ProgenipoietinTM is typically estimated to be between 2-10%, but can vary as appreciated by one of skill in the art.

Ex vivo activation of CTL/HTL responses

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Alternatively, ex vivo CTL or HTL responses to 101P3A11 antigens can be induced by incubating, in tissue culture, the patient's, or genetically compatible, CTL or HTL precursor cells together with a source of APC, such as DC, and immunogenic peptides. After an appropriate incubation time (typically about 7-28 days), in which the precursor cells are activated and expanded into effector cells, the cells are infused into the patient, where they will destroy (CTL) or facilitate destruction (HTL) of their specific target cells, i.e., tumor cells.

Example 28: An Alternative Method of Identifying and Confirming Motif-Bearing Peptides

Another method of identifying and confirming motif-bearing peptides is to elute them from cells bearing defined MHC molecules. For example, EBV transformed B cell lines used for tissue typing have been extensively characterized to determine which HLA molecules they express. In certain cases these cells express only a single type of HLA molecule. These cells can be transfected with nucleic acids that express the antigen of interest, e.g. 101P3A11. Peptides produced by endogenous antigen processing of peptides produced as a result of transfection will then bind to HLA molecules within the cell and be transported and displayed on the cell's surface. Peptides are then eluted from the

HLA molecules by exposure to mild acid conditions and their amino acid sequence determined, e.g., by mass spectral analysis (e.g., Kubo et al., J. Immunol. 152:3913, 1994). Because the majority of peptides that bind a particular HLA molecule are motif-bearing, this is an alternative modality for obtaining the motif-bearing peptides correlated with the particular HLA molecule expressed on the cell.

Alternatively, cell lines that do not express endogenous HLA molecules can be transfected with an expression construct encoding a single HLA allele. These cells can then be used as described, *i.e.*, they can then be transfected with nucleic acids that encode 101P3A11 to isolate peptides corresponding to 101P3A11 that have been presented on the cell surface. Peptides obtained from such an analysis will bear motif(s) that correspond to binding to the single HLA allele that is expressed in the cell.

As appreciated by one in the art, one can perform a similar analysis on a cell bearing more than one HLA allele and subsequently determine peptides specific for each HLA allele expressed. Moreover, one of skill would also recognize that means other than transfection, such as loading with a protein antigen, can be used to provide a source of antigen to the cell.

Example 29: Complementary Polynucleotides

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Sequences complementary to the 101P3A11-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring 101P3A11. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using, e.g., OLIGO 4.06 software (National Biosciences) and the coding sequence of 101P3A11. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the 101P3A11-encoding transcript.

Example 30: Purification of Naturally-occurring or Recombinant 101P3A11 Using 101P3A11 Specific Antibodies

Naturally occurring or recombinant 101P3A11 is substantially purified by immunoaffinity chromatography using antibodies specific for 101P3A11. An immunoaffinity column is constructed by covalently coupling anti-101P3A11 antibody to an activated chromatographic resin, such as CrBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing 101P3A11 are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of 101P3A11 (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/101P3A11 binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and GCR.P is collected.

Example 31: Identification of Molecules Which Interact with 101P3A11

101P3A11, or biologically active fragments thereof, are labeled with 121 1 Bolton-Hunter reagent. (See, e.g., Bolton et al. (1973) Biochem. J. 133:529.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled 101P3A11, washed, and any wells with labeled 101P3A11 complex are assayed. Data

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obtained using different concentrations of 101P3A11 are used to calculate values for the number, affinity, and association of 101P3A11 with the candidate molecules.

Example 32: In Vivo Assay for 101P3A11 Tumor Growth Promotion

The effect of the 101P3A11 protein on tumor cell growth can be confirmed *in vivo* by gene overexpression in a variety of cancer cells, including prostate, kidney, colon and bladder. For example, SCID mice can be injected SQ on each flank with 1 x 10⁶ prostate, kidney, colon or bladder cancer cells (such as PC3, LNCaP, SCaBER, UM-UC-3, HT1376, SK-CO, Caco, RT4, T24, Caki, A-498 and SW839 cells) containing tkNeo empty vector or 101P3A11.

At least two strategies can be used:

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- (1) Constitutive 101P3A11 expression under regulation of a promoter such as a constitutive promoter obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and Simian Virus 40 (SV40), or from heterologous mammalian promoters, *e.g.*, the actin promoter or an immunoglobulin promoter, provided such promoters are compatible with the host cell systems.
- (2) Regulated expression under control of an inducible vector system, such as ecdysone, tet, etc., can be used provided such promoters are compatible with the host cell systems. Tumor volume is then monitored at the appearance of palpable tumors and is followed over time to validate that 101P3A11-expressing cells grow at a faster rate and that tumors produced by 101P3A11-expressing cells demonstrate characteristics of altered aggressiveness (e.g., enhanced metastasis, vascularization, reduced responsiveness to chemotherapeutic drugs). Figure 21 compares subcutaneous growth of control 3T3-neo and 3T3-101P3A11 cells. One million cells stably expressing neo or 101P3A11 were injected subcutaneously in SCID mice along with matrigel. Tumor volume was evaluated by caliper measurements. This experiment demonstrates that expression of 101P3A11 in NIH 3T3 cells is induces tumor formation in 6/6 mice. Figure 42 shows demonstrates that 101P3A11 induces orthotopic growth of tumors. The results indicated that expression of 101P3A11 is sufficient to induce tumor formation in vivo. Additionally, were implanted with the same cells orthotopically in the prostate to determine if 101P3A11 has an effect on local growth in the prostate or on the ability of the cells to metastasize, specifically to lungs or lymph nodes. In an analogous manner, cells cab be implanted orthotopically in , bladder, colon or kidney. (Saffran, D., et al., PNAS 10:1073-1078; Fu, X., et al., Int. J. Cancer, 1991. 49: p. 938-939; Chang, S., et al., Anticancer Res., 1997. 17: p. 3239-3242; Peralta, E. A., et al., J. Urol., 1999. 162: p. 1806-1811).

Furthermore, this assay is useful to confirm the 101P3A11 inhibitory effect of candidate therapeutic compositions, such as for example, 101P3A11 antibodies or intrabodies, and 101P3A11 antisense molecules or ribozymes, or 101P3A11 directed small molecules. In Figure 22, we depict the effect of a small molecule, pertussis toxin (PTX) on tumor formation by 3T3-101P3A11 cells. In this experiment, SCID mice were injected with 3T3-101P3A11 alone or in conjunction with PTX. Each mouse was given 5 doses of PTX at 3-4 days interval. Tumor volume was evaluated by caliper measurements. Figure 22 shows that PTX inhibits tumor growth in a dose dependent manner. In addition to demonstrating that 101A3P11 plays an important role in tumor growth, Figures 21 and 22 identify a signaling pathway associated with 101P3A11 and indicate that 101P3A11 produced its effect on tumor growth by activating an adenylate cyclase dependent pathway.

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Example 33: 101P3A11 Monoclonal Antibody-mediated Inhibition of Tumors In Vivo

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The significant expression of 101P3A11 in cancer tissues, together with its restricted expression in normal tissues, makes 101P3A11 an excellent target for antibody therapy. In cases where the monoclonal antibody target is a cell surface protein, as is 101P3A11, antibodies have been shown to be efficacious at inhibiting tumor growth (See, e.g., Saffran, D., et al., PNAS 10:1073-1078 or www.pnas.org/cgi/doi/10.1073/pnas.051624698). In cases where the target is not on the cell surface, such as PSA and PAP in prostate cancer, antibodies have also been shown to recognize and inhibit growth of cells expressing those proteins (Saffran, D.C., et al., Cancer and Metastasis Reviews, 1999. 18: 437-449). As with any cellular protein with a restricted expression profile, 101P3A11 is a target for T cell-based immunotherapy.

Accordingly, the therapeutic efficacy of anti-101P3A11 mAbs in human colon, kidney, bladder and prostate cancer mouse models is modeled in 101P3A11-expressing kidney, colon, bladder or prostate cancer xenografts or cancer cell lines, such as those described in the Example entitled "In Vivo Assay for 101P3A11 Tumor Growth Promotion", that have been engineered to express 101P3A11.

Antibody efficacy on tumor growth and metastasis formation is confirmed, e.g., in a mouse orthotopic prostate, colon, bladder or kidney cancer xenograft model. The antibodies can be unconjugated, or can be conjugated to a therapeutic modality, as appreciated in the art. It is confirmed that anti-101P3A11 mAbs inhibit formation of 101P3A11-expressing kidney, colon, bladder and prostate tumors. Anti-101P3A11 mAbs also retard the growth of established orthotopic tumors and prolong survival of tumor-bearing mice. These results indicate the utility of anti-101P3A11 mAbs in the treatment of local and advanced stages of cancer. (See, e.g., Saffran, D., et al., PNAS 10:1073-1078 or www.pnas.org/cgi/doi/10.1073/pnas.051624698)

Administration of anti-101P3A11 mAbs retard established orthotopic tumor growth and inhibit metastasis to distant sites, resulting in a significant prolongation in the survival of tumor-bearing mice. These studies indicate that 101P3A11 is an attractive target for immunotherapy and demonstrate the therapeutic potential of anti-101P3A11 mAbs for the treatment of local and metastatic kidney, colon, bladder and prostate cancer. Similar studies manifest that 101P3A11 is safe and effective when used in combination with other therapeutic modalities such as surgery, radiation therapy, hormone therapy or chemotherapy.

This example demonstrates that unconjugated 101P3A11 monoclonal antibodies effectively to inhibit the growth of human bladder tumors grown in SCID mice; accordingly a combination of such efficacious monoclonal antibodies is also effective.

Example 34: Induction Of Specific CTL Response In Humans

A human clinical trial for an immunogenic composition comprising CTL and HTL epitopes of the invention is set up as an IND Phase I, dose escalation study and carried out as a randomized, double-blind, placebo-controlled trial. Such a trial is designed, for example, as follows:

A total of about 27 individuals are enrolled and divided into 3 groups:

Group I: 3 subjects are injected with placebo and 6 subjects are injected with 5 µg of peptide composition;

Group II: 3 subjects are injected with placebo and 6 subjects are injected with 50 µg peptide composition;

Group III: 3 subjects are injected with placebo and 6 subjects are injected with 500 µg of peptide composition. After 4 weeks following the first injection, all subjects receive a booster inoculation at the same dosage.

The endpoints measured in this study relate to the safety and tolerability of the peptide composition as well as its immunogenicity. Cellular immune responses to the peptide composition are an index of the intrinsic activity of this the peptide composition, and can therefore be viewed as a measure of biological efficacy. The following summarize the clinical and laboratory data that relate to safety and efficacy endpoints.

Safety: The incidence of adverse events is monitored in the placebo and drug treatment group and assessed in terms of degree and reversibility.

Evaluation of Vaccine Efficacy: For evaluation of vaccine efficacy, subjects are bled before and after injection. Peripheral blood mononuclear cells are isolated from fresh heparinized blood by Ficoll-Hypaque density gradient centrifugation, aliquoted in freezing media and stored frozen. Samples are assayed for CTL and HTL activity.

The vaccine is found to be both safe and efficacious.

Example 35: Splice Variants of 101P3A11

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Splice variants are also called alternative transcripts. When a gene is transcribed from genomic DNA, the initial RNA is generally spliced to produce functional mRNA, which has only exons and is used for translation into an amino acid sequence. Accordingly, a given gene can have zero to many alternatively spliced mRNA products. Alternative transcripts each have a unique exon makeup, and can have different coding and/or non-coding (5' or 3' end) portions, from the original transcript. Alternative transcripts can code for similar proteins with the same or a similar function or may encode proteins with different functions, and may be expressed in the same tissue at the same time, or at different tissue at different times, proteins encoded by alternative transcripts can have similar or different cellular or extracellular localizations, e.g., be secreted.

Splice variants are identified by a variety of art-accepted methods. For example, splice variants are identified by use of EST data. First, all human ESTs were grouped into clusters which show direct or indirect identity with each other. Second, ESTs in the same cluster were further grouped into sub-clusters and assembled into a consensus sequence. The starting gene is compared to the consensus sequence(s). Each consensus sequence is a potential splice variant for that gene (see, e.g., http://www.doubletwist.com/products/c11_agentsOverview.jhtml). Even when a variant is identified that is not a full-length clone, that portion of the variant is very useful for antigen generation and for further cloning of the full-length splice variant, using techniques known in the art.

Moreover, computer programs are available in the art that identify splice variants based on genomic sequences. Genomic-based variant identification programs include FgenesH (A. Salamov and V. Solovyev, "Ab initio gene finding in Drosophila genomic DNA," Genome Research. 2000 April;10(4):516-22); Grail (http://compbio.ornl.gov/Grail-bin/EmptyGrailForm) and GenScan (http://genes.mit.edu/GENSCAN.html). For a general discussion of splice variant identification protocols see., e.g., Southan, C., "A genomic perspective on human proteases," FEBS Lett. 2001 Jun 8;498(2-3):214-8; de Souza, SJ, et al., "Identification of human chromosome 22 transcribed sequences with ORF expressed sequence tags," Proc. Natl Acad Sci U S A. 2000 Nov 7;97(23):12690-3.

For variants identified by the EST-based method, Table XXI shows the nucleotide sequences of the splice variants. Table XXII shows the alignment of the splice variant with the 101P3A11 nucleic acid sequence. Table XXIII

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displays the single longest alignment of an amino acid sequence encoded by a splice variant, out of all six potential reading frames with 101P3A11. Thus, for each splice variant, a variant's reading frame that encodes the longest single contiguous peptide homology between 101P3A11 and the variant is the proper reading frame orientation for the variant. Due to the possibility of sequencing errors in EST or genomic data, other peptides in the relevant reading frame orientation (5' to 3' or 3' to 5') can also be encoded by the variant. Table XXIV lays out all three frame shifted amino acid translations of the splice variant for the identified reading frame orientation. Tables XXI through XXIV are set forth herein on a variant-by-variant basis.

For variants identified by any one of the genomic sequence-based methods, Table XXI shows the nucleotide sequences of the splice variant. Table XXII shows the alignment of the splice variant with the 101P3A11 nucleic acid sequence. Table XXIII displays the alignment of amino acid sequence of the predicted transcript(s) with 101P3A11. The genomic-based computer programs predict a transcript from genomic sequence, and not only predict exons but also set open reading frame as the first forward open reading frame. The predicted transcript does not contain 5' or 3' untranslated region (UTR). It starts with ATG and ends with a stop codon, TAG, TGA or TAA. In case the transcript is predicted on the reverse strand of the genomic sequence, the sequence of the transcript is reverse-complemented to the genomic sequence of the exons. Thus, the genomic-based programs provide the correct transcript sequence, with 5' to 3' orientation and +1 as the open reading frame. However, due to the possibility of inaccurate prediction of exons or the possibility of sequencing errors in genomic data, other peptides in other forward open reading frames can also be encoded by the variant. Table XXIV lays out all amino acid translations of the splice variant in each of the three forward reading frames. Tables XXI through XXIV are set forth herein on a variant-by-variant basis.

To further confirm the parameters of a splice variant, a variety of techniques available in the art are used, such as full-length cloning, proteomic validation, PCR-based validation, and 5' RACE validation, etc. (see e.g., Proteomic Validation: Brennan SO, Fellowes AP, George PM.; "Albumin banks peninsula: a new termination variant characterized by electrospray mass spectrometry." Biochim Biophys Acta. 1999 Aug 17;1433(1-2):321-6; Ferranti P, et al., "Differential splicing of pre-messenger RNA produces multiple forms of mature caprine alpha(s1)-casein." Eur J Biochem. 1997 Oct 1;249(1):1-7; PCR-based Validation: Wellmann S, et al., "Specific reverse transcription-PCR quantification of vascular endothelial growth factor (VEGF) splice variants by LightCycler technology." Clin Chem. 2001 Apr;47(4):654-60; Jia HP, et al., Discovery of new human beta-defensins using a genomics-based approach," Gene. 2001 Jan 24;263(1-2):211-8; PCR-based and 5' RACE Validation: Brigle I.E., et al., "Organization of the murine reduced folate carrier gene and identification of variant splice forms," Biochim Biophys Acta. 1997 Aug 7; 1353(2): 191-8.

It is known in the art that genomic regions are modulated in cancers. When the genomic region to which 101P3A11 maps is modulated in a particular cancer, the splice variants of 101P3A11 are modulated as well. Disclosed herein is that 101P3A11 has a particular expression profile. Splice variants of 101P3A11 that are structurally and/or functionally similar to 101P3A11 share this expression pattern, thus serving as tumor-associated markers/antigens.

Using the EST assembly approach, we identified one splice variant.

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Example 36: Splice Variant Protein Characteristics

The present variant protein set forth in Table XXI is understood to be partial, and thus to comprise domains of the full protein. Nucleotides 1-217 of variant align with nucleotides 66-290 of 101P3A11. Similarly, amino acids 3-155 of the 1-1P3A11 variant protein align with amino acids 70-222 of 101P3A11 with 100% identity, while the remaining downstream amino acids diverge from the 101P3A11. This pattern of high homology to one section of the parent protein coupled to a high divergence from the remaining portions of the parent protein form the hallmark of a splice variant.

Protein blast analysis of 101P3A11 variant shows that the 101P3A11 variant is homologous to the human olfactory receptor 52D1 (gi 14423837) with 35% identify over 68 amino acids located towards the amino terminal of 101P3A11 variant 1. 101P3A11 variant 1 shows homology to rat olfactory receptor RA1c (gi 3420759) with 53% identity over 32 amino acids. Analysis by pFam or Prosite did not identify any motifs.

Based on TMPred bioinformatic analysis the 101P3A11 variant appears to contain one transmembrane domain. In contrast, Sosui predicts no transmembrane domain but suggests that 101P3A11 variant 1 forms a soluble intracellular protein. Due to its homology to 101P3A11 and olfactory receptors, the 101P3A11 variant 1 is associated with cancer and signaling.

Example 37: Identification of 101P3A11 sequences involved in ligand binding.

As shown in Figure 4, the transmembrane regions of 101P3A11 and mouse olfactory receptor S25 (ORS25)predicted using the TMHMM algorithm are highlighted in gray. The amino acids of ORS25 predicted by Floriano, et al. to be involved in binding of the ligand hexanol and/or involved in the formation of the ligand binding pocket are italicized and bolded in Figure 4, and are listed below. (Floriano, W.B., et al, 2000, Proc. Natl. Acad. Sci., USA, 97:10712-10716)

	Leu 131	. Ala 230
25	Val 134	Ile 231
	Val 135	Gly 234
	Gly 138	Thr 284
	Thr139	Phe 287
	Ser 193	Gln 300
30	Ser 197,	Lys 302
	Phe 225	

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Sequences of 101P3A11 involved in ligand binding are identified based on homology to mouse olfactory receptor S25. Shown is the amino acid alignment of 101P3A11 with mouse olfactory receptor S25 depicting the predicted transmembrane domains of each GPCR. The amino acids of S25 involved in the recognition and binding of its ligand hexanol or that lie in the proximity of the binding pocket (Floriano, W.B., et al, 2000, Proc. Natl. Acad. Sci., USA, 97:10712-10716), are also shown. These amino acids lie close to or within the transmembrane domains of ORS25. Accordingly, the structurally homologous regions of 101P3A11 are involved in the binding of its cognate

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ligand. These regions encode the amino acids of the first extracellular loop and of the amino terminal end of transmembrane domain 3 (amino acids 82-112), the amino acids at the carboxyl terminal end of transmembrane domain 4 and into the second extracellular loop (amino acids 160-185), the amino acids at the end of the second extracellular loop and into transmembrane domain 5 (amino acids 186-212), and the amino acids at the carboxyl terminal end of transmembrane domain 6, the third extracellular loop, and the amino terminal end of transmembrane domain 7 (amino acids 250-280). Thus, ligands of 101P3A11 are identified that interact with at least 3 of the following regions of 101P3A11: amino acids 82-112, amino acids 160-185, amino acids 186-212, and, amino acids 250-280.

Example 38: Homology Comparison of 101P3A11 to Known Sequences

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The 101P3A11 protein of Figure 3 has 317 amino acids with calculated molecular weight of 35.2 kDa, and pI of 8.7. 101P3A11 is predicted to be a cell surface protein. Cellular localization was demonstrated by FACS analysis and immunofluorescence in cells engineered to express 101P3A11.

101P3A11 shows best homology to rat olfactory receptor RA1c (gi 3420759, http://www.ncbi.nlm.nih.gov) sharing 59% identity and 76% homology with that protein. 101P3A11 also shows homology to human prostate specific GPCR (gi 13540539) and human olfactory receptor 5II12 (gi 14423836), sharing 59% identities/ 77% homology, and 53% identities/ 69% homology with each, respectively (Figures 23-25).

Sequence and motif analysis indicate that 101P3A11 belongs to the family of olfactory receptors. Bioinformatic analysis revealed 101P3A11 to be a 7 transmembrane protein, with strong domain and structural homology to G-protein coupled receptors (GPCRs) (see Table XXV, TM Pred, Sosui, Pfam, Blocks, Print). Proteins that are members of the G-protein coupled receptor family exhibit an extracellular amino-terminus, three extracellular loops, three intracellular loops and an intracellular carboxyl terminus. G-protein coupled receptors are seven-transmembrane receptors that are stimulated by polypeptide hormones, neurotransmitters, chemokines and phospholipids (Civelli O et al, Trends Neurosci. 2001, 24:230; Vrecl M et al Mol Endocrinol. 1998, 12:1818). Ligand binding traditionally occurs between the first and second extracellular loops of the GPCR. Upon ligand binding GPCRs transduce signals across the cell surface membrane by associating with trimeric G proteins. Their signals are transmitted via trimeric guanine-nucleotide binding proteins (G proteins) to effector enzymes or ion channels (Simon et al., 1991, Science 252: 802). Signal transduction and biological output mediated by GPCR can be modulated through various mechanisms including peptide mimics, small molecule inhibitors and GPCR kinases or GRK (Pitcher JA et al, J Biol Chem. 1999, 3;274:34531; Fawzi AB, et al. 2001, Mol. Pharmacol., 59:30).

Recently, GPCRs have also been shown to link to mitogenic signaling pathways of tyrosine kinases (Luttrell et al., 1999, Science 283: 655; Luttrell et al., 1999 Curr Opin Cell Biol 11: 177). GPCRs are regulated by phosphorylation mediated by GPCR kinases (GRKs), which themselves are indirectly activated by the GPCRs (Pitcher et al., 1998, Ann. Rev. Biochem. 67: 653). Olfactory GPCRs transmit their signals by activating the cAMP pathway via adenylate cyclase resulting in downstream signaling to protein kinase A, and by activating the phospholipase C pathway by generating inositol 1,4,5-trisphosphate (IP3) and diacyl-glycerol (DAG) (Breer, 1993, Ciba Found Symp 179: 97; Bruch, 1996, Comp Biochem Physiol B Biochem Mol Biol 113:451). IP3 results in an increase in intracellular calcium, while DAG activates protein kinase C.

Recent studies have associated GPCRs with cellular transformation. In particular, KSHV G protein-coupled receptor was found to transform NIH 3T3 cells *in vitro* and induces multifocal KS-like lesions in KSHV-GPCR-sd-63712

transgenic mice (Schwarz M, Murphy PM. J Immunol 2001, 167:505). KSHV-GPCR was capable of producing its effect on endothelial cells and fibroblasts by activating defined signaling pathways, including the AKT survival pathway (Montaner S et al, Cancer Res 2001, 61:2641). In addition, KSHV-GPCR induced the activation of mitogenic pathways such as AP-1 and NFkB, resulting in the expression of pro-inflammatory genes (Schwarz M, Murphy PM. J Immunol 2001, 167:505). Other GPCR associated with tumor formation include G2A, and the PAR-1, which has been found to induce transformation of NIH 3T3 cells (Whitehead I et al, Oncogene 2001, 20:1547).

This information indicates that 101P3A11 plays a role in the transformation of mammalian cells, induces mitogenic responses including activation of various signaling pathways, and regulate gene transcription by transmitting cell surface signals to the nucleus, see also, the Example entitled, "In Vivo Assay for 101P3A11 Tumor Growth Promotion".

Accordingly, when 101P3A11 functions as a regulator of cell transformation, tumor formation, or as a modulator of transcription involved in activating genes associated with inflammation, tumorigenesis or proliferation, 101P3A11 is used for therapeutic, diagnostic, prognostic and/or preventative purposes, in manners analogous to or that track other GPCRs as discussed herein and in the art..

Example 39: Identification and Confirmation of Potential Signal Transduction Pathways

Many mammalian proteins have been reported to interact with signaling molecules and to participate in regulating signaling pathways. (J Neurochem. 2001; 76:217-223). In particular, GPCRs have been reported to activate MAK cascades as well as G proteins, and been associated with the EGFR pathway in epithelial cells (Naor, Z., et al, Trends Endocrinol Metab. 2000, 11:91; Vacca F et al, Cancer Res. 2000, 60:5310; Della Rocca G.J., et al, J Biol Chem. 1999, 274:13978). Using immunoprecipitation and Western blotting techniques, proteins are identified that associate with 101P3A11 and mediate signaling events. Several pathways known to play a role in cancer biology can be regulated by 101P3A11, including phospholipid pathways such as PI3K, AKT, etc, adhesion and migration pathways, including FAK, Rho, Rac-1, etc, as well as mitogenic/survival cascades such as ERK, p38, etc (Cell Growth Differ. 2000,11:279; J Biol Chem. 1999, 274:801; Oncogene. 2000, 19:3003; J. Cell Biol. 1997, 138:913).

Using Western blotting and other techniques, the ability of 101P3A11 to regulate these pathways is confirmed. Cells expressing or lacking 101P3A11 are either left untreated or stimulated with cytokines, androgen and anti-integrin antibodies. Cell lysates were analyzed using anti-phospho-specific antibodies (Cell Signaling, Santa Cruz Biotechnology) in order to detect phosphorylation and regulation of ERK, p38, AKT, PI3K, PLC and other signaling molecules. Using such techniques, we showed that 101P3A11 alters the tyrosine phosphorylation pattern of NIH 3T3 cells (Figure 26) indicating that 101P3A11 is regulating protein kinases and phosphatases. In the experiment shown in Figure 26, control 3T3-neo and 3T3-101P3A11 cells were either treated with 0.5 or 10% FBS and whole cell lysates were analyzed by anti-phosphotyrosine Western blotting. Expression of 101P3A11 resulted in reduced phosphorylation of several proteins in NIH-3T3 cells, while inducing the phosphorylation of proteins at 79-81 and 28-32 kDa.

Using anti-Phospho-ERK antibodies, we demonstrated that expression of 101P3A11 induced ERK phosphorylation in the prostate cancer cell line PC3 (Figures 27A and Figure 27B), and that ERK phosphorylation in 101P3A11 expressing cells was regulated by GPCR ligands. In this experiment, control PC3-neo cells and PC3-101P3A11 cells were left untreated (0.1% FBS) or were stimulated with 10% FBS, lipophosphatidic acid (LPA), gastrin

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releasing peptide (GRP), leukotriene (LKB4) or platelet activating factor (PAF). The cells were lysed and analyzed by Western blotting using anti-Phospho-ERK (Figure 27A) or anti-ERK (Figure 27B) mAb. The results showed that expression of 101P3A11 mediated significant ERK phosphorylation by FBS, LPA, GRP and PAF, while LKB4 resulted in a more modest level of ERK phosphorylation in PC3-101P3A11 cells. In contrast, none of the GPCR ligands induced significant ERK phosphorylation in PC3-Neo cells, demonstrating the specificity of GPCR ligands-mediated responses in 101P3A11 expressing cells. The ERK overlay demonstrated equal loading, supporting the specificity of this data. In order to delineate the signaling pathway by which 101P3A11 mediates ERK phosphorylation in cancer cells, it was confirmed which of the two pathway inhibitors: MEK inhibitor PD98059 or the p38 inhibitor SB203580 regulate 101P3A11 mediated ERK phosphorylation (Figure 28). To obtain this data, PC3-neo and PC3-101P3A11 cells were treated with media alone or in the presence of PD98059, SB203580, or genistein were stimulated with FBS or GRP. Cells were lysed and analyzed by Western blotting using anti-Phospho-ERK or anti-ERK mAb. Treatment with 10% FBS or with GRP induced the phosphorylation of ERK in PC3-101P3A11 but not in control PC3-neo cells. 101P3A11-mediated ERK phosphorylation was inhibited by the MEK-1 inhibitor PD98059 but not the p38 inhibitor SB203580 or genistein. The ERK overlay demonstrated equal loading, supporting the specificity of the results. These results were confirmed by those obtained in two additional sets of experiments. The inhibition of 101P3A11-mediated ERK phosphorylation by PD98059 demonstrates that 101P3A11 activated the classical MEK-ERK cascade, a pathway associated with mitogenesis, proliferation and tumorigenesis.

Results in Figures 26-28 indicate that 101P3A11 regulates the activity of kinases, including ERK, and phosphatases. In order to confirm the association of 101P3A11 with phosphatase activity, the effect of the protein phosphatase inhibitor sodium orthovanadate on 101P3A11 mediated ERK phosphorylation was determined (Figure 29). PC3-neo and PC3-101P3A11 cells were grown in media alone or in the presence of sodium orthovanadate (Na3VO4), and were stimulated with 0.1% or 10% FBS. Cells were lysed and analyzed by Western blotting using anti-Phospho-ERK or anti-ERK mAb. Treatment with Na3VO4 resulted in a 4.5-fold increase in ERK phosphorylation in PC3-101P3A11 cells, compared to a two-fold increase in PC3-neo cells. Results in Figure 29 confirm the contribution of protein phosphatases to 101P3A11 mediated signaling.

Several GPCRs have been shown to transactivate receptor tyrosine kinases associated with the cell membrane, such as the EGF receptor (EGFR) (Pierce K.L., et al, J Biol Chem. 2001, 276:23155; Nath, D., et al, J Cell Sci. 2001, 114:1213). In order to determine whether 101P3A11 signaling results in the activation of EGFR, we compared the effect of the EGFR inhibitor, AG1517, on EGFR- and 101P3A11-mediated ERK phosphorylation (Figure 30). In Figure 30, PC3-neo and PC3-101P3A11 cells were grown in media alone (0.1% FBS) or in the presence of AG1517. The cells were stimulated with 0.1% or 10% FBS, GRP or EGF, lysed and analyzed by Western blotting using anti-Phospho-ERK or anti-ERK mAb. Treatment with 10% FBS, GRP and EGF induced ERK phosphorylation in PC3-101P3A11 cells. ERK phosphorylation by EGF was completely inhibited by AG1517. 101P3A11 mediated ERK phosphorylation in cells treated with 10% FBS was partially inhibited by AG1517. Data in Figure 30 indicate that some cross talk occurred between 101P3A11 and EGFR signaling pathways.

In addition to activating the ERK cascade, 101P3A11 activated a parallel MAK pathway, namely p38. In Figure 31A and Figure 31B, PC3-neo and PC3-101P3A11 cells were grown in 1% or 10% FBS. Cells were lysed and analyzed by Western blotting using anti-Phospho-p38 (Figure 31A) or anti-p38 (Figure 31B) monoclonal antibody

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(mAb). Our results demonstrate that expression of 101P3A11 mediated p38 phosphorylation in cells treated with 10% FBS. Equal loading was demonstrated in the p38 overlay.

Results shown in Figures 26-30 and Figure 31A-31B confirm that 101P3A11 activates several signaling pathways in cancer cells, including the ERK and p38 cascades. In addition to MAPK, 101P3A11 signaling was associated with protein phosphatase activity and EGFR transactivation. These signaling pathways have been associated with cell growth, survival and transcriptional activation, all of which play an important role in tumor initiation and progression. When 101P3A11 plays a role in the regulation of signaling pathways, whether individually or communally, it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes.

To confirm that 101P3A11 directly or indirectly activates known signal transduction pathways in cells, luciferase (luc) based transcriptional reporter assays are carried out in cells expressing individual genes. These transcriptional reporters contain consensus-binding sites for known transcription factors that lie downstream of well-characterized signal transduction pathways. The reporters and examples of these associated transcription factors, signal transduction pathways, and activation stimuli are listed below.

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- 1. NFkB-luc, NFkB/Rel; Ik-kinase/SAPK; growth/apoptosis/stress
- 2. SRE-luc, SRF/TCF/ELK1; MAPK/SAPK; growth/differentiation
- 3. AP-1-luc, FOS/JUN; MAPK/SAPK/PKC; growth/apoptosis/stress
- 4. ARE-luc, androgen receptor; steroids/MAPK; growth/differentiation/apoptosis
- 5. p53-luc, p53; SAPK; growth/differentiation/apoptosis
- 6. CRE-luc, CREB/ATF2; PKA/p38; growth/apoptosis/stress

Gene-mediated effects can be assayed in cells showing mRNA expression. Luciferase reporter plasmids can be introduced by lipid-mediated transfection (TFX-50, Promega). Luciferase activity, an indicator of relative transcriptional activity, is measured by incubation of cell extracts with luciferin substrate and luminescence of the reaction is monitored in a luminometer.

Signaling pathways activated by 101P3A11 are mapped and used for the identification and validation of therapeutic targets. When 101P3A11 is involved in cell signaling, it is used as target for diagnostic, prognostic, preventative and/or therapeutic purposes.

Example 40: 101P3A11 Functions as a GPCR

Sequence and homology analysis of 101P3A11 indicated that 101P3A11 is a member of the olfactory receptor family of GPCR. Olfactory receptors are known to regulate biological responses by activating adenylate cyclase. In order to confirm that 101P3A11 functions as a GPCR and mediates the activation of adenylate cyclase, cAMP accumulation in PC3 and PC3-101P3A11 cells were compared (Figure 32). Control PC3 and PC3-101P3A11 cells were grown in a low concentration of fetal bovine serum (FBS) for 14 hrs in the presence or absence of pertussis toxin (PTX). The cells were stimulated with 0.1% or 10% FBS, washed in PBS and lysed using a lysis buffer provided by Amersham Pharmacia. Intracellular concentration of cAMP was measured using a commercially available enzyme immunoassay (EIA) according to the manufacturer's recommendations (Amersham Pharmacia). Each assay was performed in duplicate.

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Calculations of cAMP concentrations were based on OD450 of the standard curve. Expression of 101P3A11 induced a four-fold increase in cAMP accumulation in the absence of stimulation. Treatment with 10% FBS further enhanced cAMP accumulation in PC3-101P3A11 cells to nearly seven-fold over control PC3-neo cells. 101P3A11 mediated cAMP accumulation was inhibited by PTX. These results were confirmed by two separate sets of experiments. Results shown in Figure 32 demonstrate that 101P3A11 functions as a GPCR in prostate cancer cells and exhibits classical GPCR characteristics, such as cAMP accumulation that is inhibited by PTX.

Since adenylate cyclase activity modulates intracellular levels of cAMP and induce downstream signaling events such as activation of protein kinase A, calcium and ERK MAPK signaling (Pierce K.L., et al, Oncogene. 2001, 20:1532), we determined that PTX, an inhibitor of adenylate cyclase signaling, prevents 101P3A11-mediated ERK phosphorylation along with inhibiting cAMP accumulation (Figure 33 and Figure 34). PC3-neo and PC3-101P3A11 cells were grown overnight in 0.1% FBS in media alone or in the presence of pertussis toxin (PTX). Cells were stimulated with 0.1% or 10% FBS (Figure 33) or 10% FBS, EGF or GRP (Figure 34). Cells were lysed and analyzed by Western blotting using anti-Phospho-ERK mAb. Expression of 101P3A11 mediated ERK phosphorylation by 10% FBS in PC3 cells, which was inhibited by PTX (Figure 33 and Figure 34). In contrast, GRP and EGF-mediated ERK phosphorylation was relatively unaffected by PTX (Figure 34), demonstrating the specificity of 101P3A11 mediated responses. These results were replicated in additional experiments.

GPCR transmit their signal by activating trimeric G proteins. Once GPCRs are activated, the associated Ga subunit binds GTP, dissociates from the receptor and participates in downstream signaling events (Schild, D., and Restrepo, D. Physiol Rev. 1998, 78:429-66). In order to determine that inhibition of Ga subunits has an effect on 101P3A11 mediated cell growth, the effect of two Ga inhibitors on the proliferation of 3T3-101P3A11 cells was investigated. Control 3T3 and 3T3-101P3A11 cells were grown in the presence or absence of suramin or its derivative NF 449 (Sigma). Cells were analyzed for proliferation 72 hours later (Figure 35). The experiment was performed in triplicate. The data showed that suramin and NF449 inhibited the proliferation of 3T3-101P3A11 cells by 60% and 80%, respectively. This response was 101P3A11 specific as suramin and NF449 had no effect on the proliferation of control 3T3 cells. Thus, as 101P3A11 is involved in GPCR activity, it is used as target for diagnostic, prognostic, preventative and/or therapeutic purposes.

GPCRs can be activated by a variety of ligands, including hormones, neuropeptides, chemokines, odorants and phospholipids. In the case of olfactory receptors, individual olfactory receptors may recognize multiple odorants, and can are activated by a diverse array of molecules. These ligands and molecules recognized by a receptor (as described above) are small molecules as described herein.

In order to identify 101P3A11 (small molecule) ligand(s), the possibility that epithelial cells may be secreting 101P3A11 activators was investigated (Figure 36A and 36B). Prostate cancer epithelial cells, (PC3, PC3-101P3A11, LAPC4²hT), normal prostate cells (PrEC), fibroblasts (3T3, 3T3-101P3A11), and human kidney epithelial cells (293T) were grown in the presence or absence of FBS. Cell supernatants were collected and used to stimulate PC3 and PC3-101P3A11 cells. Cell lysates from resting and supernatant treated PC3 and PC3-101P3A11 cells were lysed and analyzed by Western blotting with anti-Phospho-ERK (Figure 36A) and anti-ERK (Figure 36B) mAb. As shown in Figure 36A and Figure 36B, supernatants form normal prostate cells, PrEC, and prostate cancer cells, PC3, PC3-101P3A11 and LAPC4²hT, induced the phosphorylation of ERK in PC3-101P3A11 but not control PC3 cells. In

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contrast, no specific ERK phosphorylation was observed using supernatants from 3T3 or 293T cells. Our results show that prostate cells, grown in the absence of serum, produce one or more factors that contribute to the activation of 101P3A11 mediated signaling events. Thus, as 101P3A11 responds to stimuli and functions in signaling and GPCR activity, it is used as target for diagnostic, prognostic, preventative and/or therapeutic purposes.

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Example 41: Inhibitors of 101P3A11 GPCR Function

As mentioned in the Example entitled "Homology Comparison of 101P3A11 to Known Sequences," GPCRs are activated by ligand binding to the extracellular loops, resulting in the activation of trimeric G proteins and the initiation of several signaling cascades. Using this information, several therapeutic and small molecule strategies are utilized to inhibit GPCR activation or downstream signaling events.

One strategy inhibits receptor and ligand binding. Recent studies using several types of GPCRs, have demonstrated the effectiveness of this strategy (Fawzi AB, et al. 2001, Mol. Pharmacol., 59:30). Using a compound named SCH-202676, they inhibited agonist and antagonist binding to GPCRs by allosterically hindering ligand-GPCR interaction. Using this and even more specific allosteric (small molecule) inhibitors, signal transduction through 101P3A11 is inhibited, thereby providing therapeutic, prognostic, diagnostic and/or prophylactic benefit.

A second approach is to inhibit G alpha subunit activation. Activation of GPCRs results in the exchange of GTP for GDP on the G alpha subunit of the trimeric G protein. Inhibition of Gα activation prevents the activation of downstream signaling cascades and therefore biological effects of GPCR. One molecule used to inhibit GDP exchange on Gα subunits is Suranim (Freissmuth M et al, 1996, Mol. Pharmacol, 49:602). Since suranim functions as a universal Gα inhibitor, it prevents the activation of most Gα subunits. Using techniques described, for example and without limitation, in the present Examples entitled "*In Vivo* Assay for 101P3A11 Tumor Growth Promotion;" "Identification and Confirmation of Potential Signal Transduction Pathways," "101P3A11 Functions as a GPCR," and "Regulation of Transcription", small molecules are identified that selectively inhibit the Gα subunit that associates with 101P3A11, thereby providing therapeutic, prognostic, diagnostic and/or prophylactic benefit.

A third approach is to inhibit $G\alpha$ subunit association with GPCR. In order for trimeric G proteins to be activated following GPCR/ligand interaction, it is necessary for them to associate with their corresponding GPCR. Mutational analysis has mapped the interaction of $G\alpha$ to the first and third intracellular loops of GPCRs (Heller R at al. 1996, Biochem. Biophys. Res. Commun). Several studies have used synthetic (small molecule) peptides corresponding to the intracellular sequence of loops 1 and 3 as inhibitors (Mukherjee, S., et al. 1999, J. Biol. Chem.). Using such short peptides that serve as receptor mimics, they are used to compete for binding of Ga subunits to 101P3A11 and thereby provide therapeutic, prognostic, diagnostic and/or prophylactic benefit.

Thus, compounds and small molecules designed to inhibit 101P3A11 function and downstream signaling events are used for therapeutic diagnostic, prognostic and/or preventative purposes.

Example 42: Involvement in Tumor Progression

The 101P3A11 gene can contribute to the growth of cancer cells. The role of 101P3A11 in tumor growth is confirmed in a variety of primary and transfected cell lines including prostate, colon, bladder and kidney cell lines, as well as NIH 3T3 cells engineered to stably express 101P3A11. Parental cells lacking 101P3A11 and cells expressing sd-63712

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101P3A11 are evaluated for cell growth using a well-documented proliferation assay (Fraser SP, et al., Prostate 2000;44:61, Johnson DE, Ochieng J, Evans SL. Anticancer Drugs. 1996, 7:288). Using such a technique, we demonstrated (see Figure 37) that 101P3A11 imparts a growth advantage on NIH 3T3 cells. 3T3-neo and 3T3-101P3A11 cells were grown in 0.5% or 10% FBS and analyzed 48 hours later. The assay was performed in triplicate. Expression of 101P3A11 resulted in 6-fold increase in proliferation relative to control 3T3 cells grown in 0.5% FBS.

To confirm the role of 101P3A11 in the transformation process, its effect in colony forming assays was investigated. Parental NIH-3T3 cells lacking 101P3A11 were compared to NIH-3T3 cells expressing 101P3A11, using a soft agar assay under stringent and more permissive conditions (Song Z. et al. Cancer Res. 2000;60:6730). The results are shown in Figure 43, where 101P3A11 induces colony formation of over 100 fold increase relative to neo resistant controls. We previously showed that expression of 101P3A11 in NIH 3T3 cells induces the growth of these cells in soft agar (129-24usu1), indicating that 101P3A11 participates in the process of transformation.

To confirm the role of 101P3A11 in invasion and metastasis of cancer cells, a well-established assay is used. A non-limiting example is the use of an assay which provides a basement membrane or an analog thereof used to detect whether cells are invasive (e.g., a Transwell Insert System assay (Becton Dickinson) (Cancer Res. 1999; 59:6010)). Control cells, including prostate, colon, bladder and kidney cell lines lacking 101P3A11 are compared to cells expressing 101P3A11. Cells are loaded with the fluorescent dye, calcein, and plated in the top well of a support structure coated with a basement membrane analog (e.g. the Transwell insert) and used in the assay. Invasion is determined by fluorescence of cells in the lower chamber relative to the fluorescence of the entire cell population.

101P3A11 can also play a role in cell cycle and apoptosis. Parental cells and cells expressing 101P3A11 are compared for differences in cell cycle regulation using a well-established BrdU assay (Abdel-Malek ZA. J Cell Physiol. 1988, 136:247). In short, cells are grown under both optimal (full serum) and limiting (low serum) conditions are labeled with BrdU and stained with anti-BrdU Ab and propidium iodide. Cells are analyzed for entry into the G1, S, and G2M phases of the cell cycle. Alternatively, the effect of stress on apoptosis is evaluated in control parental cells and cells expressing 101P3A11, including normal and tumor prostate, colon and lung cells. Engineered and parental cells are treated with various chemotherapeutic agents, such as etoposide, flutamide, etc, and protein synthesis inhibitors, such as cycloheximide. Cells are stained with annexin V-FITC and cell death is measured by FACS analysis. The modulation of cell death by 101P3A11 can play a critical role in regulating tumor progression and tumor load.

When 101P3A11 plays a role in cell growth, transformation, invasion or apoptosis, it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes.

Example 43: Involvement in Angiogenesis

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Angiogenesis or new capillary blood vessel formation is necessary for tumor growth (Hanahan, D.; Folkman, J.; Cell. 1996, 86:353; Folkman J. Endocrinology. 1998 139:441). Several assays have been developed to measure angiogenesis *in vitro* and *in vivo*, such as the tissue culture assays endothelial cell tube formation and endothelial cell proliferation. Using these assays as well as *in vitro* neo-vascularization, the role of 101P3A11 in angiogenesis, enhancement or inhibition, is confirmed.

For example, endothelial cells engineered to express 101P3A11 are evaluated using tube formation and proliferation assays. The effect of 101P3A11 is also confirmed in animal models *in vivo*. For example, cells either

expressing or lacking 101P3A11 are implanted subcutaneously in immunocompromised mice. Endothelial cell migration and angiogenesis are evaluated 5-15 days later using immunohistochemistry techniques. 101P3A11 affects angiogenesis, and it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes

Example 44: Regulation of Transcription

The cell surface localization of 101P3A11 and its similarity to GPCRs indicate that 101P3A11 is effectively used as a modulator of the transcriptional regulation of eukaryotic genes. Regulation of gene expression is confirmed, e.g., by studying gene expression in cells expressing or lacking 101P3A11. For this purpose, two types of experiments are performed.

In the first set of experiments, RNA from parental and 101P3A11-expressing cells are extracted and hybridized to commercially available gene arrays (Clontech) (Smid-Koopman E et al. Br J Cancer. 2000. 83:246). Resting cells as well as cells treated with FBS or androgen are compared. Differentially expressed genes are identified in accordance with procedures known in the art. The differentially expressed genes are then mapped to biological pathways (Chen K et al. Thyroid. 2001. 11:41.).

In the second set of experiments, specific transcriptional pathway activation is evaluated using commercially available (Stratagene) luciferase reporter constructs including: NFkB-luc, SRE-luc, ELK1-luc, ARE-luc, p53-luc, and CRE-luc. These transcriptional reporters contain consensus binding sites for known transcription factors that lie downstream of well-characterized signal transduction pathways, and represent a good tool to ascertain pathway activation and screen for positive and negative modulators of pathway activation.

Thus, 101P3A11 plays a role in gene regulation, and it is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes.

Example 45: Involvement in Cell Adhesion

Cell adhesion plays a critical role in tissue colonization and metastasis. 101P3A11 can participate in cellular organization, and as a consequence cell adhesion and motility. To confirm that 101P3A11 regulates cell adhesion, control cells lacking 101P3A11 are compared to cells expressing 101P3A11, using techniques previously described (see, e.g., Haier et al, Br. J. Cancer. 1999, 80:1867; Lehr and Pienta, J. Natl. Cancer Inst. 1998, 90:118). Briefly, in one embodiment, cells labeled with a fluorescent indicator, such as calcein, are incubated on tissue culture wells coated with media alone or with matrix proteins. Adherent cells are detected by fluorimetric analysis and percent adhesion is calculated. In another embodiment, cells lacking or expressing 101P3A11 are analyzed for their ability to mediate cell-cell adhesion using similar experimental techniques as described above. Both of these experimental systems are used to identify proteins, antibodies and/or small molecules that modulate cell adhesion to extracellular matrix and cell-cell interaction. Cell adhesion plays a critical role in tumor growth, progression, and, colonization, and 101P3A11 is involved in these processes. Thus, it serves as a diagnostic, prognostic, preventative and/or therapeutic modality.

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Example 46: Protein-Protein Association

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Several GPCRs have been shown to interact with other proteins, thereby regulating signal transduction, gene transcription, transformation and cell adhesion (Sexton PM et al, Cell Signal. 2001, 13:73; Turner CE, J Cell Sci. 2000, 23:4139). Using immunoprecipitation techniques as well as two yeast hybrid systems, proteins are identified that associate with 101P3A11. Immunoprecipitates from cells expressing 101P3A11 and cells lacking 101P3A11 are compared for specific protein-protein associations.

Studies are performed to confirm the extent of association of 101P3A11 with effector molecules, such as receptors, adaptor proteins and paxillin, kinases, phsophates and Ga proteins. Studies comparing 101P3A11 positive and 101P3A11 negative cells as well as studies comparing unstimulated/resting cells and cells treated with epithelial cell activators, such as cytokines, growth factors, androgen and anti-integrin Ab reveal unique interactions.

In addition, protein-protein interactions are confirmed using two yeast hybrid methodology (Curr Opin Chem Biol. 1999, 3:64). A vector carrying a library of proteins fused to the activation domain of a transcription factor is introduced into yeast expressing a 101P3A11-DNA-binding domain fusion protein and a reporter construct. Protein-protein interaction is detected by colorimetric reporter activity. Specific association with effector molecules and transcription factors directs one of skill to the mode of action of 101P3A11, and thus identifies therapeutic, prognostic, preventative and/or diagnostic targets for cancer. This and similar assays are also used to identify and screen for small molecules that interact with 101P3A11.

Thus it is found that 101P3A11 associates with proteins and small molecules. Accordingly, 101P3A11 and these proteins and small molecules are used for diagnostic, prognostic, preventative and/or therapeutic purposes.

Example 47: Biological effect of anti-101P3A11 antibodies.

The generation of anti-101P3A11 polyclonal Ab (pAb) using an amino-terminal peptide encoding amino acids 1-14 (MVDPNGNESSATYF; SEQ ID NO: \(\frac{YY2880}{Y}\) as antigen was reported in our Priority Application. The effect of this antibody on 101P3A11 mediated ERK phosphorylation (Figure 38) and cAMP accumulation (Figure 39) was determined. 293T cells were transfected with control or 101P3A11 cDNA. Cells were allowed to rest overnight, and treated with anti-101P3A11 or control Ab in the presence of 0.5% or 10% FBS. Cells were lysed and analyzed by Western blotting with anti-Phospho-ERK and anti-ERK mAb. Figure 38 shows that expression of 101P3A11 induces ERK phosphorylation in cells treated with 0.5 or 10% FBS. Anti-101P3A11 pAb reduced the phosphorylation of ERK in 293T-101P3A11 cells treated with 0.5% FBS. The ERK overlay demonstrated equal loading, supporting the specificity of this data.

In order to confirm that anti-101P3A11 pAb has a detectable effect on cAMP accumulation, PC3 and PC3-101P3A11 cells were grown in 0.1% FBS and treated with anti-101P3A11 pAb. Cells were analyzed for cAMP content as described in Figure 32. Expression of 101P3A11 induced a 5-fold increase in cAMP accumulation in PC3 cells, which was partially inhibited by PTX. Treatment of PC3-101P3A11 cells with anti-101P3A11 pAb resulted in a 4-fold increase in cAMP accumulation in PC3-101P3A11 but not control PC3 cells. Results shown in Figure 38 and Figure 39 indicate that anti-101P3A11 pAb produces a measurable biological effect in cells expressing 101P3A11. Accordingly, 101P3A11 is used as a target for diagnostic, prognostic, preventative and/or therapeutic purposes.

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Example 48: 101P3A11 Promoters

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A eukaryotic cells promoter is a short DNA sequence located in the 5' region of a gene. It provides binding sites for RNA polymerase and associated transcriptional cofactors, which upon assembly promotes transcription of the gene. In humans, most genes are transcribed by RNA polymerase II. The promoter DNA sequence normally contains binding motifs for RNA polymerases and its associated cofactors and activators including a TATA-box, cap signal, CCAAT-box and the GC-box. A eukaryotic cell enhancer is DNA sequence where transcriptional factors and their associated coactivators or suppressors bind and interact with promoter-bound RNA polymerase to regulate the expression of the gene located next to the promoter. While a promoter(s) locates close to the transcription starting site(s) of a gene (usually 25-30 base pairs), enhancers can be found up to 50,000 base pairs in either direction to the transcription starting site(s) of the gene. There are many different gene regulatory proteins, namely transcription factors and their associated coactivators and cosuppressors that bind to specific enhancer sequences and regulate gene expression. These proteins, upon interaction with specific DNA regulatory sequences and with each other, allow each gene to be regulated up or down in different tissues and cell types. Chapter 9. "Control of gene expression" in Molecular Biology of the Cell. 3rd ed. Ed. by Alberts et al., (New York and London) Garland Publishing, 1994).

Tissue specific gene expression is associated with the presence of specific combinations of transcription factors and their associated coactivators and suppressors, the presence of specific binding sites present in the DNA regulatory region of the gene for these factors, and the activation or inactivation of signaling pathways that modulate their relative activity. For example, prostate specific expression of prostate specific antigen, (PSA, or human kallikrein 3), is dependent on the presence of androgen receptor binding elements in defined 5' upstream enhancer and promoter sequences of the gene and intact androgen receptor signaling pathway (Pang S, et al., Cancer Res 1997 Feb 1; 57(3):495-9). It is also dependent on the presence of other cis-acting DNA regulatory elements in the promoter region (Zhang J, et al., Nucleic Acids Res. 1997, Aug. 1; 25(15):3143-50.) and on the expression of other transcription factors, such as the prostate specific ets-like transcription factor (Oettgen P, et al., J Biol Chem. 2000, Jan. 14; 275(2):1216-25).

With the accumulation of data and knowledge on human gene expression, promoters and enhancers are identified using different algorithms and computer programs (Chapter 8 "Gene Prediction" in Bioinformatics - Sequence and Genome Analysis, ed. by David W. Mount. Cold Spring Harbor Laboratory Press, 2001). Accordingly, we identified (Table XXX) promoters in a 5.04 kB 5' upstream genomic region of the 101P3A11 coding sequence using Neural Network Promoter Prediction computer program (http://www.fruitfly.org/seq_tools/nnppAbst.html; Reese, M.G. and Eeckman, F.H. (1995) Novel Neural Network Algorithms for Improved Eukaryotic Promoter Site Recognition. Accepted talk for The Seventh International Genome Sequencing and Analysis Conference, Hyatt Regency, Hilton Head Island, South Carolina, September 16-20, 1995), indicated by the underlined sequences in Table XXIX. Using a program called SIGNAL SCAN (http://bimas.dcrt.nih.gov/molbio/signal/; Prestridge, D.S. (1991) SIGNAL SCAN: A computer program that scans DNA sequences for eukaryotic transcriptional elements. CABIOS 7, 203-206.), which searches a comprehensive database of regulatory element binding sites, we found numerous transcriptional binding sites for various known transcription factors in the 5.04 kB sequence 5' to the 101P3A11 gene, suggesting the presence of specific enhancer regions that may mediate tissue specific 101P3A11 transcription. Such transcription factors include, but are not limited to, NFAT, NF-1, NF-E, CP2, AP1, AP-2, Spl, OCT-1, OCT-2, NFKB, CREB, CTF, TFIIA, TFIIB, TFIID, Pit-1, C/EBP, SRF, and various steroid receptors, such as glucocorticoid receptor (GR) and androgen receptors

(AR) (Mitchell P J and Tijan R (1989) Science 245: 371). Comparison of the 5 kB upstream sequence of the 101P3A11 gene to the 5 kB upstream sequence of the PSA gene, 5 homologous regions were found that are important for prostate cell specific expression. Figure XXXI shows the alignment of the these sequences and also indicates predicted transcription factor binding sites common to both sequences identified using SIGNAL SCAN.

Experimentally, one defines the regions in the 5' genomic upstream regions of the 101P3A11 gene using various methods well known in the art, such as deletion and mutation analysis of the putative regulatory regions fused to a transcriptional reporter gene such as luciferase or chloramphenicol acetyl-transferase. These transcriptional reporter vectors are then introduced into cell lines, tissues, or transgenic animals to analyze the tissue and cell type specificity of transcription and expression of the reporter gene. To identify transcription factors and proteins that interact with specific 101P3A11 transcriptional regulatory sequences, one employs one or more of various techniques known in the art such as DNAse footprinting, gel mobility shift assays, and DNA/protein affinity chromatography. Various techniques concerning use of promoters are set forth, e.g., U.S. Patent 5,919,652 which concerns embodiments of nucleic acid compositions that comprise prostate specific antigen (PSA) promoter alone or in combination with a cytomegalovirus (CMV) promoter and related uses.; and, U.S. Patent 6,110,702 which concerns PSA positive regulatory sequence (PSAR) and related uses.

Once regulatory sequences are identified that mediate 101P3A11 tissue-specific expression, these sequences are employed in various gene therapeutic strategies for cancer, such as driving tissue-specific expression of a toxic gene or a cell suicide gene. Such cell suicide strategies are currently employed using the PSA-promoter enhancer using the thymidine kinase/ganciclovir system (Suzuki S, Tadakuma T, Asano T, Hayakawa M. Cancer Res. 2001 Feb 15;61(4):1276-90). Unlike PSA, which is an androgen regulated gene, 101P3A11 does not exhibit androgen regulated expression. Thus, identification and use of regulatory sequences of the 101P3A11 gene that mediate, e.g., prostate-specific, but androgen insensitive gene expression, is useful for the treatment of both early stage androgen sensitive and late stage androgen insensitive or hormone refractory prostate cancer.

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Throughout this application, various website data content, publications, patent applications and patents are referenced. (Websites are referenced by their Uniform Resource Locator, or URL, addresses on the World Wide Web.) The disclosures of each of these references are hereby incorporated by reference herein in their entireties.

The present invention is not to be limited in scope by the embodiments disclosed herein, which are intended as single illustrations of individual aspects of the invention, and any that are functionally equivalent are within the scope of the invention. Various modifications to the models and methods of the invention, in addition to those described herein, will become apparent to those skilled in the art from the foregoing description and teachings, and are similarly intended to fall within the scope of the invention. Such modifications or other embodiments can be practiced without departing from the true scope and spirit of the invention.

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TABLES

5 TABLE I: Exemplary Tissues that Express 101P3A11

Normal Tissues:

Prostate

10 Ovary (by RT-PCR only)

Malignant Tissues:

15 Rectum

Prostate

Colon

Kidney

Breast

20 Uterus

Cervix

Stomach

Metastases Pool

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TABLE II: AMINO ACID ABBREVIATIONS

SINGLE LETTER	THREE LETTER	FULL NAME
F	Phe	phenylalanine
L .	Leu	leucine
S	Ser	serine
Y	Tyr	tyrosine
С	Cys	cysteine
W	Тгр	tryptophan
P	Pro	proline
Н	His	histidine
Q	Gln	glutamine
R	Arg	arginine
I	Ile	isoleucine
M	Met	methionine
T	Thr	threonine
N	Asn	asparagine
K	Lys	lysine

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V	Val	valine
A	Ala	alanine
D	Asp	aspartic acid
E	Glu	glutamic acid
G	Gly	glycine

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TABLE III: AMINO ACID SUBSTITUTION MATRIX

Adapted from the GCG Software 9.0 BLOSUM62 amino acid substitution matrix (block substitution matrix). The higher the value, the more likely a substitution is found in related, natural proteins. (See URL

5 www.ikp.unibe.ch/manual/blosum62.html)

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С
                  D
                     Ε
                       F
                          G
                             H I
                                               Ρ
                                                      R
                                                         S
                                                            Т
                                                               V
                                                                     Υ.
                                    K
                                      L
                                         Μ
                                            N
                                                   Q
               0 -2 -1 -2
                                                                0 - 3 - 2 A
                           0 -2 -1 -1 -1 -2 -1 -1 -1
                                                         1
               9 -3 -4 -2 -3 -3 -1 -3 -1 -1 -3 -3 -3 -3 -1 -1 -1 -2 -2 C
10
                  6 2 -3 -1 -1 -3 -1 -4 -3
                                             1 -1
                                                   0 -2
                                                          0 -1 -3 -4 -3 D
                     5 -3 -2 0 -3 1 -3 -2 0 -1
                                                   2
                                                      0
                                                         0 -1 -2 -3 -2 E
                        6 -3 -1 0 -3 0 0 -3 -4 -3 -3 -2 -2 -1
                                                                  1
                                                                     3 F
                           6 -2 -4 -2 -4 -3
                                             0 -2 -2 -2
                                                          0 -2 -3 -2 -3 G
                              8 -3 -1 -3 -2
                                            1 -2
                                                   0
                                                      0 -1 -2 -3 -2
15
                                  4 -3 2 1 -3 -3 -3 -2 -1
                                    5 -2 -1 0 -1
                                                   1
                                                      2
                                                          0 -1 -2 -3 -2 K
                                          2 -3 -3 -2 -2 -2 -1
                                                                1 -2 -1 L
                                          5 -2 -2
                                                   0 -1 -1 -1
                                                                1 -1 -1 M
                                             6 -2
                                                   0
                                                      0
                                                         1
                                                           \cdot 0 -3 -4 -2 N
20
                                                7 -1 -2 -1 -1 -2 -4 -3 P
                                                   5
                                                      1
                                                          0 -1 -2 -2 -1 Q
                                                      5 -1 -1 -3 -3 -2 R
                                                            1 -2 -3 -2 S
                                                             5
                                                               0 -2 -2 T
25
                                                                4 -3 -1 V
                                                                  11
                                                                     2 W
                                                                      7 Y
```

TABLE IV (A):

SUPERMOTIFS	POSITION	POSITION	POSITION
	2 (Primary Anchor)	3 (Primary Anchor)	C Terminus (Primary
			Anchor)
Al	TILVMS		FWY
A2	LIVMATQ		IVMATL
A3	VSMA <i>TLI</i>		RK
A24	YFWIVLMT		FIYWLM
B7	P		VILFMWYA
B27	RHK		FYL <i>WMIVA</i>
B44	E D		FWYLIMVA
B58	ATS		FWY <i>LIVMA</i>
B62	QLIVMP		FWYMIVLA
MOTIFS			
A1	TSM		Y
Al		DEAS	Y
A2.1	LMVQIAT		VLIMAT
`A3	LMVISATFCGD		KYR <i>HFA</i>
A11	VTMLISAGNCDF		KRYH
A24	YFWM		FLIW
A*3101	MVTALIS		RK
A*3301	MVALF/ST		RK
A*6801	AVTMSL1		RK
B*0702	P		LMFWYAIV
B*3501	P		LMFWY <i>IVA</i>
B51	P		LIVFWYAM
B*5301	P		IMFWY <i>ALV</i>
B*5401	P		ATIV <i>LMFWY</i>

Bolded residues are preferred, italicized residues are less preferred: A peptide is considered motif-bearing if it has primary anchors at each primary anchor position for a motif or supermotif as specified in the above table.

TABLE IV (B): HLA CLASS II SUPERMOTIF

10

1	6	9
W, F, Y, V, .I, L	A, V, I, L, P, C, S, T	A, V, I, L, C, S, T, M, Y

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6	MH	WDE	AVM		2	g				
∞				Q		Z				
7	MH	8	Σ	GDE	M	GRD				
1° anchor 6	VSTCPALIM		VMATSPLIC		IVMSACTPL		1° anchor 6		KRH	VMSTACPLI
5				CWD			5			
4		*	PAMQ	FD	A	Ŋ	1° anchor 4	Ω	DNQEST	
3	T			· CH	M		3			
7	M			၁	M	C	2			
1° anchor 1	FMYLIVW		MFLIVWY		MFLIVWY		1° anchor 1	LIVMFY	LIVMFAY	MFLIVWY
	preferred	deleterious	preferred	deleterious	preferred	deleterious	MOTIFS			
MOTIFS	DR4		DR1		DR7		DR3	motif a preferred	motif b preferred	DR Supermotif

Italicized residues indicate less preferred or "tolerated" residues.

TABLE IV (D):
POSITION

		-	2	3 3	POSITION 4	٠	9	7	∞	C-terminus
SUPERMOTIFS										
A1			1° Anchor TILVMS							1° Anchor FWY
A2			1° Anchor LIVMATQ			·				l° Anchor LIVMAT
A 3	preferred		1° Ancher VSMA <i>TLI</i>	YFW(4/5)			YFW (3/5)	YFW (4/5)	P (4/5)	1° Anchor RK
	deleterious	DE (3/5); P (5/5)		DE (4/5)						
A24			1° Anchor YFWIVLMT							1° Anchor FIY WLM
B7	preferred	FWY (5/5) LIVM (3/5)	1° Anchor P	FWY(4/5)					FWY (3/5)	<u>1°Anchor</u> VILF <i>MWYA</i>
	deleterious	DE (3/5); P(5/5); G(4/5); A(3/5); QN(3/5)				DE(3/5)	G(4/5)	QN(4/5)	DE(4/5)	
B27			1° Anchor RHK							1°Anchor FYLWMIVA
B44			1° Anchor ED							1° Anchor FWYLIMVA
B58			1° Anchor ATS							1° Anchor FWY <i>LIVMA</i>
B62			1° Anchor QLIVMP							1° Anchor FWY <i>MIVLA</i>
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ominio.	C-terminds		<u>hor</u>		<u>hor</u>		1°Anchor Y		l°Anchor Y				1°Anchor VLIMAT		
o	,	or C-terminus	/ <u>l°Anchor</u> Y		l°Anchor Y		۵.	K	YFW	NÒ	<u>1°Anchor</u> V <i>LIMAT</i>			RKH	<u>1°Anchor</u> KYR <i>HFA</i>
œ		•	ı YFW		DE	GP	GDE	RHK	Ð	PRHK	Ь		FYWL	DERKH	<u>a</u>
۲	,		DEQN		LIVM	PG	PASTC (RHKYF I W	PG	<u>a.</u>	A	DERKH	E.	RKH DI	
¥			ď	∢	ASTC	RHK	PA .	QNA RH		g		RKH DE	\mathcal{G}	~	YFW
				Ö		PQN	NO.		≫		*	X.			
TABLE IV (E):			YFW	¥	GSTC	DE	YFWQN	RHK	YFW	<u>a</u>	YFW			<u>d</u>	(F A
TABL	+		DEA	RHKLIVMP	1°Anchor DEAS		A	1 DE	A		STC		Ð	RKHA	PRHKYF W
"				RHK		FW	DEAQN	RHKGLIVM	1°Anchor DEAS		YFW	DERKH	LVIM	DE	YFW
·	7		1°Anchor STM		ASTCLIVM	RHKDEPYFW	1°Anchor STM		STCLIVM	RHKDEPY FW	<u>1°Anchor</u> LM <i>IVQAT</i>		<u>1°Anchor</u> LMIVQAT		I°Anchor LMVISATFCG D
			GFYW	DE	GRHK	¥	1°A S		STC	RHK	LM/				LMVIS
	-		preferred	deleterious	preferred	deleterious	YFW	GP	YFW	RHK	YFW	DEP	AYFW	DEP	RHK
			A1 pre 9-mer	dele	A1 pre 9-mer	dele	preferred	deleterious	preferred	deleterious	preferred	deleterious	preferred	deleterious	preferred
			· 6		1,6		Al 10-mer		A1 10-mer		A2.1 9-mer		A2.1 10-mer		A3

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C-terminus						1°Anchor FLIW						:			
6		1°Anchor KRYH		1°Anchor FLIW			DEA	1°Anchor RK		1°Anchor RK		1°Anchor RK		1°Anchor LMFWYAIV	
∞		<u>a</u>	ŋ	YFW	AQN		N O	AP	DE			Ь	V	PA	DE
7		YFW	Y	YFW	Ö	ď	¥	YFW	DE	AYFW		YFW		RHK	NO
9		YFW			DERHK		DE	YFW	DE					RHK	GDE
S		V		:	ÓNP	YFWP	RHK	:	ADE			YFWLIV M	RHK	RHK	DE
4		YFW		STC	g	Q.	N N	d.							DE
ю	DE	YFW			DE		GDE	YFW	DE	YFW	DE		DEG	RHK	DEP
2		<u>I°Anchor</u> VTLMISAGN <i>C</i> <i>DF</i>		1°Anchor YFWM		1°Anchor YFWM		1°Anchor MVT <i>ALIS</i>		1°Anchor MVALF <i>IST</i>	,	1°Anchor AVTMSLI		1°Anchor P	
-	DEP	V	DEP	YFWRH K	DEG			RHK	DEP		GP	YFWSTC	GP	RHKFW Y	DEQNP
	deleterious	ргебеттед	deleterious	ргебетед	deleterious	preferred	deleterious	ргебетед	deleterious	preferred	deleterious	preferred	deleterious	preferred	deleterious
		AII		A24 9-mer		A24 10-mer		A3101		A3301		A6801		B0702	

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C-terminus								
6	1°Anchor LMFWY <i>IVA</i>		1°Anchor LIVFWYAM		I ^o Anchor IMFWY <i>ALV</i>		1°Anchor ATIV <i>LMFWY</i>	
∞			FWY	GDE	FWY	DE	FWYAP	DE
7	FWY		Ð	DEQN	LIVMFW Y	RHKQN	ALIVM	QNDGE
. 9		Ö		Ö		Ð		DE
ν,		Ð	FWY	DE	FWY		LIVM	RHKDE
4			STC		STC			
ю	FWY		FWY		FWY		FWYLIVM	GDESTC
7	1°Anchor P		1°Anchor P		1°Anchor P		1°Anchor P	
-	FWYLIV M	AGP	LIVMFW Y	AGPDER HKSTC	LIVMFW Y	AGPQN	FWY	GPQNDE
	preferred	deleterious	preferred	deleterious AGPDER HKSTC	preferred	deleterions	preferred	deleterious GPQNDE
	B3501		B51		B5301		B5401	

Italicized residues indicate less preferred or "tolerated" residues. The information in this Table is specific for 9-mers unless otherwise specified.

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RANK	START POSITION	ING RESULTS -1011 SUBSEQUENCE RESIDUE	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE	SEQ.ID#
	TOSITION	LISTING	CONTAINING THIS SUBSEQUENCE)	
1	210	GLDSLLISF	25.000	1.
2	1	MVDPNGNES	5.000	2.
3	29	LAFPLCSLY	5.000	3.
4	21	GLEEAQFWL	4.500	4.
5	212	DSLLISFSY	3.750	5.
6	41	VLGNLTIIY	2.500	6.
7	245	HVCAVFIFY	2.500	7.
8	57	LHEPMYIFL	2.250	8.
9	111	GMESTVLLA	2.250	9.
10	6	GNESSATYF	2.250	10.
11	259	LSMVHRFSK	1.500	11.
12	114	STVLLAMAF	1.250	12.
13	190	ACDDIRVNV	1.000	13.
14	74	LISTSSMPK	1.000	14.
15	158	MAPLPVFIK	1.000	15.
16	78	SSMPKMLAI	0.750	16.
17	172	RSNILSHSY	0.750	17.
18	134	HATVLTLPR	0.500	18.
19	44	NLTIIYIVR	0.500	19.
20	117	LLAMAFDRY	0.500	20.
21	116	VLLAMAFDR	0.500	21.
22	70	GIDILISTS	0.500	22.
23	231	TREAQAKAF	0.450	23.
24	180	YCLHQDVMK	0.400	24.
25	137	VLTLPRVTK	0.400	25.
26	183	HQDVMKLAC	0.375	26.
27	5	NGNESSATY	0.250	27.
28	138	LTLPRVTKI	0.250	28.
29	52	RTEHSLHEP	0.225	29.
30	145	KIGVAAVVR	0.200	30.
31	56	SLHEPMYIF	0.200	31.
32	242	CVSHVCAVF	0.200	32.
33	256	FIGLSMVHR	0.200	33.
34	287	VLNPIVYGV	0.200	34.
35	156	ALMAPLPVF	0.200	35.
36	125	YVAICHPLR	0.200	36.
37	248	AVFIFYVPF	0.200	37.
38	273	LPVILANIY	0.125	38.
39	95	QFDACLLQI	0.125	39.
40	271	SPLPVILAN	0.125	40.
41	11	ATYFILIGL	0.125	41.
42	159	APLPVFIKQ	0.125	42.
43	128	ICHPLRHAT	0.100	43.
44	157	LMAPLPVFI	0.100	44.
45	283	LVPPVLNPI	0.100	45.
46	77	TSSMPKMLA	0.075	46.
47	68	LSGIDILIS	0.075	47.
48	176	LSHSYCLHQ	0.075	48.
49	9	SSATYFILI	0.075	49.

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RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
50	8	ESSATYFIL	0.075	50.
51	218	FSYLLILKT	0.075	51.
52	216	ISFSYLLIL	0.075	52.
53	90	NSTTIQFDA	0.075	53.
54	55	HSLHEPMYI	0.075	54.
55	270	DSPLPVILA	0.075	55.
56	285	PPVLNPIVY	0.062	56.
57	233	EAQAKAFGT	0.050	57.
58	203	IVIISAIGL	0.050	58.
59	188	KLACDDIRV	0.050	59.
60	97	DACLLQIFA	0.050	60.
61	19	LPGLEEAQF	0.050	61.
62	253	YVPFIGLSM	0.050	62.
63	98	ACLLQIFAI	0.050	63.
64	164	FIKQLPFCR	0.050	64.
65	33	LCSLYLIAV	0.050	65.
66	288	LNPIVYGVK	0.050	66.
67	260	SMVHRFSKR	0.050	67.
68	67	MLSGIDILI	0.050	68.
69	126	VAICHPLRH	0.050	69.
70	240	GTCVSHVCA	0.050	70.
71	54	EHSLHEPMY	0.050	71.
72	276	ILANIYLLV	0.050	72.
73	215	LISFSYLLI	0.050	73.
74	217	SFSYLLILK	0.050	74.
75	83	MLAIFWFNS	0.050	75.
76	39	IAVLGNLTI	0.050	76.
77	246	VCAVFIFYV	0.050	77.
78	162	PVFIKQLPF	0.050	78.
79	208	AIGLDSLLI	0.050	79.
80	250	FIFYVPFIG	0.050	80.
81	155	AALMAPLPV	0.050	81.
82	65	LCMLSGIDI	0.050	82.
83	10	SATYFILIG	0.050	83.
84	268	RRDSPLPVI	0.050	84.
85	92	TTIQFDACL	0.050	85.
86	181	CLHQDVMKL	0.050	86.
87	62	YIFLCMLSG	0.050	87.
88	197	NVVYGLIVI	0.050	88.
89	135	ATVLTLPRV	0.050	89.
90	100	LLQIFAIHS	0.050	90.
91	282	LLVPPVLNP	0.050	91.
92	45	LTIIYIVRT	0.050	92.
93	205	IISAIGLDS	0.050	93.
94	22	LEEAQFWLA	0.045	94.
95	297	TKEIRQRIL	0.045	95.
96	113	ESTVLLAMA	0.030	96.
97	243	VSHVCAVFI	0.030	97.
98	75	ISTSSMPKM	0.030	98.
99	34	CSLYLIAVL	0.030	99.

TABLE V:				
HLA PE	EPTIDE SCORI	NG RESULTS -1011	P3A11- A1, 9-MERS	
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
100	224	LKTVLGLTR	0.025	100.

TABLE	VI:		· · · · · · · · · · · · · · · · · · ·	
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11- A1. 10-MERS	
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
1	1	MVDPNGNESS	5.000	101.
2	157	LMAPLPVFIK	5.000	102.
3	78	SSMPKMLAIF	3.000	103.
4	40	AVLGNLTIIY	2.500	104.
5	191	CDDIRVNVVY	2.500	105.
6	57	LHEPMYIFLC	2.250	106.
7	. 22	LEEAQFWLAF	2.250	107.
8	297	TKEIRQRILR	2.250	. 108.
9	52	RTEHSLHEPM	2.250	109.
10	111	GMESTVLLAM	2.250	110.
11	287	VLNPIVYGVK	2.000	111.
12	216	ISFSYLLILK	1.500	112.
13	73	ILISTSSMPK	1.000	113.
14	258	GLSMVHRFSK	1.000	114.
15	21	GLEEAQFWLA	0.900	115.
16	113	ESTVLLAMAF	0.750	116.
17	243	VSHVCAVFIF	0.750	117.
18	284	VPPVLNPIVY	0.625	118.
19	29	LAFPLCSLYL	0.500	119.
20	70	GIDILISTSS	0.500	120.
21	116	VLLAMAFDRY	0.500	121.
22	115	TVLLAMAFDR	0.500	122.
23	210	GLDSLLISFS	0.500	123.
24	190	ACDDIRVNVV	0.500	124.
25	28	WLAFPLCSLY	0.500	125.
26	6	GNESSATYFI	0.450	126.
27	136	TVLTLPRVTK	0.400	127.
28	55	HSLHEPMYIF	0.300	128.
29	95	QFDACLLQIF	0.250	129.
30	138	LTLPRVTKIG	0.250	130.
31	128	ICHPLRHATV	0.200	131.
32	155	AALMAPLPVF	0.200	132.
33	247	CAVFIFYVPF	0.200	133.
34	283	LVPPVLNPIV	0.200	134.
35	241	TCVSHVCAVF	0.200	135.
36	18	GLPGLEEAQF	0.200	136.
37	259	LSMVHRFSKR	0.150	137.
38	43	GNLTIIYIVR	0.125	138.
39	244	SHVCAVFIFY	0.125	139.
40	209	IGLDSLLISF	0.125	140.
41	161	LPVFIKQLPF	0.125	141.
42	211	LDSLLISFSY	0.125	142.
43	76	STSSMPKMLA	0.125	143.

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HLA PEPTIDE SCORING RESULTS -101P3A11- A1, 10-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE) FIGLSMVHRF** 44 256 0.100 144. 45 LLVPPVLNPI 145. 282 0.100 46 231 TREAOAKAFG 0.090 146. 47 77 **TSSMPKMLAI** 0.075 147. 48 270 **DSPLPVILAN** 0.075 148. 49 8 **ESSATYFILI** 0.075 149. 50 68 **LSGIDILIST** 0.075 150. 51 9 **SSATYFILIG** 0.075 151. 52 272 **PLPVILANIY** 0.050 152. 53 214 LLISFSYLLI 0.050 153. 54 245 **HVCAVFIFYV** 0.050 154. 55 175 **ILSHSYCLHQ** 0.050 155. 56 289 **NPIVYGVKTK** 0.050 156. 57 41 VLGNLTIIYI 0.050 157. 58 1.05 **AIHSLSGMES** 0.050 158. 59 125 YVAICHPLRH 0.050 159. 202 60 LIVIISAIGL 0.050 160. 61 260 **SMVHRFSKRR** 0.050 161. 97 62 **DACLLQIFAI** 162. 0.050 63 56 **SLHEPMYIFL** 0.050 163. SLYLIAVLGN 64 35 0.050 164. 65 195 **RVNVVYGLIV** 0.050 165. 66 215 LISFSYLLIL 0.050 166. 67 221 LLILKTVLGL 0.050 167. 68 67 **MLSGIDILIS** 0.050 168. 69 268 RRDSPLPVIL 0.050 169. 70 271 **SPLPVILANI** 0.050 170. 71 207 **SAIGLDSLLI** 0.050 171 72 102 0.050 **QIFAIHSLSG** 172. 73 64 173. **FLCMLSGIDI** 0.050 74 208 **AIGLDSLLIS** 0.050 174. 75 204 **VIISAIGLDS** 175. 0.050 76 93 TIQFDACLLQ 0.050 176. 77 275 VILANIYLLV 177. 0.050 78 10 **SATYFILIGL** 0.050 178. 79 198 **VVYGLIVIIS** 0.050 179. 80 24 **EAQFWLAFPL** 0.050 180. 81 154 **GAALMAPLPV** 0.050 181. 82 91 **STTIQFDACL** 0.050 182. 83 228 **LGLTREAQAK** 0.050 183. 84 99 **CLLQIFAIHS** 0.050 184. LIAVLGNLTI 85 38 0.050 185. **TEHSLHEPMY** 86 53 0.050 186. 87 223 **ILKTVLGLTR** 0.050 187. 158 88 **MAPLPVFIKQ** 0.050 188. 89 250 **FIFYVPFIGL** 0.050 189. 90 180 YCLHQDVMKL 0.050 190. 91 109 **LSGMESTVLL** 191. 0.030 92 34 CSLYLIAVLG 0.030 192.

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TABLE VI:

TABLE VI:						
HLA PE	HLA PEPTIDE SCORING RESULTS –101P3A11– A1, 10-MERS					
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#		
93	225	KTVLGLTREA	0.025	193.		
94	5	NGNESSATYF	0.025	194.		
95	230	LTREAQAKAF	0.025	195.		
96	171	CRSNILSHSY	0.025	196.		
97	87	FWFNSTTIQF	0.025	197.		
98	17	IGLPGLEEAQ	0.025	198.		
99	82	KMLAIFWFNS	0.025	199.		
100	4	PNGNESSATY	0.025	200.		

TABLE	VII:			
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11- A-0201, 9-MERS	
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
1	213	SLLISFSYL	825.977	201.
2	287	VLNPIVYGV	271.948	202.
3	188	KLACDDIRV	243.432	203.
4	28	WLAFPLCSL	226.014	204.
5	246	VCAVFIFYV	215.192	205.
6	21	GLEEAQFWL	178.815	206.
7	276	ILANIYLLV	177.358	207.
8	220	YLLILKTVL	149.071	208.
9	214	LLISFSYLL	138.001	209.
10	14	FILIGLPGL	114.985	210.
11	157	LMAPLPVFI	70.450	211.
12	279	NIYLLVPPV	70.387	212.
13	108	SLSGMESTV	69.552	213.
14	82	KMLAIFWFN	54.625	214.
15	181	CLHQDVMKL	49.134	215.
16	25	AQFWLAFPL	46.480	216.
17	275	VILANIYLL	42.494	217.
18	222	LILKTVLGL	42.494	218.
19	37	YLIAVLGNL	29.382	219.
20	66	CMLSGIDIL	26.377	220.
21	118	LAMAFDRYV	25.398	221.
22	201	GLIVIISAI	23.995	222.
23	306	RLFHVATHA	18.382	223.
24	67	MLSGIDILI	17.736	224.
25	85	AIFWFNSTT	14.407	225.
26	49	YIVRTEHSL	13.512	226.
27	174	NILSHSYCL	10.868	227.
28	167	QLPFCRSNI	10.433	228.
29	60	PMYIFLCML	9.493	229.
30	101	LQIFAIHSL	8.469	230.
31	254	VPFIGLSMV	6.568	231.
32	120	MAFDRYVAI	5.605	232.
33	83	MLAIFWFNS	4.747	233.
34	198	VVYGLIVII	4.683	234.

		G RESULTS -101P3 SUBSEQUENCE	SCORE (ESTIMATE OF HALF TIME OF	SEQ.ID#
RANK	START POSITION	RESIDUE	DISASSOCIATION OF A MOLECULE	SEQ.ID#
25		LISTING	CONTAINING THIS SUBSEQUENCE)	1 225
35	160	PLPVFIKQL	4.108	235.
36	304	ILRLFHVAT	3.659	236.
37	215	LISFSYLLI	3.658	237.
38	155	AALMAPLPV	3.574	238.
39	283	LVPPVLNPI	3.569	239.
40	98	ACLLQIFAI	3.361	240.
41	40	AVLGNLTII	3.185	241.
42	203	IVIISAIGL	3.178	242.
43	93	TIQFDACLL	2.937	243.
44	31	FPLCSLYLI	2.666	244.
45	135	ATVLTLPRV	2.222	245.
46	11	ATYFILIGL	2.184	246.
47	110	SGMESTVLL	2.115	247.
48	138	LTLPRVTKI	2.096	248.
49	253	YVPFIGLSM	. 2.000	249.
50	303	RILRLFHVA	1.969	250.
51	33	LCSLYLIAV	1.775	251.
52	38	LIAVLGNLT	1.742	252.
53	218	FSYLLILKT	1.647	253.
54	43	GNLTIIYIV	1.584	254.
55	119	AMAFDRYVA	1.471	255.
56	42	LGNLTIIYI	1.465	256.
57	208	AIGLDSLLI	1.435	257.
58	200	YGLIVIISA	1.270	258.
59	281	YLLVPPVLN	1.268	259.
60	92	TTIQFDACL	1.127	260.
61	7	NESSATYFI	1.116	261.
62	241	TCVSHVCAV	1.044	262.
_63	197	NVVYGLĪVĪ	0.861	263.
64	216	ISFSYLLIL	0.827	264.
65	190	ACDDIRVNV	0.745	265.
66	104	FAIHSLSGM	0.730	266.
67	243	VSHVCAVFI	0.637	267.
68	78	SSMPKMLAI	0.580	268.
69	116	VLLAMAFDR	0.544	269.
70	111	GMESTVLLA	0.528	270.
71	272	PLPVILANI	0.528	271.
72	34	CSLYLIAVL	0.487	272.
73	100	LLQIFAIHS	0.481	273.
74	69	SGIDILIST	0.459	274.
75	142	RVTKIGVAA	0.435	275.
76	150	AVVRGAALM	0.435	276.
77	65	LCMLSGIDI	0.428	277.
78	250	FIFYVPFIG	0.415	278.
79	45	LTIIYIVRT	0.405	279.
80	62	YIFLCMLSG	0.401	280.
81	112	MESTVLLAM	0.378	281.
82	168	LPFCRSNIL	0.360	282.
83	84	LAIFWFNST	0.334	283.

TABLE	VII:			
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11- A-0201, 9-MERS	
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
84	226	TVLGLTREA	0.322	284.
85	153	RGAALMAPL	0.321	285.
86	284	VPPVLNPIV	0.316	286.
87	196	VNVVYGLIV	0.316	287.
88	127	AICHPLRHA	0.314	288.
89	149	AAVVRGAAL	0.297	289.
90	207	SAIGLDSLL	0.297	290.
91	76	STSSMPKML	0.297	291.
92	56	SLHEPMYIF	0.288	292.
93	206	ISAIGLDSL	0.267	293.
94	156	ALMAPLPVF	0.260	294.
95	39	IAVLGNLTI	0.246	295.
96	35	SLYLIAVLG	0.238	296.
97	17	IGLPGLEEA	0.230	297.
98	. 274	PVILANIYL	0.226 .	298.
99	186	VMKLACDDI	0.220	299.
100	99	CLLQIFAIH	0.215	300.

TABLE VIII:				
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11- A-0201, 10-MERS	
	START	SUBSEQUENCE	SCORE (ESTIMATE OF HALF TIME OF	SEQ.ID#
RANK	POSITION	RESIDUE	DISASSOCIATION OF A MOLECULE	
		LISTING	CONTAINING THIS SUBSEQUENCE)	
1	56	SLHEPMYIFL	722.583	301.
2	117	LLAMAFDRYV	494.237	302.
3	213	SLLISFSYLL	300.355	303.
4	41	VLGNLTIIYI	224.357	304.
5	156	ALMAPLPVFI	212.307	305.
6	250	FIFYVPFIGL	94.987	306.
7	275	VILANIYLLV	90.231	307.
8	221	LLILKTVLGL	83.527	308.
9	100	LLQIFAIHSL	83.527	309.
10	139	TLPRVTKIGV	69.552	310.
11	253	YVPFIGLSMV	64.388	311.
12	94	IQFDACLLQI	62.741	312.
13	245	HVCAVFIFYV	57.690	313.
14	62	YIFLCMLSGI	56.155	314.
15	64	FLCMLSGIDI	47.991	315.
16	248	AVFIFYVPFI	42.727	316.
17	137	VLTLPRVTKI	40.792	317.
18	282	LLVPPVLNPI	40.792	318.
19	37	YLIAVLGNLT	34.279	319.
20	237	KAFGTCVSHV	28.772	320.
21	66	CMLSGIDILI	27.879	321.
22	234	AQAKAFGTCV	26.797	322.
23	214	LLISFSYLLI	26.604	323.
24	82	KMLAIFWFNS	26.114	324.
25	83	MLAIFWFNST	24.070	325.

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HLA PEPTIDE SCORING RESULTS -101P3A11- A-0201, 10-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE)** 26 21 **GLEEAQFWLA** 18.576 326. 27 44 **NLTIIYIVRT** 17.140 327. 218 28 **FSYLLILKTV** 15.371 328. 29 303 **RILRLFHVAT** 14.407 329. 30 142 RVTKIGVAAV 13.997 330. 31 166 **KQLPFCRSNI** 13.698 331. 181 32 **CLHQDVMKLA** 11.426 332. 33 119 **AMAFDRYVAI** 11.302 333. 34 29 LAFPLCSLYL 10.264 334. 108 35 **SLSGMESTVL** 8.759 335. 36 167 **QLPFCRSNIL** 8.759 336. 227 37 **VLGLTREAQA** 8.446 337. 38 301 **RQRILRLFHV** 7.149 338. 39 189 LACDDIRVNV 6.733 339. 40 205 **IISAIGLDSL** 5.628 340. 41 180 YCLHQDVMKL 5.459 341. 42 85 **AIFWFNSTTI** 5.308 342. 43 242 **CVSHVCAVFI** 5.021 343. 44 202 4.993 LIVIISAIGL 344. 45 229 4.968 **GLTREAQAKA** 345. 46 291 **IVYGVKTKEI** 4.966 346. 47 215 LISFSYLLIL 4.709 347. 48 159 **APLPVFIKQL** 4.510 348. 49 283 LVPPVLNPIV 4.242 349. 50 279 NIYLLVPPVL 3.854 350. 51 25 **AQFWLAFPLC** 3.541 351. 52 32 **PLCSLYLIAV** 3.519 352. 53 298 KEIRQRILRL 3.344 353. 54 74 LISTSSMPKM 2.671 354. 55 200 2.666 355. **YGLIVIISAI** 56 RVNVVYGLIV 195 2.495 356. 57 38 LIAVLGNLTI 2.439 357. 58 240 **GTCVSHVCAV** 2.222 358. 59 222 1.927 LILKTVLGLT 359. 120 60 **MAFDRYVAIC** 1.678 360. 61 20 **PGLEEAOFWL** 1.485 361. 62 7 **NESSATYFIL** 1.482 362. 16 63 LIGLPGLEEA 1.309 363. 64 273 LPVILANIYL 1.304 364. 65 220 YLLILKTVLG 1.268 365. 66 286 **PVLNPIVYGV** 1.139 366. 67 110 **SGMESTVLLA** 1.132 367. 68 91 **STTIQFDACL** 1.127 368. 69 99 **CLLQIFAIHS** 1.048 369. 70 278 **ANIYLLVPPV** 1.044 370. 71 127 **AICHPLRHAT** 1.025 371. 72 154 GAALMAPLPV 0.966 372. 73 89 **FNSTTIQFDA** 0.865 373. 74 197 374. **NVVYGLIVII** 0.861

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TABLE VIII:

TABLE	TABLE VIII:				
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11- A-0201, 10-MERS		
		SUBSEQUENCE	SCORE (ESTIMATE OF HALF TIME OF	SEQ.ID#	
RANK	START POSITION	RESIDUE	DISASSOCIATION OF A MOLECULE		
	POSITION	LISTING	CONTAINING THIS SUBSEQUENCE)		
75	232	REAQAKAFGT	0.840	375.	
76	128	ICHPLRHATV	0.772	376.	
77	42	LGNLTIIYIV	0.728	377.	
78	190	ACDDIRVNVV	0.702	378.	
79	10	SATYFILIGL	0.682	379.	
80	84	LAIFWFNSTT	0.669	380.	
81	187	MKLACDDIRV	0.608	381.	
82	281	YLLVPPVLNP	0.583	382.	
83	271	SPLPVILANI	0.580	383.	
84	35	SLYLIAVLGN	0.548	384.	
85	111	GMESTVLLAM	0.528	385.	
86	148	VAAVVRGAAL	0.504	386.	
87	162	PVFIKQLPFC	0.448	387.	
88	116	VLLAMAFDRY	0.436	388.	
89	150	AVVRGAALMA	0.435 .	389.	
90	65	LCMLSGIDIL	0.405	390.	
91	212	DSLLISFSYL	0.404	391.	
92	112	MESTVLLAMA	0.378	392.	
93	210	GLDSLLISFS	0.377	393.	
94 .	28	WLAFPLCSLY	0.343	394.	
95	31	FPLCSLYLIA	0.339	395.	
96	59	EPMYIFLCML	0.338	396.	
97	92	TTIQFDACLL	0.297	397.	
98	79	SMPKMLAIFW	0.296	398.	
99	306	RLFHVATHAS	0.276	399.	
100	109	LSGMESTVLL	0.237	400.	

TABLE	IX:			
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11– A3, 9-MERS	
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
1	229	GLTREAQAK	60.000	401.
2	137	VLTLPRVTK	30.000	402.
3	56	SLHEPMYIF	20.250	403.
4	260	SMVHRFSKR	18.000	404.
5	210	GLDSLLISF	18.000	405.
_6	116	VLLAMAFDR	18.000	406.
7	44	NLTIIYIVR	12.000	407.
8	117	LLAMAFDRY	12.000	408.
9	21	GLEEAQFWL	8.100	409.
10	214	LLISFSYLL	8.100	410.
11	41	VLGNLTIIY	8.000	411.
12	156	ALMAPLPVF	6.750	412.
13	201	GLIVIISAI	6.075	413.
14	287	VLNPIVYGV	4.050	414.
15	74	LISTSSMPK	4.000	415.
16	181	CLHQDVMKL	3.600	416.

RANK	START	SUBSEQUENCE RESIDUE	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE	SEQ.ID#
KANK	POSITION	LISTING	CONTAINING THIS SUBSEQUENCE)	}
17	245	HVCAVFIFY	3.600	417.
18	111	GMESTVLLA	3.600	418.
19	79	SMPKMLAIF	3.000	419.
20	248	AVFIFYVPF	3.000	420.
21	158	MAPLPVFIK	2.700	421.
22	213	SLLISFSYL	2.700	422.
23	67	MLSGIDILI	2.700	423.
24	306	RLFHVATHA	1.500	424.
25	60	PMYIFLCML	1.350	425.
26	28	WLAFPLCSL	1.350	426.
27	99	CLLQIFAIH	1.350	420.
28	66	CMLSGIDIL	1.350	427.
29	82	KMLAIFWFN	1.215	428.
30	145	KIGVAAVVR	1.213	430.
31	188	KLACDDIRV	1.200	430.
32	164	FIKQLPFCR	1.200	431
33	220	YLLILKTVL	0.900	432.
34	157	LMAPLPVFI	0.900	434.
35	11	ATYFILIGL	0.675	435.
36	198	VVYGLIVII	0.675	436.
37	37	YLIAVLGNL	0.608	437.
38	167	QLPFCRSNI	0.600	437.
39	256	FIGLSMVHR	0.600	439.
40	276	ILANIYLLV	0.600	440.
41	119	AMAFDRYVA	0.600	441.
42	186	VMKLACDDI	0.600	442.
43	261	MVHRFSKRR	0.600	443.
44	222	LILKTVLGL	0.540	444.
45	259	LSMVHRFSK	0.450	444.
46	290	PIVYGVKTK	0.450	446.
47	282	LLVPPVLNP	0.405	447.
48	275	VILANIYLL	0.405	448.
49	125	YVAICHPLR	0.400	449.
50	83	MLAIFWFNS	0.360	450.
51	304	ILRLFHVAT	0.300	451.
52	29	LAFPLCSLY	0.300	452.
53	180	YCLHQDVMK	0.300	453.
54	108	SLSGMESTV	0.300	454.
55	242	CVSHVCAVF	0.300	455.
56	279	NIYLLVPPV	0.300	456.
57	114	STVLLAMAF	0.300	457.
58	174	NILSHSYCL	0.270	458.
59	25	AQFWLAFPL	0.270	459.
60	197	NVVYGLIVI	0.270	460.
61	100	LLQIFAIHS	0.240	461.
62	296	KTKEIRQRI	0.203	462.
63	138	LTLPRVTKI	0.203	463.
64	40	AVLGNLTII	0.203	464.
65	283	LVPPVLNPI	0.203	465.

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TABLE IX:

TABLE IX:		
HI A PEPTIC	F SCORING RESULTS _101P3A11_	A3 9-MFRS

RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING		SEQ.ID#
66	175	ILSHSYCLH	0.200	466.
67	162	PVFIKQLPF	0.200	467.
68	203	IVIISAIGL	0.180	468.
69	85	AIFWFNSTT	0.150	469.
70	35	SLYLIAVLG	0.150	470.
71	47	IIYIVRTEH	0.150	471.
72	14	FILIGLPGL	0.135	472.
73	92	TTIQFDACL	0.135	473.
74	272	PLPVILANI	0.135	474.
75	216	ISFSYLLIL	0.135	475.
76	160	PLPVFIKQL	0.135	476.
77	208	AIGLDSLLI	0.120	477.
78	215	LISFSYLLI	0.120	478.
79	298	KEIRQRILR	0.108	479.
80	18	GLPGLEEAQ	. 0.090	480.
81	49	YIVRTEHSL	0.090	481.
82	303	RILRLFHVA	0.090	482.
83	134	HATVLTLPR	0.080	483.
84	223	ILKTVLGLT	0.068	484.
85	217	SFSYLLILK	0.060	485.
86	253	YVPFIGLSM	0.060	486.
87	273	LPVILANIY	0.060	487.
88	93	TIQFDACLL	0.060	488.
89	151	VVRGAALMA	0.060	489.
90	9	SSATYFILI	0.054	490.
91	299	EIRQRILRL	0.054	491.
92	237	KAFGTCVSH	0.045	492.
93	281	YLLVPPVLN	0.045	493.
94	250	FIFYVPFIG	0.045	494.
95	98	ACLLQIFAI	0.041	495.
96	212	DSLLISFSY	0.041	496.
97	31	FPLCSLYLI	0.041	497.
98	101	LQIFAIHSL	0.041	498.
99	32	PLCSLYLIA	0.040	499.
100	45	LTIIYIVRT	0.034	500.

TABLE X:						
HLA PE	PTIDE SCORIN	NG RESULTS -101P3	A11- A3, 10-MERS			
RANK START POSITION SUBSEQUENCE RESIDUE DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)						
1	157	LMAPLPVFIK	405.000	501.		
2	258	GLSMVHRFSK	180.000	502.		
3	73	ILISTSSMPK	60.000	503.		
4	116	VLLAMAFDRY	18.000	504.		
5	287	VLNPIVYGVK	13.500	505.		
6	18	GLPGLEEAQF	9.000	506.		
7	260	SMVHRFSKRR	9.000	507.		

HLA PEPTIDE SCORING RESULTS -101P3A11- A3, 10-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE)** 8 213 **SLLISFSYLL** 8.100 508. 9 21 **GLEEAQFWLA** 8.100 509. 223 10 **ILKTVLGLTR** 8.000 510. 221 LLILKTVLGL 11 5.400 511. 136 TVLTLPRVTK 4.500 512. 12 WLAFPLCSLY 13 28 4.000 513. 14 186 **VMKLACDDIR** 4.000 514. 15 282 LLVPPVLNPI 3.038 515. 16 66 **CMLSGIDILI** 2.700 516. 248 17 **AVFIFYVPFI** 2.700 517. 18 111 **GMESTVLLAM** 2.700 518. 19 250 **FIFYVPFIGL** 2.700 519. 20 115 TVLLAMAFDR 1.800 520. 21 137 **VLTLPRVTKI** 1.800 521. 22 40 **AVLGNLTIIY** 1.800 522. 23 56 523. SLHEPMYIFL 1.800 24 214 **LLISFSYLLI** 1.800 524. 25 41 **VLGNLTIIYI** 1.800 525. 26 82 **KMLAIFWFNS** 1.620 526. 27 216 **ISFSYLLILK** 1.500 527. 28 64 1.200 **FLCMLSGIDI** 528. 29 100 **LLQIFAIHSL** 0.900 529. 30 108 SLSGMESTVL 0.900 530. 31 156 0.900 **ALMAPLPVFI** 531. 32 83 **MLAIFWFNST** 0.900 532. 33 256 **FIGLSMVHRF** 0.600 533. 34 304 **ILRLFHVATH** 0.600 534. 35 119 **AMAFDRYVAI** 0.600 535. 36 167 **QLPFCRSNIL** 0.600 536. 37 35 **SLYLIAVLGN** 0.600 537. 229 38 **GLTREAQAKA** 0.600 538. 39 215 LISFSYLLIL 0.540 539. 40 44 **NLTIIYIVRT** 0.450 540. 41 279 NIYLLVPPVL 0.450 541. 42 62 **YIFLCMLSGI** 0.450 542. 43 289 **NPIVYGVKTK** 0.450 543. 44 139 **TLPRVTKIGV** 0.400 544. 79 45 **SMPKMLAIFW** 0.400 545. 272 46 **PLPVILANIY** 0.400 546. 47 **PLRHATVLTL** 0.360 131 547. 99 48 **CLLQIFAIHS** 0.360 548. 49 85 **AIFWFNSTTI** 0.300 549. 50 94 **IQFDACLLQI** 0.270 550. 281 51 YLLVPPVLNP 0.270 551. 52 198 **VVYGLIVIIS** 0.270 552. 53 245 **HVCAVFIFYV** 0.270 553. 193 0.243 54 DIRVNVVYGL 554. 55 181 **CLHQDVMKLA** 0.225 555. 291 **IVYGVKTKEI** 56 0.225 556.

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TABLE X:

RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
57	306	RLFHVATHAS	0.200	557.
58	227	VLGLTREAQA	0.200	558.
59	32	PLCSLYLIAV	0.180	559.
60	294	GVKTKEIRQR	0.180	560.
61	210	GLDSLLISFS	0.180	561.
62	202	LIVIISAIGL	0.180	562.
63	67	MLSGIDILIS	0.180	563.
64	276	ILANIYLLVP	0.180	564.
65	37	YLIAVLGNLT	0.150	565.
66	15	ILIGLPGLEE	0.135	566.
67	197	NVVYGLIVII	0.135	567.
68	237	KAFGTCVSHV	0.135	568.
69	201	GLIVIISAIG	0.135	569.
70	166	KQLPFCRSNI	0.121	570.
71	175	ILSHSYCLHQ	0.120	. 571.
72	195	RVNVVYGLIV	0.120	572.
73	38	LIAVLGNLTI	0.120	573.
74	43	GNLTIIYIVR	0.108	574.
75	55	HSLHEPMYIF	0.101	575.
76	25	AQFWLAFPLC	0.090	576.
77	240	GTCVSHVCAV	0.090	577.
78	29	LAFPLCSLYL	0.090	578.
79	259	LSMVHRFSKR	0.090	579.
80	275	VILANIYLLV	0.090	580.
81	120	MAFDRYVAIC	0.090	581.
82	243	VSHVCAVFIF	0.090	582.
83	91	STTIQFDACL	0.090	583.
84	247	CAVFIFYVPF	0.090	584.
85	205	IISAIGLDSL	0.090	585.
86	80	MPKMLAIFWF	0.090	586.
87	284	VPPVLNPIVY	0.080	587.
88	230	LTREAQAKAF	0.075	588.
89	155	AALMAPLPVF	0.068	589.
90	242	CVSHVCAVFI	0.060	590.
91	161	LPVFIKQLPF	0.060	591.
92	117	LLAMAFDRYV	0.060	592.
93	150	AVVRGAALMA	0.060	593.
94	142	RVTKIGVAAV	0.060	594.
95	244	SHVCAVFIFY	0.054	595.
96	180	YCLHQDVMKL	0.054	596.
97	46	TIIYIVRTEH	0.045	597.
98	241	TCVSHVCAVF	0.045	598.
99	92	TTIQFDACLL	0.045	599.
100	303	RILRLFHVAT	0.045	600.

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HLA PEPTIDE SCORING RESULTS -101P3A11- A-1101, 9-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE)** 1 229 **GLTREAQAK** 1.200 601. 2 74 LISTSSMPK 0.800 602. 158 3 **MAPLPVFIK** 0.600 603. 4 217 **SFSYLLILK** 0.400 604. 5 125 YVAICHPLR 0.400 605. VLTLPRVTK 6 137 0.400 606. 7 116 **VLLAMAFDR** 0.360 607. 8 180 YCLHQDVMK 0.300 608. 9 145 **KIGVAAVVR** 0.240 609 10 164 **FIKOLPFCR** 0.240 610. 11 261 **MVHRFSKRR** 0.200 611. 12 44 **NLTIIYIVR** 0.160 612. 259 13 **LSMVHRFSK** 0.120 613. 14 260 **SMVHRFSKR** 0.120 614. 298 15 KEIRQRILR 0.108 615. 256 16 **FIGLSMVHR** 0.080 616. 17 134 **HATVLTLPR** 0.080 617. 18 142 **RVTKIGVAA** 0.060 618. 19 197 NVVYGLIVI 0.060 619. 20 203 **IVIISAIGL** 0.060 620. 21 195 RVNVVYGLI 0.060 621. 0.040 22 151 **VVRGAALMA** 622. 23 245 **HVCAVFIFY** 0.040 623. 24 11 **ATYFILIGL** 0.040 624. 25 198 **VVYGLIVII** 0.040 625. 248 26 **AVFIFYVPF** 0.040 626. 27 288 LNPIVYGVK 0.040 627. 28 253 **YVPFIGLSM** 0.040 628. 29 30 i RQRILRLFH 0.036 629. 30 21 630. **GLEEAQFWL** 0.036 31 25 **AOFWLAFPL** 0.036 631. 32 290 PIVYGVKTK 0.030 632. 33 240 **GTCVSHVCA** 0.030 633. 34 296 KTKEIRQRI 0.030 634. 35 40 **AVLGNLTII** 0.030 635. 150 36 0.030 AVVRGAALM 636. 37 114 **STVLLAMAF** 0.030 637. 38 306 **RLFHVATHA** 0.024 638. 39 188 **KLACDDIRV** 0.024 639. 40 210 **GLDSLLISF** 0.024 640. 41 111 **GMESTVLLA** 0.024 641. 42 50 **IVRTEHSLH** 0.020 642. 43 242 **CVSHVCAVF** 0.020 643. 44 283 LVPPVLNPI 0.020 644. 45 303 **RILRLFHVA** 0.018 645. 46 124 RYVAICHPL 0.018 646. 47 201 **GLIVIISAI** 0.018 647. 48 92 TTIQFDACL 0.015 648.

0.015

649.

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ATVLTLPRV

135

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TABLE XI:

HLA PEPTIDE SCORING RESULTS -101P3A11- A-1101, 9-MERS **SUBSEQUENCE** SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** CONTAINING THIS SUBSEQUENCE) LISTING 650. 50 138 LTLPRVTKI 0.015 KAFGTCVSH 51 237 0.012 651. 52 174 0.012 652. NILSHSYCL 53 214 LLISFSYLL 0.012 653. 54 275 VILANIYLL 0.012 654. 55 222 LILKTVLGL 0.012 655. 56 294 **GVKTKEIRQ** 0.012 656. 57 143 VTKIGVAAV 0.010 657. 58 230 0.010 **LTREAQAKA** 658. 59 0.009 101 659. LQIFAIHSL 60 98 **ACLLQIFAI** 0.009 660. 47 61 **IIYIVRTEH** 0.008 661. 279 62 NIYLLVPPV 0.008 662. 0.008 63 56 **SLHEPMYIF** 663. 64 65 **LCMLSGIDI** 0.008 664. 65 67 **MLSGIDILI** 0.008 665. 66 208 **AIGLDSLLI** 0.008 666. 67 224 **LKTVLGLTR** 0.008 667. 181 0.008 68 **CLHQDVMKL** 668. 69 276 0.008 **ILANIYLLV** 669. 70 287 **VLNPIVYGV** 0.008 670. 71 156 **ALMAPLPVF** 0.008 671. 72 215 LISFSYLLI 0.008 672. 73 251 **IFYVPFIGL** 0.008 673. 74 119 **AMAFDRYVA** 0.008 674. 75 41 **VLGNLTIIY** 0.008 675. 76 162 **PVFIKQLPF** 0.008 676. 77 187 MKLACDDIR 0.006 677. YGVKTKEIR 78 293 0.006 678. 79 213 679. **SLLISFSYL** 0.006 80 147 0.006 **GVAAVVRGA** 680. 81 39 **IAVLGNLTI** 0.006 681. 82 220 YLLILKTVL 0.006 682. 83 155 **AALMAPLPV** 0.006 683. 84 274 **PVILANIYL** 0.006 684. 85 0.006 66 **CMLSGIDIL** 685. 86 31 **FPLCSLYLI** 0.006 686. 87 VAICHPLRH 126 0.006 687. 88 14 **FILIGLPGL** 0.006 688. 89 37 YLIAVLGNL 0.006 689. 90 99 **CLLQIFAIH** 0.006 690. 91 280 **IYLLVPPVL** 0.006 691. 92 49 YIVRTEHSL 0.006 692. 93 246 **VCAVFIFYV** 0.006 693. 94 76 STSSMPKML 0.005 694. 225 95 KTVLGLTRE 0.005 695. 96 179 **SYCLHODVM** 0.004 696. 97 29 **LAFPLCSLY** 0.004 697. 98 167 **QLPFCRSNI** 0.004 698.

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TABLE XI:

TABLE XI:							
HLA PE	HLA PEPTIDE SCORING RESULTS -101P3A11- A-1101, 9-MERS						
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#			
99	28	WLAFPLCSL	0.004	699.			
100	117	LLAMAFDRY	0.004	700.			

HLA PEPTIDE SCORING RESULTS -101P3A11- A-1101, 10-MERS					
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#	
1	258	GLSMVHRFSK	3.600	701.	
2	136	TVLTLPRVTK	3.000	702.	
3	115	TVLLAMAFDR	1.800	703.	
4	157	LMAPLPVFIK	1.200	704.	
5	73	ILISTSSMPK	1.200	705.	
6	287	VLNPIVYGVK	0.400	706.	
7	179	SYCLHQDVMK	. 0.400	707.	
8	124	RYVAICHPLR	0.360	708.	
9	163	VFIKQLPFCR	0.180	709.	
10	223	ILKTVLGLTR	0.160	710.	
11	289	NPIVYGVKTK	0.150	711.	
12	294	GVKTKEIRQR	0.120	712.	
13	195	RVNVVYGLIV	0.120	713.	
14	292	VYGVKTKEIR	0.080	714.	
15	216	ISFSYLLILK	0.080	715.	
16	186	VMKLACDDIR	0.080	716.	
17	43	GNLTIIYIVR	0.072	717.	
18	147	GVAAVVRGAA	0.060	718.	
19	245	HVCAVFIFYV	0.060	719.	
20	142	RVTKIGVAAV	0.060	720.	
21	150	AVVRGAALMA	0.060	721.	
22	40 260	AVLGNLTIIY	0.060	722.	
24	301	SMVHRFSKRR	0.060	723.	
25	248	RQRILRLFHV AVFIFYVPFI	0.054	724.	
26	125	YVAICHPLRH	0.040	725.	
27	228	LGLTREAQAK	0.040	726. 727.	
28	52	RTEHSLHEPM	0.030	727.	
29	240	GTCVSHVCAV	0.030	729.	
30	197	NVVYGLIVII	0.030	730.	
31	166	KQLPFCRSNI	0.027	731.	
32	133	RHATVLTLPR	0.024	731.	
33	94	IQFDACLLQI	0.024	733.	
34	111	GMESTVLLAM	0.024	734.	
35	21	GLEEAQFWLA	0.024	735.	
36	291	IVYGVKTKEI	0.020	736.	
37	242	CVSHVCAVFI	0.020	737.	
38	253	YVPFIGLSMV	0.020	738.	
39	76	STSSMPKMLA	0.020	739.	
40	283	LVPPVLNPIV	0.020	740.	
41	250	FIFYVPFIGL	0.016	741.	

HLA PEPTIDE SCORING RESULTS -101P3A11- A-1101, 10-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING CONTAINING THIS SUBSEQUENCE) 42 92 TTIQFDACLL 742. 0.015 43 202 743. LIVIISAIGL 0.012 44 275 VILANIYLLV 744. 0.012 45 221 LLILKTVLGL 0.012 745. 229 46 **GLTREAQAKA** 0.012 746. 47 214 747. LLISFSYLLI 0.012 48 18 **GLPGLEEAQF** 0.012 748. 49 154 **GAALMAPLPV** 749. 0.012 50 66 **CMLSGIDILI** 0.012 750. 51 213 SLLISFSYLL 0.012 751. 52 252 **FYVPFIGLSM** 0.012 752. 53 237 KAFGTCVSHV 0.012 753. 54 185 **DVMKLACDDI** 0.012 754. 55 91 **STTIOFDACL** 0.010 755. 56 143 VTKIGVAAVV 0.010 756. 57 56 **SLHEPMYIFL** 0.008 757. 58 79 **SMPKMLAIFW** 0.008 758. 59 297 **TKEIRQRILR** 0.008 759. 60 41 760. VLGNLTIIYI 0.008 118 61 LAMAFDRYVA 0.008 761. 62 62 YIFLCMLSGI 0.008 762. 63 215 LISFSYLLIL 0.008 763. 64 64 **FLCMLSGIDI** 0.008 764. 199 65 VYGLIVIISA 0.008 765. 66 156 **ALMAPLPVFI** 0.008 766. 67 139 **TLPRVTKIGV** 0.008 767. 68 85 **AIFWFNSTTI** 0.008 768. 69 38 LIAVLGNLTI 0.008 769. 70 **254 VPFIGLSMVH** 770. 0.008 71 29 771. LAFPLCSLYL 0.008 259 72 LSMVHRFSKR 0.008 772. 73 198 **VVYGLIVIIS** 0.008 773. 74 279 NIYLLVPPVL 0.008 774. 75 TKIGVAAVVR 144 775. 0.006 76 255 **PFIGLSMVHR** 0.006 776. 77 207 0.006 **SAIGLDSLLI** 777. 78 36 LYLIAVLGNL 0.006 778. 79 46 TIIYIVRTEH 0.006 779. 80 286 **PVLNPIVYGV** 0.006 780. 81 49 YIVRTEHSLH 0.006 781. 82 80 **MPKMLAIFWF** 0.006 782. 83 161 **LPVFIKQLPF** 0.006 783. 84 264 RFSKRRDSPL 0.006 784. 85 174 **NILSHSYCLH** 0.006 785. 116 0.006 86 **VLLAMAFDRY** 786. 87 219 SYLLILKTVL 0.006 787. 88 180 YCLHODVMKL 0.006 788. 89 31 **FPLCSLYLIA** 0.006 789. 48 90 **IYIVRTEHSL** 0.006 790.

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TABLE XII:

TABLE	TABLE XII:						
HLA PE	HLA PEPTIDE SCORING RESULTS -101P3A11- A-1101, 10-MERS						
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#			
91	274	PVILANIYLL	0.006	791.			
92	273	LPVILANIYL	0.006	792.			
93	282	LLVPPVLNPI	0.006	793.			
94	234	AQAKAFGTCV	0.006	794.			
95	82	KMLAIFWFNS	0.005	795.			
96	298	KEIRQRILRL	0.005	796.			
97	230	LTREAQAKAF	0.005	797.			
98	225	KTVLGLTREA	0.005	798.			
99	108	SLSGMESTVL	0.004	799.			
100	10	SATYFILIGL	0.004	800.			

TABLE	XIII:			
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11- A24 , 9-MERS	
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
11	124	RYVAICHPL	840.000	801.
2	280	IYLLVPPVL	420.000	802.
3	292	VYGVKTKEI	55.000	803.
4	30	AFPLCSLYL	30.000	804.
5	179	SYCLHQDVM	25.000	805.
6	251	IFYVPFIGL	24.000	806.
7	88	WFNSTTIQF	15.000	807.
8	219	SYLLILKTV	10.500	808.
9	153	RGAALMAPL	9.600	809.
10	252	FYVPFIGLS	9.000	810.
11	61	MYIFLCMLS	9.000	811.
12	37	YLIAVLGNL	8.400	812.
13	36	LYLIAVLGN	7.500	813.
14	249	VFIFYVPFI	7.500	814.
15	63	IFLCMLSGI	7.500	815.
16	48	IYIVRTEHS	7.500	816.
17	213	SLLISFSYL	7.200	817.
18	207	SAIGLDSLL	7.200	818.
19	220	YLLILKTVL	7.200	819.
20	34	CSLYLIAVL	7.200	820.
21	21	GLEEAQFWL	7.200	821.
22	110	SGMESTVLL	7.200	822.
23	199	VYGLIVIIS	7.000	823.
24	92	TTIQFDACL	6.000	824.
25	66	CMLSGIDIL	6.000	825.
26	174	NILSHSYCL	6.000	826.
27	49	YIVRTEHSL	6.000	827.
28	275	VILANIYLL	6.000	828.
29	149	AAVVRGAAL	6.000	829.
30	101	LQIFAIHSL	6.000	830.
31	222	LILKTVLGL	6.000	831.
32	130	HPLRHATVL	6.000	832.
33	214	LLISFSYLL	6.000	833.

HLA PEPTIDE SCORING RESULTS -101P3A11- A24, 9-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# START **RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE)** 834. 34 93 **TIQFDACLL** 6.000 35 **FILIGLPGL** 14 6.000 835. 36 203 6.000 **IVIISAIGL** 836. 37 11 ATYFILIGL 5.600 837. 38 86 **IFWFNSTTI** 5.000 838. 39 95 **QFDACLLQI** 5.000 839. 40 25 **AQFWLAFPL** 4.800 840. 41 168 **LPFCRSNIL** 841. 4.800 42 CLHODVMKL 181 4.400 842. 43 195 **RVNVVYGLI** 4.200 843. 44 296 KTKEIRORI 4.032 844. 45 265 **FSKRRDSPL** 4.000 845. 46 206 **ISAIGLDSL** 4.000 846. 47 216 **ISFSYLLIL** 4.000 847. 48 28 WLAFPLCSL 4.000 848. 49 76 STSSMPKML 4.000 849. 50 8 **ESSATYFIL** 4.000 850. 51 109 **LSGMESTVL** 4.000 851. 52 299 **EIRQRILRL** 4.000 852. 53 114 **STVLLAMAF** 3.600 853. 54 283 LVPPVLNPI 3.024 854. 55 156 **ALMAPLPVF** 3.000 855. 257 **IGLSMVHRF** 56 3.000 856. 57 **GNESSATYF** 3.000 6 857. 79 58 **SMPKMLAIF** 3.000 858. 59 242 **CVSHVCAVF** 2.800 859. 60 210 **GLDSLLISF** 2.400 860. 61 56 **SLHEPMYIF** 2.400 861. 62 201 **GLIVIISAI** 2.100 862. 63 19 2.000 **LPGLEEAQF** 863. 64 248 **AVFIFYVPF** 2.000 864. 65 138 LTLPRVTKI 1.980 865. 66 78 SSMPKMLAI 1.800 866. 67 197 **NVVYGLIVI** 1.500 867. 68 98 **ACLLQIFAI** 1.500 868. 69 42 **LGNLTIIYI** 1.500 869. 70 55 **HSLHEPMYI** 1.500 870. 71 65 **LCMLSGIDI** 1.500 871. 72 39 **IAVLGNLTI** 1.500 872. 73 167 **QLPFCRSNI** 1.500 873. 74 40 AVLGNLTII 1.500 874. 75 31 **FPLCSLYLI** 1.500 875. 76 157 **LMAPLPVFI** 1.440 876. 77 67 **MLSGIDILI** 1.400 877. 78 59 **EPMYIFLCM** 1.260 878. 79 120 **MAFDRYVAI** 1.200 879. 80 9 **SSATYFILI** 1.200 880. 81 198 VVYGLIVII 1.200 881. 269 RDSPLPVIL 82 1.152 882.

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TABLE XIII:

RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
83	253	YVPFIGLSM	1.050	883.
84	243	VSHVCAVFI	1.000	884.
85	208	AIGLDSLLI	1.000	885.
86	186	VMKLACDDI	1.000	886.
87	215	LISFSYLLI	1.000	887.
88	194	IRVNVVYGL	0.840	888.
89	72	DILISTSSM	0.750	889.
90	163	VFIKQLPFC	0.750	890.
91	104	FAIHSLSGM	0.750	891.
92	150	AVVRGAALM	0.750	892.
93	57	LHEPMYIFL	0.720	893.
94	12	TYFILIGLP	0.600	894.
95	160	PLPVFIKQL	0.600	895.
96	26	QFWLAFPLC	0.600	896.
97	297	TKEIRQRIL	0.600	897.
98	274	PVILANIYL	0.600	898.
99	75	ISTSSMPKM	0.550	899.
100	82	KMLAIFWFN	0.504	900.

TABLE	XIV:			•
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11- A24, 10-MERS	
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
1	36	LYLIAVLGNL	420.000	901.
2	219	SYLLILKTVL	360.000	902.
3	48	IYIVRTEHSL	300.000	903.
4	252	FYVPFIGLSM	63.000	904.
5	264	RFSKRRDSPL	40.000	905.
6	13	YFILIGLPGL	30.000	906.
7	95	QFDACLLQIF	14.400	907.
8	296	KTKEIRQRIL	9.600	908.
9	30	AFPLCSLYLI	7.500	909.
10	280	IYLLVPPVLN	7.500	910.
11	24	EAQFWLAFPL	7.200	911.
12	167	QLPFCRSNIL	7.200	912.
13	159	APLPVFIKQL	7.200	913.
14	212	DSLLISFSYL	7.200	914.
15	199	VYGLIVIISA	7.000	915.
16	180	YCLHQDVMKL	6.600	916.
17	65	LCMLSGIDIL	6.000	917.
18	92	TTIQFDACLL	6.000	918.
19	27	FWLAFPLCSL	6.000	919.
20	213	SLLISFSYLL	6.000	920.
21	59	EPMYIFLCML	6.000	921.
22	273	LPVILANIYL	6.000	922.
23	221	LLILKTVLGL	6.000	923.
24	202	LIVIISAIGL	6.000	924.

		SUBSEQUENCE	A11- A24, 10-MERS SCORE (ESTIMATE OF HALF TIME OF	SEQ.ID#
RANK	START	RESIDUE	DISASSOCIATION OF A MOLECULE	SEQ.ID#
14,1,1,1	POSITION	LISTING	CONTAINING THIS SUBSEQUENCE)	
25	173	SNILSHSYCL	6.000	925.
26	100	LLQIFAIHSL	6.000	926.
27	279	NIYLLVPPVL	5.600	927.
28	193	DIRVNVVYGL	5.600	928.
29	10	SATYFILIGL	5.600	929.
30	250	FIFYVPFIGL	4.800	930.
31	29	LAFPLCSLYL	4.800	931.
32	56	SLHEPMYIFL	4.800	932.
33	33	LCSLYLIAVL	4.800	933.
34	206	ISAIGLDSLL	4.800	934.
35	209	IGLDSLLISF	4.320	935.
36	241	TCVSHVCAVF	4.200	936.
37	215	LISFSYLLIL	4.000	937.
38	108	SLSGMESTVL	4.000	938.
39	148	VAAVVRGAAL	. 4.000	939.
40	205	IISAIGLDSL	4.000	940.
41	91	STTIQFDACL	4.000	941.
42	75	ISTSSMPKML	4.000	942.
43	109	LSGMESTVLL	4.000	943.
44	166	KQLPFCRSNI	3.600	944.
45	78	SSMPKMLAIF	3.600	945.
46	5	NGNESSATYF	3.600	946.
47	282	LLVPPVLNPI	3.024	947.
48	55	HSLHEPMYIF	3.000	948.
49	161	LPVFIKQLPF	3.000	949.
50	155	AALMAPLPVF	3.000	950.
51	247	CAVFIFYVPF	3.000	951.
52	18	GLPGLEEAQF	3.000	952.
53	299	EIRQRILRLF	2.800	953.
54	271	SPLPVILANI	2.520	954.
55	103	IFAIHSLSGM	2.500	955.
56	230	LTREAQAKAF	2.400	956.
57	113	ESTVLLAMAF	2.400	957.
58	66	CMLSGIDILI	2.100	958.
59	200	YGLIVIISAI	2.100	959.
60	256	FIGLSMVHRF	2.000	960.
61	243	VSHVCAVFIF	2.000	961.
62	87	FWFNSTTIQF	2.000	962.
63	80	MPKMLAIFWF	2.000	963.
64	156	ALMAPLPVFI	1.800	964.
65	52	RTEHSLHEPM	1.800	965.
66	197	NVVYGLIVII	1.800	966.
67	6	GNESSATYFI	1.500	967.
68	124	RYVAICHPLR	1.500	968.
69	214	LLISFSYLLI	1.500	969.
70	207	SAIGLDSLLI	1.500	970.
71	196	VNVVYGLIVI	1.500	971.
72	185	DVMKLACDDI	1.500	972.
73	39	IAVLGNLTII	1.500	973.

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TABLE XIV:

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RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
74	94	IQFDACLLQI	1.200	974.
75	298	KEIRQRILRL	1.200	975.
76	8	ESSATYFILI	1.200	976.
77	291	IVYGVKTKEI	1.100	977.
78	137	VLTLPRVTKI	1.100	978.
79	111	GMESTVLLAM	1.050	979.
80	85	AIFWFNSTTI	1.000	980.
81	41	VLGNLTIIYI	1.000	981.
82	97	DACLLQIFAI	1.000	982.
83	119	AMAFDRYVAI	1.000	983.
84	77	TSSMPKMLAI	1.000	984.
85	38	LIAVLGNLTI	1.000	985.
86	248	AVFIFYVPFI	1.000	986.
87	64	FLCMLSGIDI	1.000	987.
88	. 62	YIFLCMLSGI	1.000	988.
89	242	CVSHVCAVFI	1.000	989.
90	268	RRDSPLPVIL	0.960	990.
91	61	MYIFLCMLSG	0.750	991.
92	149	AAVVRGAALM	0.750	992.
93	20	PGLEEAQFWL	0.720	993.
94	12	TYFILIGLPG	0.700	994.
95	238	AFGTCVSHVC	0.700	995.
96	217	SFSYLLILKT	0.660	996.
97	129	CHPLRHATVL	0.600	997.
98	274	PVILANIYLL	0.600	998.
99	123	DRYVAICHPL	0.560	999.
100	74	LISTSSMPKM	0.550	1000

TABLE XV:

HLA PEPTIDE SCORING RESULTS -101P3A11- B7, 9-MERS

RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
1	130	HPLRHATVL	80.000	1001.
2	168	LPFCRSNIL	80.000	1002.
3	59	EPMYIFLCM	60.000	1003.
4	299	EIRQRILRL	40.000	1004.
5	140	LPRVTKIGV	40.000	1005.
6	149	AAVVRGAAL	36.000	1006.
7	203	IVIISAIGL	20.000	1007.
8	150	AVVRGAALM	15.000	1008.
9	110	SGMESTVLL	12.000	1009.
10	11	ATYFILIGL	12.000	1010.
11	25	AQFWLAFPL	12.000	1011.
12	207	SAIGLDSLL	12.000	1012.
13	31	FPLCSLYLI	8.000	1013.
14	28	WLAFPLCSL	6.000	1014.
15	40	AVLGNLTII	6.000	1015.
16	151	VVRGAALMA	5.000	1016.

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HLA PEPTIDE SCORING RESULTS -101P3A11-B7, 9-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE)** 253 **YVPFIGLSM** 17 5.000 1017. 18 109 LSGMESTVL 4.000 1018. 19 275 VILANIYLL 4.000 1019. **ISFSYLLIL** 20 216 4.000 1020. 21 220 YLLILKTVL 4.000 1021. 22 181 **CLHQDVMKL** 4.000 1022. 23 254 **VPFIGLSMV** 4.000 1023. 24 222 LILKTVLGL 4.000 1024. 25 TIOFDACLL 4.000 93 1025. 26 76 **STSSMPKML** 4.000 1026. 27 49 YIVRTEHSL 4.000 1027. 28 153 **RGAALMAPL** 4.000 1028. 29 92 TTIQFDACL 4.000 1029. 30 14 **FILIGLPGL** 4.000 1030. 31 101 **LQIFAIHSL** 4.000 .1031. 32 213 **SLLISFSYL** 4.000 1032. 33 174 **NILSHSYCL** 4.000 1033. 34 265 **FSKRRDSPL** 4.000 1034. 35 37 4.000 YLIAVLGNL 1035. 36 284 4.000 **VPPVLNPIV** 1036. LLISFSYLL 37 214 4.000 1037. 38 34 **CSLYLIAVL** 4.000 1038. 39 66 **CMLSGIDIL** 4.000 1039. 40 206 **ISAIGLDSL** 4.000 1040. 41 8 **ESSATYFIL** 4.000 1041. 42 104 **FAIHSLSGM** 3.000 1042. 43 283 LVPPVLNPI 2.000 1043. 44 289 **NPIVYGVKT** 2.000 1044. 45 195 RVNVVYGLI 2.000 1045. 197 46 NVVYGLIVI 2.000 1046. 47 3 **DPNGNESSA** 2.000 1047. 48 198 **VVYGLIVII** 2.000 1048. 49 274 **PVILANIYL** 2.000 1049. 50 118 LAMAFDRYV 1.800 1050. 51 155 **AALMAPLPV** 1.800 1051. 52 39 1.200 **IAVLGNLTI** 1052. 21 53 **GLEEAQFWL** 1.200 1053. 54 98 **ACLLQIFAI** 1.200 1054. 55 120 **MAFDRYVAI** 1.200 1055. 56 208 **AIGLDSLLI** 1.200 1056. 57 78 **SSMPKMLAI** 1.200 1057. 58 65 **LCMLSGIDI** 1.200 1058. 59 30 1.200 1059. AFPLCSLYL 60 304 **ILRLFHVAT** 1.000 1060. 61 72 **DILISTSSM** 1.000 1061. 62 75 **ISTSSMPKM** 1.000 1062. 230 63 LTREAOAKA 1.000 1063. 64 235 **QAKAFGTCV** 0.600 1064. 251 65 **IFYVPFIGL** 0.600 1065.

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TABLE XV:

TABLE XV:	
HLA PEPTIDE SCORING RESULTS -101P3A11- B7, 9-MERS	

	CW A DE	SUBSEQUENCE	SCORE (ESTIMATE OF HALF TIME OF	SEQ.ID#
RANK	START	RESIDUE	DISASSOCIATION OF A MOLECULE	
	POSITION	LISTING	CONTAINING THIS SUBSEQUENCE)	
66	167	QLPFCRSNI	0.600	1066.
67	296	KTKEIRQRI	0.600	1067.
68	135	ATVLTLPRV	0.600	1068.
69	159	APLPVFIKQ	0.600	1069.
70	147	GVAAVVRGA	0.500	1070.
71	50	IVRTEHSLH	0.500	1071.
72	136	TVLTLPRVT	0.500	1072.
73	226	TVLGLTREA	0.500	1073.
74	142	RVTKIGVAA	0.500	1074.
75	119	AMAFDRYVA	0.450	1075.
76	127	AICHPLRHA	0.450	1076.
77	148	VAAVVRGAA	0.450	1077.
78	273	LPVILANIY	0.400	1078.
79	271	SPLPVILAN	0.400	1079.
80	215	LISFSYLLI	. 0.400	1080.
81	269	RDSPLPVIL	0.400	1081.
82	80	MPKMLAIFW	0.400	1082.
83	124	RYVAICHPL	0.400	1083.
84	9	SSATYFILI	0.400	1084.
85	42	LGNLTIIYI	0.400	1085.
86	60	PMYIFLCML	0.400	1086.
87	280	IYLLVPPVL	0.400	1087.
88	67	MLSGIDILI	0.400	1088.
89	132	LRHATVLTL	0.400	1089.
90	55	HSLHEPMYI	0.400	1090.
91	157	LMAPLPVFI	0.400	1091.
92	138	LTLPRVTKI	0.400	1092.
93	160	PLPVFIKQL	0.400	1093.
94	19	LPGLEEAQF	0.400	1094.
95	194	IRVNVVYGL	0.400	1095.
96	186	VMKLACDDI	0.400	1096.
97	201	GLIVIISAI	0.400	1097.
98	243	VSHVCAVFI	0.400	1098.
99	85	AIFWFNSTT	0.300	1099.
100	248	AVFIFYVPF	0.300	1100.

TABLE XVI:

HLA PEPTIDE SCORING RESULTS -101P3A11- B7, 10-MERS

RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
1	59	EPMYIFLCML	240.000	1101.
2	159	APLPVFIKQL	240.000	1102.
3	273	LPVILANIYL	80.000	1103.
4	193	DIRVNVVYGL	40.000	1104.
5	140	LPRVTKIGVA	20.000	1105.
6	65	LCMLSGIDIL	12.000	1106.
7	10	SATYFILIGL	12.000	1107.
8	24	EAQFWLAFPL	12.000	1108.

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HLA PEPTIDE SCORING RESULTS -101P3A11-B7, 10-MERS SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# SUBSEQUENCE **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE)** 9 29 LAFPLCSLYL 12.000 1109. 10 148 VAAVVRGAAL 12.000 1110. 149 **AAVVRGAALM** 9.000 1111. 11 12 271 SPLPVILANI 8.000 1112. 13 185 **DVMKLACDDI** 6.000 1113. 14 250 **FIFYVPFIGL** 6.000 1114. 15 248 **AVFIFYVPFI** 6.000 1115. 16 108 SLSGMESTVL 4.000 1116. 17 91 **STTIOFDACL** 4.000 1117. 92 18 TTIOFDACLL 4.000 1118. 19 206 **ISAIGLDSLL** 4.000 1119. 20 202 LIVIISAIGL 4.000 1120. 21 296 KTKEIRQRIL 4.000 1121. 22 212 **DSLLISFSYL** 4.000 1122. 23 205 **IISAIGLDSL** 4.000 1123. 24 131 **PLRHATVLTL** 4.000 1124. 25 100 LLQIFAIHSL 4.000 1125. 26 109 **LSGMESTVLL** 4.000 1126. 27 33 LCSLYLIAVL 4.000 1127. 28 221 LLILKTVLGL 4.000 1128. 29 4.000 56 **SLHEPMYIFL** 1129. 30 215 LISFSYLLIL 4.000 1130. 31 173 **SNILSHSYCL** 4.000 1131. 32 167 **QLPFCRSNIL** 4.000 1132. 33 279 4.000 NIYLLVPPVL 1133. 34 75 **ISTSSMPKML** 4.000 1134. 35 213 SLLISFSYLL 4.000 1135. 36 180 YCLHQDVMKL 4.000 1136. 37 156 ALMAPLPVFI 3.600 1137. 38 274 **PVILANIYLL** 2.000 1138. 39 301 RORILRLFHV 2.000 1139. 40 197 **NVVYGLIVII** 2.000 1140. 41 130 **HPLRHATVLT** 2.000 1141. 42 31 **FPLCSLYLIA** 2.000 1142. 43 242 CVSHVCAVFI 2.000 1143. 44 291 **IVYGVKTKEI** 1144. 2.000 45 3 **DPNGNESSAT** 2.000 1145. 46 150 AVVRGAALMA 1.500 1146. 47 118 **LAMAFDRYVA** 1.350 1147. 48 39 **IAVLGNLTII** 1.200 1148. 49 97 **DACLLQIFAI** 1.200 1149. 50 85 **AIFWFNSTTI** 1.200 1150. 51 119 **AMAFDRYVAI** 1.200 1151. 52 207 **SAIGLDSLLI** 1.200 1152. 53 195 **RVNVVYGLIV** 1.000 1153. 54 178 **HSYCLHQDVM** 1.000 1154. 55 253 **YVPFIGLSMV** 1.000 1155. 56 142 **RVTKIGVAAV** 1.000 1156. 57 283 LVPPVLNPIV 1.000 1157.

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TABLE XVI:

TABLE	XVI:			
HIAPE	PTIDE SCORIN	IG RESULTS -101P3	A11-B7 10-MFRS	
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#
58	245	HVCAVFIFYV	1.000	1158.
59	74	LISTSSMPKM	1.000	1159.
60	189	LACDDIRVNV	0.900	1160.
61	147	GVAAVVRGAA	0.750	1161.
62	27	FWLAFPLCSL	0.600	1162.
63	154	GAALMAPLPV	0.600	1163.
64	278	ANIYLLVPPV	0.600	1164.
65	166	KQLPFCRSNI	0.600	1165.
66	134	HATVLTLPRV	0.600	1166.
67	267	KRRDSPLPVI	0.600	1167.
68	237	KAFGTCVSHV	0.600	1168.
69	234	AQAKAFGTCV	0.600	1169.
70	50	IVRTEHSLHE	0.500	1170.
71	151	VVRGAALMAP	0.500	1171.
72 .	127	AICHPLRHAT	0.450	1172.
73	126	VAICHPLRHA	0.450	1173.
74	36	LYLIAVLGNL	0.400	1174.
75	7	NESSATYFIL	0.400	1175.
76	129	CHPLRHATVL	0.400	1176.
77	214	LLISFSYLLI	0.400	1177.
78	80	MPKMLAIFWF	0.400	1178.
79	66	CMLSGIDILI	0.400	1179.
80	77	TSSMPKMLAI	0.400	1180.
81	168	LPFCRSNILS	0.400	1181.
82	137	VLTLPRVTKI	0.400	1182.
83	19	LPGLEEAQFW	0.400	1183.
84	8	ESSATYFILI	0.400	1184.
85	123	DRYVAICHPL	0.400	1185.
86	298	KEIRQRILRL	0.400	1186.
87	20	PGLEEAQFWL	0.400	1187.
88	94	IQFDACLLQI	0.400	1188.
89	284	VPPVLNPIVY	0.400	1189.
90	196	VNVVYGLIVI	0.400	1190.
91	41	VLGNLTIIYI	0.400	1191.
92	264	RFSKRRDSPL	0.400	1192.
93	38	LIAVLGNLTI	0.400	1193.
94	282	LLVPPVLNPI	0.400	1194.
95	48	IYIVRTEHSL	0.400	1195.
96	13	YFILIGLPGL	0.400	1196.
97	62	YIFLCMLSGI	0.400	1197.
98	64	FLCMLSGIDI	0.400	1198.
99	161	LPVFIKQLPF	0.400	1199.
100	152	VRGAALMAPL	0.400	1200.

TILATE		SUBSEQUENCE	A11- B-3501, 9-MERS SCORE (ESTIMATE OF HALF TIME OF	SEQ.ID#
RANK	START	RESIDUE	DISASSOCIATION OF A MOLECULE	J. S. Z. L. S.
IUIII	POSITION	LISTING	CONTAINING THIS SUBSEQUENCE)	
1	273	LPVILANIY	40.000	1201.
2	59	EPMYIFLCM	40.000	1202.
3	19	LPGLEEAQF	30.000	1203.
4	80	MPKMLAIFW	30.000	1204.
5	130	HPLRHATVL	20.000	1205.
6	172	RSNILSHSY	20.000	1206.
7	168	LPFCRSNIL	20.000	1207.
8	265	FSKRRDSPL	15.000	1208.
9	140	LPRVTKIGV	12.000	1209.
10	212	DSLLISFSY	10.000	1210.
11	75	ISTSSMPKM	10.000	1211.
12	31	FPLCSLYLI	8.000	1212.
13	109	LSGMESTVL	7.500	1213.
14	29	LAFPLCSLY	6.000	1214.
15	104	FAIHSLSGM	6.000 .	1215.
16	34	CSLYLIAVL	5.000	1216.
17	8	ESSATYFIL	5.000	1217.
18	206	ISAIGLDSL	5.000	1218.
19	216	ISFSYLLIL	5.000	1219.
20	296	KTKEIRQRI	4.800	1220.
21	284	VPPVLNPIV	4.000	1221.
22	5	NGNESSATY	4.000	1222.
23	254	VPFIGLSMV	4.000	1223.
24	285	PPVLNPIVY	4.000	1224.
25	299	EIRQRILRL	3.000	1225.
26	55	HSLHEPMYI	3.000	1226.
27	149	AAVVRGAAL	3.000	1227.
28	207	SAIGLDSLL	3.000	1228.
29	120	MAFDRYVAI	2.400	1229.
30	110	SGMESTVLL	2.000	1230.
31	150	AVVRGAALM	2.000	1231.
32	56	SLHEPMYIF	2.000	1232.
33	78	SSMPKMLAI	2.000	1233.
34	117	LLAMAFDRY	2.000	1234.
35	245	HVCAVFIFY	2.000	1235.
36	243	VSHVCAVFI	2.000	1236.
37	41	VLGNLTIIY	2.000	1237.
38	271	SPLPVILAN	2.000	1238.
39	3	DPNGNESSA	2.000	1239.
40	72	DILISTSSM	2.000	1240.
41	153	RGAALMAPL	2.000	1241.
42	253	YVPFIGLSM	2.000	1242.
43	9	SSATYFILI	2.000	1243.
44	289	NPIVYGVKT	2.000	1244.
45	235	QAKAFGTCV	1.800	1245.
46_	181	CLHQDVMKL	1.500	1246.
47	93	TIQFDACLL	1.500	1247.
48	39	IAVLGNLTI	1.200	1248.
49	186	VMKLACDDI	1.200	1249.

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TABLE XVII:

HLA PEPTIDE SCORING RESULTS -101P3A11- B-3501, 9-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE)** 50 275 VILANIYLL 1250. 1.000 **CMLSGIDIL** 1.000 1251. 51 66 52 49 YIVRTEHSL 1.000 1252. 242 CVSHVCAVF 53 1.000 1253. 54 203 IVIISAIGL 1.000 1254. 55 14 **FILIGLPGL** 1.000 1255. 56 222 LILKTVLGL 1.000 1256. 57 25 **AQFWLAFPL** 1.000 1257. 58 214 LLISFSYLL 1.000 1258. 59 114 **STVLLAMAF** 1.000 1259. 60 101 LQIFAIHSL 1.000 1260. 92 TTIQFDACL 61 1.000 1261. 62 257 **IGLSMVHRF** 1.000 1262. 28 WLAFPLCSL 1.000 63 1263. 64 79 **SMPKMLAIF** 1.000 1264. 178 65 **HSYCLHODV** 1.000 1265. 66 37 YLIAVLGNL 1.000 1266. 67 174 **NILSHSYCL** 1.000 1267. 68 156 **ALMAPLPVF** 1.000 1268. 69 SLLISFSYL 1.000 213 1269. 70 220 YLLILKTVL 1.000 1270. 71 248 **AVFIFYVPF** 1.000 1271. 72 76 STSSMPKML 1.000 1272. 73 11 **ATYFILIGL** 1.000 1273. 0.900 74 189 LACDDIRVN 1274. 75 195 **RVNVVYGLI** 0.800 1275. 76 68 **LSGIDILIS** 0.750 1276. 77 118 **LAMAFDRYV** 0.600 1277. 78 143 VTKIGVAAV 0.600 1278. 79 21 **GLEEAOFWL** 1279. 0.600 80 188 KLACDDIRV 0.600 1280. 81 208 **AIGLDSLLI** 0.600 1281. 82 155 **AALMAPLPV** 0.600 1282. 230 83 **LTREAQAKA** 0.600 1283. 84 113 **ESTVLLAMA** 0.500 1284. 85 90 0.500 **NSTTIOFDA** 1285. 86 218 **FSYLLILKT** 0.500 1286. 87 77 **TSSMPKMLA** 0.500 1287. 88 107 **HSLSGMEST** 0.500 1288. 89 270 **DSPLPVILA** 0.500 1289. 90 167 **QLPFCRSNI** 0.400 1290. 91 198 VVYGLIVII 0.400 1291. 92 201 **GLIVIISAI** 0.400 1292. **NVVYGLIVI** 93 197 0.400 1293. 94 98 1294. **ACLLQIFAI** 0.400 95 67 **MLSGIDILI** 0.400 1295. 96 138 LTLPRVTKI 0.400 1296. 97 157 LMAPLPVFI 0.400 1297. 98 283 LVPPVLNPI 0.400 1298.

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TABLE XVII:

TABLE XVII:						
HLA PE	HLA PEPTIDE SCORING RESULTS -101P3A11- B-3501, 9-MERS					
RANK	START POSITION	SUBSEQUENCE RESIDUE LISTING	SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)	SEQ.ID#		
99	40	AVLGNLTII	0.400	1299.		
100	215	LISFSYLLI	0.400	1300.		

TABLE XVIII: HLA PEPTIDE SCORING RESULTS -101P3A11-B-3501, 10-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEQ.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING **CONTAINING THIS SUBSEQUENCE)** 80 1 **MPKMLAIFWF** 60.000 1301. 2 284 **VPPVLNPIVY** 40.000 1302. 3 273 **LPVILANIYL** 20.000 1303. 4 159 **APLPVFIKQL** 20.000 1304. 5 59 **EPMYIFLCML** 20.000 1305. 6 161 **LPVFIKOLPF** 20.000 1306. • 7 19 15.000 **LPGLEEAQFW** 1307. 8 296 KTKEIRORIL 12.000 1308. 9 178 **HSYCLHQDVM** 10.000 1309. 10 271 8.000 **SPLPVILANI** 1310. 11 140 **LPRVTKIGVA** 6.000 1311. 230 12 **LTREAQAKAF** 6.000 1312. 13 149 AAVVRGAALM 6.000 1313. 5.000 14 206 **ISAIGLDSLL** 1314. 15 75 ISTSSMPKML 5.000 1315. 16 243 **VSHVCAVFIF** 5.000 1316. 212 17 **DSLLISFSYL** 5.000 1317. 18 113 **ESTVLLAMAF** 5.000 1318. 19 78 **SSMPKMLAIF** 5.000 1319. 20 55 **HSLHEPMYIF** 5.000 1320. 21 109 **LSGMESTVLL** 5.000 1321. 22 10 SATYFILIGL 3.000 1322. 23 193 **DIRVNVVYGL** 3.000 1323. 24 155 **AALMAPLPVF** 3.000 1324. 299 25 **EIRQRILRLF** 3.000 1325. 26 3 **DPNGNESSAT** 3.000 1326. 27 148 VAAVVRGAAL 3.000 1327. 28 LAFPLCSLYL 29 3.000 1328. 29 24 **EAQFWLAFPL** 3.000 1329. 30 247 **CAVFIFYVPF** 3.000 1330. SLHEPMYIFL 31 56 2.000 1331. 74 2.000 32 LISTSSMPKM 1332. 33 130 **HPLRHATVLT** 2.000 1333. 34 77 **TSSMPKMLAI** 2.000 1334. 35 5 **NGNESSATYF** 2.000 1335. 36 116 VLLAMAFDRY 2.000 1336. 37 28 WLAFPLCSLY 2.000 1337. 38 31 **FPLCSLYLIA** 2.000 1338. 39 168 **LPFCRSNILS** 2.000 1339.

2.000

2.000

1340.

1341.

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ESSATYFILI

AVLGNLTIIY

40

41

8

40

HLA PEPTIDE SCORING RESULTS -101P3A11- B-3501, 10-MERS SUBSEQUENCE SCORE (ESTIMATE OF HALF TIME OF SEO.ID# **START RANK** RESIDUE DISASSOCIATION OF A MOLECULE **POSITION** LISTING CONTAINING THIS SUBSEQUENCE) 42 209 **IGLDSLLISF** 1342. 2.000 43 207 SAIGLDSLLI 1.800 1343. 44 18 **GLPGLEEAOF** 1.500 1344. 45 108 **SLSGMESTVL** 1.500 1345. 46 180 YCLHQDVMKL 1.500 1346. 47 92 TTIQFDACLL 1.500 1347. 48 237 KAFGTCVSHV 1.200 1348. 49 97 1.200 **DACLLQIFAI** 1349. 50 39 1.200 **IAVLGNLTII** 1350. 51 301 **RORILRLFHV** 1.200 1351. 52 52 **RTEHSLHEPM** 1.200 1352. 53 189 **LACDDIRVNV** 1.200 1353. 54 218 **FSYLLILKTV** 1.000 1354. 55 241 **TCVSHVCAVF** 1.000 1355. 56 215 1.000 LISFSYLLIL 1356. 57 33 **LCSLYLIAVL** 1.000 1357. 58 202 LIVIISAIGL 1.000 1358. 59 221 LLILKTVLGL 1.000 1359. 60 172 **RSNILSHSYC** 1.000 1360. 100 61 LLQIFAIHSL 1.000 1361. 62 173 **SNILSHSYCL** 1.000 1362. 63 279 NIYLLVPPVL 1.000 1363. 64 107 **HSLSGMESTV** 1.000 1364. 65 65 LCMLSGIDIL 1.000 1365. 66 205 **IISAIGLDSL** 1.000 1366. 67 91 **STTIOFDACL** 1.000 1367. 68 250 **FIFYVPFIGL** 1.000 1368. 69 213 **SLLISFSYLL** 1.000 1369. 70 256 **FIGLSMVHRF** 1.000 1370. 71 167 1371. **OLPFCRSNIL** 1.000 72 235 0.900 **QAKAFGTCVS** 1372. 73 94 **IQFDACLLQI** 0.800 1373. 74 166 **KQLPFCRSNI** 0.800 1374. 75 134 **HATVLTLPRV** 0.600 1375. 76 111 **GMESTVLLAM** 0.600 1376. 77 154 **GAALMAPLPV** 0.600 1377. 78 143 VTKIGVAAVV 0.600 1378. 79 120 **MAFDRYVAIC** 0.600 1379. 80 68 **LSGIDILIST** 0.500 1380. 81 90 **NSTTIQFDAC** 0.500 1381. 82 79 **SMPKMLAIFW** 0.500 1382. 83 270 **DSPLPVILAN** 0.500 1383. 84 267 KRRDSPLPVI 0.480 1384. 85 118 LAMAFDRYVA 0.450 1385. 291 86 **IVYGVKTKEI** 0.400 1386. 87 282 LLVPPVLNPI 0.400 1387. 88 242 **CVSHVCAVFI** 0.400 1388. 89 64 **FLCMLSGIDI** 0.400 1389. 90 85 **AIFWFNSTTI** 0.400 1390.

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TABLE XVIII:

TABLE	TABLE XVIII:				
HLA PE	PTIDE SCORIN	G RESULTS -101P3	A11- B-3501, 10-MERS		
RANK START POSITION SUBSEQUENCE RESIDUE LISTING SCORE (ESTIMATE OF HALF TIME OF DISASSOCIATION OF A MOLECULE CONTAINING THIS SUBSEQUENCE)		SEQ.ID#			
91	142	RVTKIGVAAV	0.400	1391.	
92	137	VLTLPRVTKI	0.400	1392.	
93	38	LIAVLGNLTI	0.400	1393.	
94	62	YIFLCMLSGI	0.400	1394.	
95	214	LLISFSYLLI	0.400	1395.	
96	156	ALMAPLPVFI	0.400	1396.	
97	41	VLGNLTIIYI	0.400	1397.	
98	196	VNVVYGLIVI	0.400	1398.	
99	248	AVFIFYVPFI	0.400	1399.	
100	200	YGLIVIISAI	0.400	1400.	

Table XIX: Motifs and Post-translational Modifications of 101P3A11

5 N-glycosylation site

Number of matches: 3

1 7-10 NESS (SEQ ID NO: 2881) 2 44-47 NLTI -(SEQ ID NO: 2882) 3 90-93 NSTT (SEQ ID NO: 2883)

3 90-93 NSTT

cAMP- and cGMP-dependent protein kinase phosphorylation site 268-271 RRDS (SEQ ID NO: 2884)

200-271 Idd55

Protein kinase C phosphorylation site

15 266-268 SKR (SEQ ID NO: 2885)

Casein kinase II phosphorylation site

Number of matches: 3

20

25

1 56-59 SLHE (SEQ ID NO: 2886) 2 69-72 SGID (SEQ ID NO: 2887) 3 110-113 SGME (SEQ ID NO: 2888)

N-myristoylation site

Number of matches: 4

1 6-11 GNESSA (SEQ ID NO: 2889) 2 21-26 GLEEAQ (SEQ ID NO: 2890) 3 111-116 GMESTV (SEQ ID NO: 2891)

4 240-245 GTCVSH

30 G-protein coupled receptors family 1 signature

112-128 MESTVLLAMAFDRYVAI (SEQ ID NO: 2892)

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Table XX	K: Frequen	tly Occurring Motifs	
Name	avrg. % identity	Description	Potential Function
			Nucleic acid-binding protein functions as
			transcription factor, nuclear location
zf-C2H2	34%	Zinc finger, C2H2 type	probable
		Cytochrome b(N-	membrane bound oxidase, generate
cytochrome b N	68%	terminal)/b6/petB	superoxide
			domains are one hundred amino acids long
			and include a conserved intradomain
ig	19%	Immunoglobulin domain	disulfide bond.
			tandem repeats of about 40 residues, each
			containing a Trp-Asp motif. Function in
<u>WD40</u>	18%	WD domain, G-beta repeat	signal transduction and protein interaction
			may function in targeting signaling
PDZ	23%	PDZ domain	molecules to sub-membranous sites
			short sequence motifs involved in protein-
LRR	28%	Leucine Rich Repeat	protein interactions
,,			conserved catalytic core common to both
			serine/threonine and tyrosine protein
			kinases containing an ATP binding site and
<u>pkinase</u>	23%	Protein kinase domain	a catalytic site
			pleckstrin homology involved in
			intracellular signaling or as constituents of
<u>PH</u>	16%	PH domain	the cytoskeleton
			30-40 amino-acid long found in the
			extracellular domain of membrane-bound
<u>EGF</u>	34%	EGF-like domain	proteins or in secreted proteins
		Reverse transcriptase	
		(RNA-dependent DNA	
<u>rvt</u>	49%	polymerase)	
			Cytoplasmic protein, associates integral
<u>ank</u>	25%	Ank repeat	membrane proteins to the cytoskeleton
	1	NADH-	
		Ubiquinone/plastoquinone	membrane associated. Involved in proton
oxidored q1	32%	(complex I), various chains	translocation across the membrane

Table	Table XX: Frequently Occurring Motifs				
Name	avrg. % identity	Description	Potential Function		
			calcium-binding domain, consists of a12		
			residue loop flanked on both sides by a 12		
efhand	24%	EF hand	residue alpha-helical domain		
			Aspartyl or acid proteases, centered on a		
<u>rv</u> p	79%	Retroviral aspartyl protease	catalytic aspartyl residue		
			extracellular structural proteins involved in		
			formation of connective tissue. The		
		Collagen triple helix repeat,	sequence consists of the G-X-Y and the		
Collagen	42%	(20 copies)	polypeptide chains forms a triple helix.		
			Located in the extracellular ligand-binding		
			region of receptors and is about 200 amino		
			acid residues long with two pairs of		
<u>fn3</u>	20%	Fibronectin type III domain	cysteines involved in disulfide bonds		
			seven hydrophobic transmembrane regions,		
			with the N-terminus located extracellularly		
		7 transmembrane receptor	while the C-terminus is cytoplasmic.		
7tm_1	19%	(rhodopsin family)	Signal through G proteins		

Table XXI: Nucleotide sequence of the splice variant. (SEQ ID NO: 1401)

```
1 CACATTCCTT CCATACGGTT GAGCCTCTAC CTGCCTGGTG CTGGTCACAG TTCAGCTTCT
5
          61 TCATGATGGT GGATCCCAAT GGCAATGAAT CCAGTGCTAC ATACTTCATC CTAATAGGCC
         121 TCCCTGGTTT AGAAGAGGCT CAGTTCTGGT TGGCCTCCCA TTGTGCTCCC TCTANCTATG
         181 CTGTGCTAGT AATTGACAAT CATCTACATG TGCGGACGAG CACGNCGCNG AGCCCNGTAT
         241 NATTCTGCNG CTTCAGCATG ACACCCTNCA GTCTCAGCCA AAGNGCATCT CNGTCAATCA
         301 NACACNTGAG CTGTCGTACG AGTTGCATCA TCCTANGGCA GGATCAATGT GCGGNAGGCN
10
         361 TGACGCAGTG CACGTACCAT GGCAGCAAGA CAGGGCCGGT ACAAATGGGG GCGAGNCGGG
         421 GTGAAGATGN ACCCTCGGGT CANAGAGTGC CTCTGCGCCA AAACCTCCAT CATGNNAACA
         481 GNGTATAACG GCGNAGAATC GGNNANGCGC AAGGCTAAGG AAANNCCCAA NNCNGGTACT
         541 TTAACCCNGC AAANGGCANC NAAACGGGNG GGTNANTGAA CAAGGAAGGN NTGNAACTGG
         601 GCCAAAACGG GNTGGGCAAN NNAAGGACTC ATGGGNCCAA GGGACGGNAA AAGGGGNAAN
15
         661 CGGGGCGAAA TGNNAAAAAC CGGGNCCCGG GGAANAANGA AGGGGAANAN GNGTGAAGGA
         721 CNGGGTTCAA GGGAAAAGNA AAACCANGGG NNAGAAACCN TTCNAANGGC CCGGGNANGA
         781 AAGGAANTNN GNNNGGNGAA AAAATCNAAA AAAAGCNGNG GCNNAAAAAN GGGGGGAANN
         841 NAAANACCNN GGNCGNNAAA AAACNNAANG NGGGGGGANT ANACACGGAA ANNNANGGGC
         901 GNNNAAGGGA AATAANNCGG GAACNAAAGN GCAAACCGNA CGGNAGGAAC GAAACCCACC
20
         961 GGAGNCGCNN AACGCCNNNC NNANCCCGAG CNGAGGTNG
    Table XXII: Nucleotide sequence alignment of 101P3A11 (SEQ ID NO: 1402) with the
    splice variant (SEQ ID NO: 1403).
25
    Score = 337 bits (175), Expect = 4e-89
    Identities = 215/223 (96%), Gaps = 6/223 (2%)
    Strand = Plus / Plus
30
    Variant: 1
                cacattccttccatacggttgagcctctacctgcctggtgctggtcacagttcagcttct 60
35
    101P3A11: 128 tcatgatggtggatcccaatggcaatgaatccagtgctacatacttcatcctaataggcc 187
                Variant : 61 tcatgatggtggatcccaatggcaatgaatccagtgctacatacttcatcctaataggcc 120
40
    101P3A11: 188 tccctggtttagaagaggctcagttctggttggccttcccattgtgctccctctacctta 247
                Variant: 121 tccctggtttagaagaggctcagttctggttggcc-tcccattqtgctccctctanct-- 177
45
    101P3A11: 248 ttgctgtgctaggtaacttgacaatcatctacattgtgcggac 290
                Variant: 178 atgctgtgcta-gtaa-ttgacaatcatctaca-tgtgcggac 217
50
    Table XXIII: Longest single amino acid sequence alignment of 101P3A11 (SEQ ID
    NO: 1404) and the splice variant (SEQ ID NO: 1405).
    Score = 134 \text{ bits } (287), \text{ Expect } (2) = 3e-29
    Identities = 51/51 (100%)
     Frame = +1 / +3
55
    101P3A11: 70 HSFHTVEPLPAWCWSQFSFFMMVDPNGNESSATYFILIGLPGLEEAQFWLA 222
                 HSFHTVEPLPAWCWSQFSFFMMVDPNGNESSATYFILIGLPGLEEAQFWLA
    Variant : 3
                 HSFHTVEPLPAWCWSQFSFFMMVDPNGNESSATYFILIGLPGLEEAOFWLA 155
```

Table XXIV: Peptide sequences from the translation of the nucleotide sequence of the splice variant .

Open reading	Amino acid sequences
frame	
Frame 1	HIPSIRLSLYLPGAGHSSASS*WWIPMAMNPVLHTSS**ASLV*KRLSSGWPPIVLPLXM
<u>SEQ ID NO: 1406</u>	LC**LTIIYMCGRARRXAXYXSAASA*HPXVSAKXHLXQSXT*AVVRVASSXGRINVRXA
	*RSARTMAARQGRYKWGRXGVKMXPRVXECLCAKTSIMXTXYNGXESXXRKAKEXPXXGT
	LTXQXAXKRXGX*TRKXXNWAKTGWAXXGLMGPRDGKRGXRGEMXKTGXRGXXKGXXXEG
	XGSREKXNXGXETXXXARXXKEXXXXKKSKKSXGXKXGGXXXPXXXKNXXXGXXTRKXXG
	XXGK*XGNXXANRTXGTKPTGXAXRXXXPEXRX
Frame 2	TFLPYG*ASTCLVLVTVQLLHDGGSQWQ*IQCYILHPNRPPWFRRGSVLVGLPLCSLXLC
<u>SEQ ID NO: 1407</u>	CASN*QSSTCADEHXAEPXXILXLQHDTLQSQPKXISVNXTXELSYELHHPXAGSMCGRX
	DAVHVPWQQDRAGTNGGEXG*RXTLGSXSASAPKPPSXXQXITAXNRXXARLRKXPXXVL
	*PXKXXXNGXVXEQGRXXTGPKRXGQXKDSWXQGTXKGXXGAKXXKPGPGEXXRGXXVKD
	XVQGKXKTXGXKPFXXPGXERXXXXXKNXKKAXAXKXGEXKXXGRXKTXXGGXXHGXXXA
	XKGNXXGTKXQTXRXERNPPEXXNAXXXPSXG
Frame 3	HSFHTVEPLPAWCWSQFSFFMMVDPNGNESSATYFILIGLPGLEEAQFWLASHCAPSXYA
<u>SEQ ID NO: 1408</u>	VLVIDNHLHVRTSTXXSPVXFCXFSMTPXSLSQXASXSIXHXSCRTSCIILXQDQCAXGX
	TQCTYHGSKTGPVQMGAXRGEDXPSGXRVPLRQNLHHXNXV*RRRIGXAQG*GXXQXXYF
	NPAXGXXTGGXXNKEGXXLGQNGXGXXRTHGXKGRXKGXXGRNXKNRXPGXXEGEXX*RT
•	GFKGKXKPXXRNXSXGPGXKGXXXGEKIXKKXXXKXGGXXXTXXXKKXXXGGXXTEXXGR
	XREIXREXKXKPXGRNETHRXRXTPXXXRAEV

Note: Frame 3 gives the longest subsequence that is identical with 101P3All amino acid sequence. In this Table each (*)indicates the product of a single stop codon, and 'X' indicates a single unknown amino acid.

5

TABLE XXV: Properties of 101P3A11

Program DRF Finder /a M Pred	URL http://www.ncbi.nlm.gov/gorf	Outcome 133-1086
/a	http://www.ncbi.nlm.gov/gorf	
		(includes stop)
M Pred	n/a	317 amino acids
WIIICU	http://www.ch.embnet.org/	7 TM at aa :
		27-52, 63-88, 104-129,
		146-165, 196-224,
		239-262, 273-295
łММТор	http://www.enzim.hu/hmmtop/	7 TM at aa:
		27-50, 63-86, 99-121,
		146-165, 201-224,
		239-262, 275-294
osui	http://www.genome.ad.jp/SOSui/	7 TM, at aa:
		29-51, 63-85, 100-122,
		203-225, 239-261,
	handless of the distance of th	273-295
MHMM	, -	7 TM, at aa:
	IVI	29-51, 63-85, 100-122,
		143-165, 202-224, 236-258, 273-295
Signal P	http://www.chs.dtu.dk/services/SignalD	
-		pl 8.7
		35.2 kDa
		Plasma membrane
SORT	intp://psort.moo.ac.jp/	64%
SOPT II	http://psort.nibb.ac.in/	Plasma membrane
SORT II	http://psort.mob.ac.jp/	56.4%
)fam	http://www.sanger.ac.uk/Pfam/	7 transmembrane
Talli	intp://www.sanger.ac.uk/r fain/	receptor (rhodopsin
		family)
rints	http://www.biochem.ucl.ac.uk/	Rhodopsin-like GPCR
11110	intep.,, www.sioonom.uo.uo.uo	superfamily
Blocks	http://www.blocks.fhcrc.org/	Rhodopsin-like GPCR
		superfamily
		•
rosite	http://www.genome.ad.ip/	G-protein coupled
	, 3 · · · · · · · · · · · · · · · · · ·	receptors family 1
	osui MHMM ignal P I/MW tool I/MW tool SORT SORT II fam rints clocks	http://www.genome.ad.jp/SOSui/ http://www.cbs.dtu.dk/services/TMHM ignal P I/MW tool I/MW tool SORT http://www.expasy.ch/tools/ http://psort.nibb.ac.jp/ http://psort.nibb.ac.jp/ http://www.sanger.ac.uk/Pfam/ http://www.biochem.ucl.ac.uk/ http://www.blocks.fhcrc.org/

<u>sd-63712</u> ______159

HLA-A*0201 nonomers

Table XXVI: HLA Class I Nonamers	Pos	1 2 3 4 5 6 7 8 9	score
HIM CIASS I NONAMEIS	11 275	VILANIYLL	26
SEQ	12 157	<u> </u>	25
<u>ID</u>	13 220	- Y L L I L K T V L	25
NO: HLA-A1 nonomers	14 276	<u> </u>	25
Pos 1 2 3 4 5 6 7 8 9 score 1409 1 245 HVCAVFTFY 24	15 279	- N - I - Y - L - L - V - P - P - V	25
	16 138	<u>-L T L P R V</u> T K I	24
	17 213	<u>-S L L I S F S Y L</u>	24
1411 3 41 V L G N L T I I Y 21 1412 4 285 P P V L N P I V Y 20	18 49	YIVRTEHSL	23
1413 5 111 GMESTVLLA 19	19 143	<u> </u>	23
1414 6 117 L L A M A F D R Y 19	20 188	<u> </u>	23
1415 7 172 R S N I L S H S Y 19	21 198	-V V Y G L I V I I	23
1416 8 192 D D I R V N V V Y 19	22 21	-G-L E E A Q F W L	22
1417 9 212 D S L L I S F S Y 19	23 40	-AVLCNLTII	22
1418 10 57 L H E P M Y I F L 18	24 206	-ISAIG <u>L</u> DSL	22
1419 11 22 LEEAQFWLA 17	25 11	-A-T Y F I <u>L</u> I-C-L	21
1420 12 9 S S A T Y F I L I 16	26 60	-P M Y I F <u>L</u> C M L	21
1421 13 52 RTEHSLHEP 16	27 135	- A T V L T <u>L P R V</u>	21
1422 14 54 E H S L H E P M Y 16	28 160	<u> </u>	21
1423 15 78 S S M P K M L A I 16	29 174	<u> N I L S H <u>S</u> Y C L</u>	21
- -	30 207	-S-A I G L <u>D</u> S-L L	21
	31 272	<u>-P-L P V I <u>L</u> A N I</u>	21
	32 283	LVPPVLNPI	21
<u> </u>	33 67	M L S-G I D I L I	20
	34 101	LQIFAIHSL	20
	35 282	LLVPPVLNP	20
	36 299	-EIRQRILRL	20
1430 22 273 L P V I L A N I Y 15	37 304	<u>- I L R L F H V A T</u>	· 20
1431 23 271 S P L P V I L A N 14	38 39	<u></u>	19
1432 24 91 S T T I Q F D A C 13	39 45		19
1433 25 121 A F D R Y V A I C 13	40 92	TTIQFDACL	19
1434 26 138 L T L P R V T K I 13 1435 27 218 F S Y L L I L K T 13	41 110	-SCMESTVLL	19
-	42 127	- A I C H P L R H A	19
- -	43 132	_	19
1437 29 190 A C D D I R V N V 12 1438 30 191 C D D I R V N V V 12	44 149	- A A V V R G A A L	19
	45 155	- A A L M A P L P V	19
	46 156	-A L M A P L P V F	19
	47 203	<u>-I V I I S A I G L</u>	19
<u>1441</u> 33 270 D <u>S</u> P L P V <u>I</u> L A 12	48 208	AIGLDSLLI	19
HLA-A*0201 nonomoro	49 216	<u>- I S F S Y L L I L</u>	19
HIM IN OLOT HOHOMOLO	50 219	- S Y L L I L K T V	19
Pos score	51 221	<u>-L </u>	19
	52 223	<u> </u>	19
	53 17	-I G L P G L E E A	18
· 	54 33	-L C S L Y L I A V	18
-	55 34	CSLYLIAVL	18
4 37 Y L I A V L C N L 28	56 38	LIAVLCNLT	18
5 222 LILKTVLGL 28	57 43	_	18
6 66 CMLSCIDIL 26	58 85	_	18
7 108 S L S C M E S T V 26	59 118	_	18
8 181 C I H Q D V M K I 26		IRVNVVYGL	18
9 201 G L I V I I S A I 26	61 210	-	18
10 214 LLISFSYLL 26		LISFSYLLI	18
od 62712			10
sd-63712	160		

5

HLA-A*0201	nonomers
Pos	

HLA A*0203 nonomers

•	Pos		score	-	Pes		score
		1 2 3 4 5 6 7 8 9				1 2 3 4 5 6 7 8 9	00020
63	246	V C A V F I F Y V	18	4	97	-DACLLQIFA	11
64	254	VPFI-GLSMV	18	5	127	A I C H P L R H A	10
65	15	<u> </u>	17	6	3	-D-P-N-C-N-E-S-S-A	9
66	63	IFLCMLSGI	17	7	17	<u>-I G L P C L E E A</u>	9
67	72	DILISTSSM	17	8	22	-L <u>E E A Q F W L A</u>	9
68	93	TIQFDACLL	17	9	32	P L C S L Y L I A	, 9
69	98	ACLUQIFAI	17	10	77	<u>T </u>	9
70	111	GMESTVLLA	17	11	90	NSTTIQEDA	9
71	120	MAFDRYVAI	17	12	111	CMESTVLLA	9
72	167	QLPFCRSNI	1.7	13	113	ESTVLLAMA	9
73	197	NVVYGLIVI	17	14	141	P R V T K I G V A	9
74	226	TVLGLTREA	17	15	142	RYTKICYNA	9
75	281	Y L L V P P V L N	17	16	151	V V R C A A L M A	9
76	31	FPLCSLYLI	16	17	182	THODVMKLA	9
77	56	SLHEPMYIF	16	18	200	Y C L I V I I S A	9
78	70	GIDILISTS	16	19	226	TVLCLTREA	9
79	78	SSMPKMLA-I	16	20	228	LCLTREAQA	9
80	79	SMPKMLAIF	16	21	230	LTREAQAKA	9
81 82	104	FAIHSLSG-M	16	22	240	CTCVSHVCA	9
82	119	-	16	23	270	D S P L P V I L A	9
83	144 147	<u> </u>	16 16	24 25	303	RILRIFHVA	9 9
85	186	<u>V M K L A C D D I</u>	16	23	306	R L F H V A T H A	9
86	230	- L T R E A Q A K A	16		HLA-A2	5 nonomers	
87	238	AFCTCVSHV	16	=		1011011011011	
88	249	VFIFYVPFI	16		Pos	123456789	score
89	302	0 R I L R L F H V	16	1	299	EIRORI LRL	30
90	303	RILRLFHVA	16	2	72	DILISTS SM	27
91	18	G L P G L E E A Q	15	3	248	AVFIFYVPF	27
92	35	SLYLIAVLG	15 15	4	210	-C-L-D-S-L-L-I-S-F	26
93	42	LCNL-TIIXI	15	5	14	FILIC PGL	24
94	4-6	-T I I Y I V R T E	15	6	56	S L H E P M Y I F	24
95	69	-S-G-I-D-I-I-S-T	15	7	117	LLAMAFDRY	24
96	76	STSSMPKML	15	8	222	LILKTVLGL	24
97	131		15	9	245	HVCAVFIFY	24
98	137	-V L T L P R V T K	15	10	11	-A-T-Y F-I-L-I-G-L	23
99	153	-R C A A L M A P L	15	11	· 37	Y L I A V L G N L	23
100	190	ACDDIRVNV	15	12	114	STVLLAMAF	23
101	191	-C D I R V N V V	15	13	156	ALMAPLPVF	23
102		VIISAIGLD	15	14	162	PVFIKQLPF	23
	204		10				
103		T C V S H V C A V	15	15	181	-C-L-H-Q-D-V-M-K-L	23
103 104	241	-			181	-C-L-H-Q-D-V-M-K-L -W-L-A-F-P-L-C-S-L	23 22
	241 251	$\overline{\mathbf{T}} \mathbf{C} \mathbf{V} \mathbf{S} \mathbf{H} \overline{\underline{\mathbf{V}}} \mathbf{C} \mathbf{A} \mathbf{V}$	15	15	181 28		
104	241 251 269	T C V S H V C A V	15 15	15 16	181 28 92	WLAFPLCSL	22
104 105	241 251 269 280	T C V S H V C A V I F Y V P F I G L R D S P L P V I L	15 15 15	15 16 17	181 28 92 160 203	W L A F P L C S L T T I Q F D A C L	22 22 22 22
104 105 106 107	241 251 269 280 306	T C V S H V C A V I F Y V P F I G L R D S P L P V I L I Y L L V P P V L R L F H V A T H A	15 15 15 15	15 16 17 18 19 20	181 28 92 160 203 213	W L A F P L C S L T T I Q F D A C L P L P V F I K Q L	22 22 22 22 22
104 105 106 107	241 251 269 280 306	T C V S H V C A V I F Y V P F I G L R D S P L P V I L I Y L L V P P V L	15 15 15 15	15 16 17 18 19 20 21	181 28 92 160 203 213 275	W L A F P L C S L T T I Q F D A C L P L P V F I K Q L I V I I S A I G L S L L I S F S Y L V I L A N I Y L L	22 22 22 22 22 22
104 105 106 107	241 251 269 280 306	T C V S H V C A V I F Y V P F I G L R D S P L P V I L I Y L L V P P V L R L F H V A T H A	15 15 15 15	15 16 17 18 19 20 21 22	181 28 92 160 203 213 275 193	W L A F P L C S L T T I Q F D A C L P L P V F I K Q L I V I I S A I G L S L L I S F S Y L V I L A N I Y L L D I R V N V V Y G	22 22 22 22 22 22 22 21
104 105 106 107	241 251 269 280 306 HLA A*(T C V S H V C A V I F Y V P F I G L R D S P L P V I L I Y L L V P P V L R L F H V A T H A 0203 nonomers	15 15 15 15 15	15 16 17 18 19 20 21 22 23	181 28 92 160 203 213 275 193 242	W L A F P L C S L T T I Q F D A C L P L P V F I K Q L I V I I S A I G L S L L I S F S Y L V I L A N I Y L L D I R V N V V Y G C V S H V C A V F	22 22 22 22 22 22 21 21
104 105 106 107	241 251 269 280 306 HLA A*(T C V S H V C A V I F Y V P F I G L R D S P L P V I L I Y L L V P P V L R L F H V A T H A 0203 nonomers 1 2 3 4 5 6 7 8 9 V A A V V R G A A	15 15 15 15 15 15	15 16 17 18 19 20 21 22 23 24	181 28 92 160 203 213 275 193 242 76	W L A F P L C S L T T I Q F D A C L P L P V F I K Q L I V I I S A I G L S L L I S F S Y L V I L A N I Y L L D I R V N V V Y G C V S H V C A V F S T S S M P K M L	22 22 22 22 22 22 21 21 21
104 105 106 107	241 251 269 280 306 306 HLA A*(Pos 148 119	T C V S H V C A V I F Y V P F I G L R D S P L P V I L I Y L L V P P V L R L F H V A T H A 0203 nonomero 1 2 3 4 5 6 7 8 9 V A A V V R G A A A M A F D R Y V A	15 15 15 15 15 15 15	15 16 17 18 19 20 21 22 23 24 25	181 28 92 160 203 213 275 193 242 76 253	W L A F P L C S L T T I Q F D A C L P L P V F I K Q L I V I I S A I G L S L L I S F S Y L V I L A N I Y L L D I R V N V V Y C C V S H V C A V F S T S S M P K M L Y V P F I G L S M	22 22 22 22 22 22 21 21 20 20
104 105 106 107	241 251 269 280 306 HLA A*(T C V S H V C A V I F Y V P F I G L R D S P L P V I L I Y L L V P P V L R L F H V A T H A 0203 nonomers 1 2 3 4 5 6 7 8 9 V A A V V R G A A	15 15 15 15 15 15	15 16 17 18 19 20 21 22 23 24	181 28 92 160 203 213 275 193 242 76 253	W L A F P L C S L T T I Q F D A C L P L P V F I K Q L I V I I S A I G L S L L I S F S Y L V I L A N I Y L L D I R V N V V Y G C V S H V C A V F S T S S M P K M L	22 22 22 22 22 22 21 21 21
104 105 106 107	241 251 269 280 306 HLA A*(Pos 148 119 147	T C V S H V C A V I F Y V P F I G L R D S P L P V I L I Y L L V P P V L R L F H V A T H A 0203 nonomero 1 2 3 4 5 6 7 8 9 V A A V V R G A A A M A F D R Y V A	15 15 15 15 15 15 15	15 16 17 18 19 20 21 22 23 24 25	181 28 92 160 203 213 275 193 242 76 253	W L A F P L C S L T T I Q F D A C L P L P V F I K Q L I V I I S A I G L S L L I S F S Y L V I L A N I Y L L D I R V N V V Y C C V S H V C A V F S T S S M P K M L Y V P F I G L S M	22 22 22 22 22 22 21 21 20 20

H	LA-A2	nonomers		Ħ	LA-A2	nonomers	
	Pes	1 2 3 4 5 6 7 8 9	score		Pos	123456789	score
27	23	EEAOFWLAF	19	79	287	V L N P I V Y G V	15
28	41	V L C N L T I I Y	19	80	296	KTKEIRORI	15
29	49	Y I V R T E H S L	19	81	303	RILRLFHVA	15
30	150	AVVRGAALM	19				
31	174	-NI-LSHSYC-L	19	H	LA-A3	-nonomers	
32	192	DDIRVNVY	19		Pos		score
33	214	LLISFSYLL	19			123456789	
34	251	IFYVPFICL	19	1	137	<u> </u>	30
35	8	ESSATYFIL	18	£	229	GLTREAQAK	27
36	21	G L E E A Q F W L	18	3	145	KIGVAAVVR	26
37	45	LTIIYIVRT	18	4	150	-A-V <u>V R G A A L M</u>	24
38	54	E H S L H E P M Y	18	5	290	-P-I V-Y G V K T-K	24
39	59	EPMYIFLCM	18	6	35	SLYLIAVLG	23
40	88	WFNSTTIQF	18	7	156	ALMAPLPVF	23
41	93	TIQFDACLL	18	8	47 50	<u>-I I Y I V R T E H</u> -I V R T E H S L H	22 22
42	185	DVMKLACDD	18	10	30 142	RVTKIGVAA	22
43	198	V V Y G L I V I I	18 17	11	151	<u> </u>	22
44 45	62 70	YIFLCMLSG	17	12	242	- C V S H V C A V F	22
46	70 79	S M P K M L A I F	17	13	248	<u> </u>	22
47	96	FDACLLOIF	17	14	116	V L L A M A F D R	21
48	104	FAIHSLS GM	17	15	192	-D D I R V N V Y	21
49	138	LTLPRVTKI	17	16	303	RILRLFHVA	21
50	143	VTKICVAAV	17	17	304		21
51	204	VIISAIC L D	17	18	108	<u> </u>	20
52	212	DSLLISFSY	17	19	198	V V Y C L I V I I	20
53	220	Y L L I L K T V L	17	20	291	IVYCVKTKE	20
54	256	FIGLSMVHR	17	21	15	I L I C L P C L E	19
55	283	LVPPVLNPI	17	22	44	<u>N L T I I Y I V R</u>	19
56	29	LAFPLCSLY	16	23	73	-I-L <u>I</u> S T <u>S S</u> M P	19
57	40	AVLCNLTII	16	24	74	-L I <u>S</u> T S <u>S</u> <u>M</u> P K	19
58	46	TIIYIVRTE	16	25	99	CLLQIFAIH	19
59	52	RTEHSLHEP	16	26	162	P V F I K Q L P F	19
60	75	ISTSSMPKM	16	27	203	IVIISAICL	19
61		STTIQFDAC	16	28		LLILKTVLG	19
62		ATVLTLPRV	16	29 30		H V C A V F I F Y R L F H V A T H A	
63		G V A A V V R G A	16	30 31		AVLGNLTI	
64 65		- C L I V I I S A I - I C L S M V H R F	16	32		AIFWFNSTT	
66	257 279	NIYLLVPPV	16 16	33		I I S A I G L D S	
67	30	A F P L C S L Y L	15	34	220		
68	101	LOIFAIHS L	15	35		YVPFIGLSM	
69	115	TV L L A M A F D	15 15	36		Y L I A V L C N L	
70		A I C H P L R H A	15 15	37		<u> </u>	
71	153	R G A A L M A P L	15 15	38		<u> </u>	
72	163	V F I K Q L P F C	15	39	131	<u>P L R H A T V L T</u>	
73	215	LISFSYLLI	15	40	136	T V L T L P R V T	17
74	216	ISFSYLLIL	15	41	180	<u> </u>	17
75	225	KTVLGLTRE	15	42	201	-C-I-I-V-I-I-S-A-I	17
7-6	272	-PLPVILANI	15	43		<u>-S-L-L I S F S Y L</u>	
77		-LLVPPVLNP	15	44		<u>FIGLSMVHR</u>	
78	286	PV-LN-PIVYC	15	45	261	MVHRFSKRR	17

_162

H	LA-A3	-nonomero		H	LA-B*(9702 nonomers	
_				-			
	Pos	1 2 3 4 5 6 7 8 9	score		Pes	123456789	score
46	276	<u>-I L A N I Y L L V</u>	17	22	159	APLPVFIKO	14
47	281	YLVPPVLN	17	23	222	- L I - L K T V L G L	14
48	286	- P V L N P I V Y C	1.7	24	271	SPLPVILAN	14
49	288	-L N P I V Y G V K	17	25	25	AQFWLAFPL	13
50	309	<u> </u>	17	26	109	LSGMESTVL	13
51	1		16	27	124	RYVAICHPL	13
52	56	-S L H E P M Y I F	16	28	216	ISFSYLLIL	13
53	70	-G-I D I L I S T S	16	29	268	RRDSPLPVI	13
54	72		16	30	280	<u> </u>	13
55	115	TVLLAMAFD	16	31	11	ATYFILIG L	12
56	125	YVAICHPLR	16	32	34	-C-S-L-Y-L-I-A-V-L	· 12
57	144	T K I C V A A V V	16	33	57	-LHEPMYIFL	12
58	167	QLPFCRSNI	16	34	76	STSSMPKML	12
59	175	$-I$ L \underline{S} H S \underline{Y} \underline{C} L H	16	35	142	RVTKIGVAA	12
60	195	<u> </u>	16	36	151	-V V R G A A L M A	12
61	197	NVYYC <u>L</u> VI	16	37	190	ACDDIRVN V	12
62	210	-C L <u>D</u> S L <u>L</u> <u>I</u> S F	16	38	194	-I R V N V V Y G L	. 12
63	282	LLVPPVLN P	16	39	206	-I S A I G L D S L	12
64	299	EIRQRILRL	16	40	207	SAIGLDSLL	12
65	301	-R Q R I L R L F H	16	41	220	<u> </u>	12
66	16	LIGLPGLEE	15	42	267	KRRDSPLP V	12
67	46	TIIYIVRTE	15	43	304	I L R L F H V A T	12
68	102	QIFAI <u>HS</u> LS	15	44	14	FILIGLPGL	11
69	193	-D I R V N V V Y G	15	45	23	EEAQFWLAF	11
70	208	AIGLD<u>S</u>LLI	15	46	37	Y L I A V L G N L	11
71	223	-I L <u>K</u> T V <u>L</u> <u>C</u> L-T	15	47	40	AVLCNLTII	11
72	237	- К А <u>F</u> С Т <u>С V</u> S Н	15	48	77	TSSMPKMLA	11
		0.500		49	78	SSMPKMLAI	11
#	TV-R.	0702 nonomers		50	80	MPKMLAIFW	11
	Pos	100156700	score	51	92	TTIQFDACL	11
	120	1 2 3 4 5 6 7 8 9	00	52	112	MESTVLLAM	11
1	130	HPLRHATVL	22	53	119	AMAFDRYVA	11
2	59	, EPMYIFLCM	21	54	127	AICHPLRHA	11
3 4	168 289		20	55	131	PLRHATVLT	11
4 5		NPIVYCVKT DPNGNESSA	19	56	155	AALMAPLP V	11
5	3 19	LPCLEEAOF	18	57	157	<u>- L M A P L P V F I</u>	11
7	140	LPRVTKICV	18 18	58	181	-C L H Q D V M K L	11
8	284	V P P V L N P I V	10 17	59	203	-I V I I S A I G L	11
9	31	FPLCSLYLI	16	60	208	AIGLDSLLI	11
10	31 25 4	V P F I G L S M V	16	61	213	SLLISFSYL	11
11	269	RDSPLPVIL	16	62	248	AVFIFYVPF	11
12	149	AAVVRCAAL	15	63	265	FSKRRDSPL	11
13	153	RCAALMAPL	15	64	-	VILANIYLI	11
14	156	ALMAPLPVF	15	65	285	-PPVLNPIV¥	11
15	251		15	ı	I.A-R*	98-nonomers	
16	299	EIRQRILRL	15	=			
4.7			10		Pos		score

3 149

-1 2 3 4 5 6 7 8 9

1 299 -E I R Q R I L R L

2 265 F S K R R D S P L

4 168 LPFCRSNIL

14

14

14

14

14

31

29

24

24

<u>sd-63712</u> 163

30 AFPLCSLYL

132 LRHATVLTL

20 110 S G M E S T V L L

28 WLAFPLCSL

17

18

19

21

<u> </u>	LA-B*	08 nonomers		<u>#</u>	LA-B+1	1510 nonomers
	Pos	-1 2 3 4 5 6 7 8 9	score	;	Pos	-1 2 3 4 5 6 7 8 9
5	294	-C V K T K E I R Q	21	18	14	FILICLPCL
6	120	-MAFDRYVAI	20	19	28	WLAFPLCSL
7	292	V Y G V K T K E I	20	20	66	CMLSCIDIL
8	21	G L E E A Q F W L	19	21	76	STSSMPKML
9	78	SSMPKMLAI	19	22	92	-T T I Q-F D A C L
10	160	-P-L-P-V-F-1-K-Q-L	19	23	109	LSCMESTVL
11	186	-V M K L A C D D I	18	24	130	-H-P L R H A T V L
12	213	S L L I S F S Y L	18	25	132	-LRHATVLT-L
13	221	- L - L - K T - V - L - G	18	26	149	-A A V V R C A-A L
14	296	KTKEIRQRI	18	27	153	RGAALMAPL
15	297	TKEIRQRIL	18	28	160	PLPVFIKQL
16	130	HPLRHATVL	17	29	181	CLHQDVMKL
17	$\frac{181}{1}$	-C-L-H-Q-D-V-M-K-L	17	30	182	-L # Q D V M K L A
18	. 223	-I-L K T V L C L T	17	31	203	-I V I I S A I C L
19	28	-W L A F P L C S L	16	32	216	-I-S-F-S-Y-L-I-L
20	37	Y L I A V L C N L	16	33	222	LILKTVLCL
21	56	SLHEPMYIF	16	34	275	<u> </u>
22	80	-M P K M L A I F W	16	35	37	Y L I A V L C N L
23	162	PVFIKQLPF	16	36	49	YIVRTEHSL
24	201	GLIVIISAI	16	37	93	TIQFDACLL
25	207	SAIGLDSLL	16	38	101	-L-Q-I F A I H S L
26	214	LLISFSYLL	16	39	129	-C H P L R H A T V
27	220	Y L L I L K T V L	16	40	133	RHATVLTLP

score

11

11

11

25

score

15 138 -L T L P R V T K I 15 Pos -1-2 3 4 5 6 7 8 9 34 164 FIKQLPFCR 15

15

16

16

16

15

41 177 S H S Y C L H Q D

42 207 -S-A I G L D S L L

43 257 - I G L S M V H R F

1 194 -I-R V N V V Y C L

HLA-B*2705 nonomers

28 233 E A Q A K A F G T

29 275 V I L A N I Y L L

30 304 I L R L F H V A T

35 222 LILKTVLCL

FILICLPCL

SCMESTVLL

31

32

33

sd-63712

14

110

55	222		10	-			
				2	268	RRDSPLPVI	24
H	LA-B*	1510 nonomers		3	132	LRHATVLTL	23
	Pos		score	4	300	IRQRILRL F	23
		123456789	50010	5	305	LRLFHVATH	23
1	57	LHEPMYIFL	23	6	231	-T-R E A Q A K A F	21
2	244	SHVCAVFIF	17	7	34	-C-S-L Y L I A V-L	18
3	269	RDSPLPVI L	16	8	299	EIRQRILRL	18
4	280	<u>- I Y L L V P P V L</u>	16	9	6	-C N-E S S A T Y F	17
5	262	VHRFSKRRD	15	10	66	CMLSGIDIL	17
6	299	-EIRQRILRL	15	11	162	-P V F I K Q L P F	17
7	106	-I H S L S C M E S	14	12	207	SAIGLDSLL	17
8	206	-I-S-A I G L D S L	14	13	210	-C-L-D-S-L-L-I-S-F	17
9	220	Y L L I L K T V L	14	14	220	-Y L L I L K T V L	17
10	$\frac{251}{2}$	IFYVPFICL	14	15	237	-K-A-F-C-T-C-V-S-H	17
11	297	TKEIRQRIL	14	16	269	R D S P L P V I L	17
12	21	CLEEAQFWL	13	17	280	IYLLVPPVL	17
13	34	-C-S-L-Y-L-I-A-V-L	13	18	295	VKTKEIRQR	17
14	54	EHSLHEPM¥	13	19	11	ATYFILICL	16
15	110	SCMESTVLL	13	20	14	FILIGLPGL	16
16	194	IRVNVYCL	13	21	21	G L E E A Q F W L	16
17	8	ESSATYFI L	12	22	25	AQFWLAFPL	16

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UT A	P+27	Ω.E	nonomers
111111	7	5	HOHOMELS

HLA-B*2705 nonomers

•	Pos		score		Pos		score
	105	1 2 3 4 5 6 7 8 9	30016		105	-1 2 3 4 5 6 7 8 9	BCOLE
23	37	Y L I A V L-C N L	16	75	293	Y G V K T K E I R	14
24	92	TTIQFDACL	16	76	301	RQRILRLFH	14
25	101	LQIFAIHSL	16	77	302	QRILRLFHV	14
26	124	RYVAICHPL	16	78		N C N E S S A T Y	13
27		HPLRHATVL	16	79		EEAQFWLAF	13
28		PRVTKIGVA	16	80		WLAFPLCSL	13
29	153	R G A A L M A P L	16	81		NLTIIYIVR	13
30	181	CLHQDVMKL	16	82		-VRTEHSLHE	13
31		GLIVIISAI	16	83		SLHEPMYIF	13
32		IVIISAICL	16	84		PMYIFLCML	13
33		ISFSYLLIL	16	85		-DILI-S-T-S-M	13
34		-L I L K T V L G L	16	86		LISTSSMPK	13
35	255	PFICLSMVH	16	87	_	-I S T S S M P K M	13
36	257	ICLSMVHRF	16	88		ACLLQIFAI	13
37		VILANIYLL	16	89		FAIHSLSGM	13
38		IIYIVRTEH	15	90		SCMESTVLL	13
39		LSCMESTVL	15	91		V L L A M A F D R	13
40		STVLLAMAF	15	92		VAICHPLRH	13
41		DRYVAICHP	15	93		-A-V-V-R-C-A-A-L	13
42		KIGVAAVVR	15	94		MAPLPVFIK	13
43		ALMAPLPVF	15	95		FIKQLPFCR	13
44		LPFCRSNIL	15	96		FCRSNILSH	13
45		RSNILSHSY	15	97		CRSNILSHS	13
46		V V Y C L I V I I	15	98	-	-M K L A C D D I R	13
47		ISAIGLDSL	15	99		SFSYLLILK	13
48		GLTREAQAK	15	100		LKTVLGLTR	13
49		A V F I F Y V P F	15	101		- C V S H V C A V F	13
50		IFYVPFIGL	15	102		FIGLSMVHR	13
51		PVILANIYL	15	103		MVHRFSKRR	13
52		PIVYGVKTK	15	104		YIVRTEHSL	12
53		KEIRQRILR	15	105		L H E P M Y I F L	12
54		LPGLEEAQF	14	106		WFNSTTIQF	12
55		LAFPLCSLY	14	107		FDACLLQIF	12
56	30	AFPLCSLYL	14	108		HATVLTLPR	12
57 58		I A V L G N L T I	14	109		VRGAALMAP SYCLHQDVM	12
58 59		•	14	110		NVVYGLIVI	12 12
55		SMPKMLAIF PKMLAIFWF	14 14	111		SHVCAVFIF	
61		CLLOIFAIH	14 14	112 113		F S K R R D S P L	12 12
62		V L T L P R V T K	14 14	114		LPVILANI¥	12
63		LTLPRVTKI	14 14	115		PPVLNPIV¥	12
64		- A V V R G A A L M	14 14	116		LNPIVYCVK	12
65		PLPVFIKQL	14	117		- K T K E I R O R I	12
66		NILSHSYCL	14	118		TKEIRORIL	12
67		Y C L H Q D V M K	14	110	237		±£
68		DDIRVNV Y	14				
69		DS L L I S F S ¥	14		HLA-B*	2709 nonomers	
70		SLLISFSYL	14		Poo		860**
71		LLISFSYLL	14		Pes	123456789	score
72		SMVHRFSKR	14	1	194	-I R V N V V Y C L	24
73		HRFSKRRDS	14	2	268	-RRDSPLP-VI	24
74		KRRDSPLPV	14	3	132	-LRHATVLTL	22
			- 1	165			
<u>sd-63</u>	114			165			•

HI A	_ D 1	12700	-nonomers
111111	7	2703	-1101101110113

_	Pos		score	HLA-B*5101 nonomers			
	103	-1 2 3 4 5 6 7 8 9	50010		Pos		score
4	267	KRRDSPLPV	21			-1 2 3 4 5 6 7 8 9	0000
5	300	-I R Q R I L R L F	20	1	39	-IAVLGNLTI	26
6	231	TREAQAKAF	19	2	31	FPLCSLYLI	25
7	302	QRILRLFHV	19	3	120	MAFDRYVAI	2 4
용	124	RYVAICHPL	16	4	130	HPLRHATVL	23
9	269	RDSPLPVIL.	16	5	118	- L A M A F D R Y V	22
10	43	-C-N-L-T-I-Y-I-V	15	6	140	LPRVTKIGV	22
11	216	-I-S-F-S-Y-L-I-L	15	7	$\frac{155}{1}$	- A - L - M - A - P - L - P - V	22
12	11	ATYFILICL	14	8	42	-L G N L T I I Y I	21
13	25	AQFWLAFPL	14	9	254	-V P F I C L S M V	21
14	153	RGAALMAPL	14	10	284	V P P V L N P I V	21
15	174	NILSHSYCL	14	11	168	LPFCRSNIL	20
16	222	-LILKTVLGL	14	12	235	QAKAFCTCV	20
17	257	-I G L S M V H R F	14	13	138	LTLPRVTKI	19
18	280	-I Y L L V P P V L	14	14	159	APLPVFIKQ	18
19	6	CNESSATYF	13	15	189	LACDDIRVN	18
20	14	FILIGLPGL .	13	16	198	V V Y C L I V I I	. 18
21	21	-CLEEAQFW-L	13	17	277	LANIYLLVP	. 18
22	66	-CMLSGIDI-L	13	18	207	SAIGLDSLL	17
23	130	HPLRHATVL	13	19	283	LVPPVLNPI	17
24	201	-G-L-I-V-I-S-A-I	13	20	63	IFLCMLSCI	16
25	203	-IVIISAIC L	13	21	86	IFWFNSTTI	16
26	214	-L-L I S F S Y L L	13	22	110	S G M E S T V L L	16
27	251	-I-F Y V P F I G-L	13	23	144	TKICVAAVV	16
28	263	HRFSKRRDS	13	24	149	AAVVRGAAL	16
29	275	VILANIYLL	13	25	197	NVYGLIVI	16
30	305	LRLFHVATH	13	26	271	SPLPVILAN	16
31	30	AFPLCSLYL	12	27	280	I Y L L V P P V L	16
32	34	CSLYLIAVL	12	28	3	DPNCNESSA	15
33	37	Y L I A V L C N L	12	29	40	AVLGNLTII	15
34	51	VRTEHSLHE	12	30	97	DACLLOIFA	15
35	60	PMYIFLCML	12	31	132	LRHATVLTL	15
36	75	ISTSSMPKM	12	32	222	-LILKTVLCL	15
37	93	TIQFDACLL	12	33	279	NIYLLVPPV	15
38	123	-D-R Y V A I C H P	12	34	285	PPVLNPIV Y	15
39	135	ATVLTLPRV	12	35	289	N P I V Y G V K T	15
40	138	-LT-LPRVTKI	12	36	9	SSATYFILI	14
41	149	AAV R G A A L	12	37	65	LCMLSCIDI	14
42	155	AALMAPLPV	12	38	84	LAIFWFNST	14
43	168	LPFCRSNIL	12	39	126	VAICHPLRH	14
44	181	C-L-H-Q-D-V-M-K-L	12	40	157	L M A P L P V F I	14
45	188	K-L-A-C-D-D I R V	12	41	158	MAPLPVFIK	14
46	190	A-C-D-I-R V N V	12	42	191	CDDIRVNVV	14
47	195	RVNVYGL-I	12	43	200	Y G L I V I I S A	14
48	210	-G L D S L L I S F	12	44	209	I G L D S L L I S	14
49	213	-S L L I-S F S Y L	12	45	215	LISFSYLLI	14
50	220	Y L L I L K T V L	12	46	219	SYLLILKTV	14
51	248	AVFIFYVPF	12	47	220	Y L L I L K T V L	14
52	279	NIYLLVPPV	12	48	237	KAFGTCVSH	14
53	287	V L N P I V Y G V	12	4-9 5-0	247	GAVFIFYVP	14
54	296	KTKEIRQRI	12	50	249	V F I F Y V P F I	14
55	299	EI-RQRILRL	12	51	251	IFYVPFIGL	14
d-637	12		166				
				•			

UT 7 _ D 1	F 5 1 0 1	nonomers
mmr o	2101	HOHOMELS

HLA-B*5101 nonomers

Second S	_	Pos	1 2 3 4 5 6 7 0 0	score		Pes	1 2 2 4 5 6 7 0 0	score
\$\frac{80}{80} = \frac{80}{80} - \frac{80}{10} - \frac{80}{10} - \frac{1}{10} - \	E 0	257	1 2 3 4 5 6 7 8 9	1.4	1471	20	-1-2-3-4-5-6-7-8-9	21
Second S								-
Second S								
Second S	-							
\$\frac{5}{5}\$\$ \$\frac{6}{1}\$\$ \$\frac{1}{1}\$\$ \$\frac					•			
\$8 67 ***RISCIPTI*** 13								
### 1472 1472 1473 1474 1475	_							
60 95 QFDACLLQT 13 1479 38 39 IAVLGNLTT 19 61 98 ACLLQIFAT 13 1480 39 45 LTIIYIVRT 19 62 104 FAIHSLSCM 13 1481 40 92 TTIQFDACL 19 63 1446 ICVAAAVVRCA 13 1481 40 92 TTIQFDACL 19 64 148 VAAVVRCAA 13 1482 41 110 SCMESTVLL 19 65 153 RCAALHAPL 13 1484 43 132 LRHATVLTL 19 66 233 EACAKAFCT 13 1486 45 155 ALMAPLPV 19 66 233 EACAKAFCT 13 1486 45 155 ALMAPLPV 19 69 296 KTKEIRORT 13 1486 45 155 ALMAPLPV 19 69 296 KTKEIRORT 13 1486 45 155 ALMAPLPV 19 19 18 18 18 18 18 18 18 18 18 18 18 18 18								
Second						_		
1491 40 92 TTIOFDACL 19 18 140 140 140 140 150 1			-		***************************************			
146								
1488								
1486 453					1483			
149			RCAALMAPL		1484			
1486 45 155 A L M A P L P V 19	66	233	<u> </u>		1485			
1490 49 46 156 ALMAPLEVE 19 19 19 19 19 19 19 1	67	243	VSHVCAVFI	13	1486			
SEQ ID SECONDARY SECON	68	292	VYCVKTKEI	13	1487			
1490 1490	69	296	KTKEIRQRI	13	· <u>1488</u>		203 I V IISAIG I	
NO: HIA-A+0201 nonomers 1490 99 216 ISFSYLLII 19 19 19 19 19 19 19	<u>SEO</u>				<u>1489</u>			
Pos		•••			<u>1490</u>		216 ISFSYLLII	19
12 3 4 5 6 7 8 9 19 1442 1 287 VLNPIVYGV 30 1494 51 221 SECRET 19 1443 2	NO:	<u>H1</u>	LA-A*U2UI nonomers		<u>1491</u>	<u>50</u>	219 SYLLILKTV	19
1442 1 287 V L N P I V Y G V 30 1494 53 17 I G L P G L E E A 18 1444 3 28 W L A F P L C S L 28 1496 55 34 C S L Y L I A V L G N L T 18 1445 5 222 L I L K T V L G L L 28 1496 55 34 C S L Y L I A V L L R R I L I G L P G L 28 1496 55 34 C S L Y L I A V L L R R I L I G L P G L 28 1496 55 34 C S L Y L I A V L L R R I L I G L P G L 28 1496 55 34 C S L Y L I A V L L R R I L I G L P G L 28 1496 55 34 C S L Y L I A V L L R R I L I G L P G L 28 1496 55 34 C S L Y L I A V L L R R I L I G L P G L 28 1496 55 34 C S L Y L I A V L L R I R I L I G L P G L 28 1496 55 34 C S L Y L I A V L L R I R I R I G L P G L 28 1498 57 43 G N L T I I Y I V R I R I R I G L P G L D S L L I S F S Y L L L S C G L D S L L I S S G M E S T V 26 1500 59 118 L A M A F D R Y V R I R I R I G L P G L D S L L I S F S Y L L L C G I S L I S F S Y L L L L L L L L L L L L L L L L L L			Pos 1 2 2 4 5 6 7 9 6	score	<u>1492</u>	<u>51</u>	221 L L L L K T V L G	19
1443 2	1442	1		=			223 I L K T V L G L 1	
1444 3 28							17 IGLPGLEE	<u>18</u>
1446 5 222 LILKTVLGNL 28 1497 56 38 LIAVLGNLT 18 1447 6 66 66 CMLSGIDIL 26 1499 58 85 AIFWFNSTT 18 1448 7 108 SLSGMESTV 26 1500 59 118 LAMAFDRYV 18 1450 9 201 GLIVIISAI 26 1501 60 194 IRVNVVVGLISF 18 1451 10 214 LLISFSYLL 26 1500 62 215 LISFSYLL 18 1452 11 27 VILANIYLL 26 1500 62 215 LISFSYLL 18 1452 11 27 VILANIYLL 25 1506 64 254 VPFIGLSMV 18 1455 12 257 VILANIYLL 25 1506 64 254 VPFIGLSMV 18 1455 13 220 YLLILKTVL 25 1506 66 65 15 IFLCMLSGI 17 1455 14 276 ILANIYLL 25 1506 66 65 15 IFLCMLSGI 17 1455 16 138 LTLFRVTK 24 1509 68 93 TIQFDACL 17 1456 17 213 SLLISFSYL 24 1509 68 93 TIQFDACL 17 1460 19 143 VTKIGVAAV 23 1511 70 111 GMESTVL A 17 1461 20 18 KLACDDIRV 23 1515 71 120 MAFDRYVAI 17 1466 22 21 198 VVYGLIVII 22 1515 76 31 FPLCSLYL 17 1466 22 21 198 VVYGLIVII 22 1515 76 31 FPLCSLYL 16 1466 25 11 ATYFIL LFRV 22 1515 76 31 FPLCSLYL 17 1466 25 15 ATYFIL 15 15 15 15 76 31 FPLCSLYL 17 1466 25 15 15 15 15 15 15 15								
1446 5 222 LILKTVLGL 28 1498 56 38 LIAVLGNLT 18 1447 6 66 CMLSGIDIL 26 1499 58 85 AIFWINSTT 18 1448 7 108 SLSGMESTV 26 1500 59 118 LAMAFDRYV 18 1449 9 18 CLHODVMKL 26 1501 60 194 IRVNVVVGL 18 1450 9 201 GLIVIISAI 26 1500 61 210 GLDSLLISF 18 1451 10 214 LLISFSYLL 26 1503 62 215 LISFSYLL 18 1452 11 275 VILANIYL 26 1504 63 246 VCAVFIFYV 18 1453 12 157 LMAPLPVFI 25 1505 64 254 VPFIGLSMV 18 1453 12 157 LMAPLPVFI 25 1506 65 15 LLIGLSMV 18 1455 13 220 YLLILKTVL 25 1506 65 15 LLIGLSMV 18 1455 13 220 YLLILKTVL 25 1506 65 15 LLIGLSMV 18 1455 15 279 NIYLLVPPV 25 1506 65 15 LLIGLSMV 17 1458 17 213 SLLISFSYL 24 1509 68 93 TIOFDACL 17 1458 17 213 SLLISFSYL 24 1510 69 98 ACLLOIFAI 17 1460 19 143 VTKIGVAAV 23 1511 70 111 GMESTVLLA 17 1460 19 143 VTKIGVAAV 23 1511 70 111 GMESTVLLA 17 1460 22 188 KLACDDIRV 23 1511 70 111 GMESTVLLA 17 1460 22 188 KLACDDIRV 23 1511 70 111 GMESTVLLA 17 1460 22 188 KLACDDIRV 23 1511 70 111 GMESTVLLA 17 1460 22 188 KLACDDIRV 23 1511 70 111 GMESTVLLA 17 1460 22 188 KLACDDIRV 23 1511 70 111 GMESTVLLA 17 1460 25 11 ATYFILIGL 21 1515 74 226 TVLGLTREA 17 1460 25 11 ATYFILIGL 21 1515 75 281 YLLVPPVLN 17 16 1468 27 135 ATVLTLPRV 21 1520 79 78 SMPKMLA 1 16 1460 28 160 PLPVFIKOL 21 1520 79 78 SMPKMLA 1 16 1460 28 160 PLPVFIKOL 21 1520 80 79 SMPKMLA 1 16 1400 14								
1448 7 108		≐ 5						
1448 7								
1449 8	1448	<u>=</u> 7						
1450 9 201 G L I V I I S A I 26 1502 61 210 G L D S L L I S F 18 1452 11 275 V I L A N I Y L L 26 1503 62 215 L I S F S Y L L I 18 1453 12 157 L M A P L P V F I 25 1505 64 254 V P F I G L S M V 18 1453 12 157 L A N I Y L L V 25 1506 65 15 I L I G L D G L E I I I I I I I I I I I I I I I I I I	1449	8						
1451 10 214	1450	9						
1452 11 275	<u>1451</u>							
1453 12 157 L M A P L P V F I 25 1505 64 254 V P F I G L S M V 18 1454 13 220 Y L L I L K T V L 25 1506 65 15 I L I G L P G L E 17 1455 14 276 I L A N I Y L L V P P V 25 1507 66 63 I F L C M L S G I 17 1456 15 279 N I Y L L V P P V 25 1508 67 72 D I L I S T S S M 17 1457 16 138 L T L P R V T K I 24 1509 68 93 T I Q F D A C L L 17 1458 17 213 S L L I S F S Y L 24 1510 69 98 A C L L Q I F A I 17 1459 18 49 Y I V R T E H S L 23 1511 70 111 G M E S T V L L A 17 1460 19 143 V T K I G V A A V 23 1512 71 120 M A F D R Y V A I 17 1462 21 198 V V Y G L I V I I 23 1513 72	<u>1452</u>	_	275 VILANIYLI					
1454 13 220	<u>1453</u>	12	157 LMAPLPVFI	<u>25</u>				====
1455		13	220 YLLILKTVI					
1456 15 279 N I Y L L V P P V 25 1508 67 72 D I L I S T S S M 17 1457 16 138 L T L P R V T K I 24 1509 68 93 T I Q F D A C L L 17 1458 17 213 S L L I S F S Y L 24 1510 69 98 A C L L Q I F A I 17 1460 19 143 V T K I G V A A V 23 1511 70 111 G M E S T V L L A 17 1461 20 188 K L A C D D I R V 23 1513 72 167 Q L P F C R S N I 17 1462 21 198 V V Y G L I V I I 23 1514 73 197 N V V Y G L I V I I 17 1463 22 21 G L E E A Q F W L 22 1515 74 226 T V L G L T R E A 17 1465 24 206 I S A I G L D S L 22 1516 75 281 Y L L V P P V L N 17 16 1466 25 11 A T Y F I L I G L 21 1518 77 56 S L H E P M Y I F 16 16 1468 27 135 A T V L T L P R V 21 1520 79 78 S S M P K M L A I F 16 1469 28 160 P L P V F I K Q L 21 1521 80 79 78 S M P K M L A I F 16 16 16 16 16 16 16		<u>14</u>	<u>276 </u>					
1457 16 138 L T L P R V T K I 24 1509 68 93 T I Q F D A C L L 17 1458 17 213 S L L I S F S Y L 24 1510 69 98 A C L L Q I F A I 17 1469 19 143 V T K I G V A A V 23 1511 70 111 G M E S T V L L A 17 1461 20 188 K L A C D D D I R V 23 1513 72 167 Q L P F C R S N I 17 1462 21 198 V V Y G L I V I I 23 1513 72 167 Q L P F C R S N I 17 1463 22 21 198 V V Y G L I V I I 23 1514 73 197 N V V Y G L I V I 17 1464 23 40 A V L G N L T I I 22 1515 74 226 T V L G L T R E A 17 1465 24 206 I S A I G L D S L 21 1516 75 281 Y L L V P P V L N 17 1466 25 11 A T Y F I L I G L 21 1518		<u>15</u>	279 N I Y L L V P P V	<u>25</u>				
1458 17 213 S			<u> 138 </u>			_		
1459 18 49 Y I V R T E H S L 23 1460 19 143 V T K I G V A A V 23 1461 20 188 K L A C D D I R V 23 1462 21 198 V V Y G L I V I I 23 1463 22 21 G L E E A Q F W L 22 1464 23 40 A V L G N L T I I 22 1465 24 206 I S A I G L D S L 22 1466 25 11 A T Y F I L I G L 21 1468 27 135 A T V L T L P R V 21 1468 27 135 A T V L T L P R V 21 1469 28 160 P L P V F I K Q L 21 1470 29 174 N I L S H S Y C L 21 1470 29 174 N I L S H S Y C L 21 1460 25 110 15 H S A I H S L S G M								17
1460 19			———————————————————————————————————————					
1461 20 188 K L A C D D I R V 23 1513 72 167 Q L P F C R S N I 17 1462 21 198 V V Y G L I V I I 23 1514 73 197 N V V Y G L I V I 17 1463 22 21 G L E E A Q F W L 22 1515 74 226 T V L G L T R E A 17 1465 24 206 I S A I G L D S L 22 1516 75 281 Y L L V P P V L N 17 1466 25 11 A T Y F I L I G L 21 1518 77 56 S L H E P M Y I F 16 1468 27 135 A T V L T L P R V 21 1519 78 70 G I D I L I S T S 16 1469 28 160 P L P V F I K Q L 21 1520 79 78 S S M P K M L A I F 16 1470 29 174 N I L S H S Y C L 21 1522 81 104 F A I H S L S G M 16								
198 V V Y G L I V I I 23 197 N V V Y G L I V I I 17 1463 22 21 G L E E A Q F W L 22 1515 74 226 T V L G L I V I R E A 17 17 1865 24 206 I S A I G L D S L 22 1516 75 281 Y L L V P P V L N 17 1866 25 11 A T Y F I L I G L 21 1518 77 56 S L H E P M Y I F 16 16 1468 27 135 A T V L T L P R V 21 1519 78 70 G I D I L I S T S 16 1469 28 160 P L P V F I K Q L 21 1520 79 78 S M P K M L A I 16 16 1470 29 174 N I L S H S Y C L 21 1522 81 104 F A I H S L S G M 16 16 16 16 16 16 16					<u>1513</u>			
1464 23 40 A V L G N L T I I 22 1515 74 226 T V L G L T R E A 17 1465 24 206 I S A I G L D S L 22 1517 76 31 F P L C S L Y L I 16 1467 26 60 P M Y I F L C M L 21 1518 77 56 S L H E P M Y I F 16 16 1468 27 135 A T V L T L P R V 21 1520 79 78 70 G I D I L I S T S 16 1470 29 174 N I L S H S Y C L 21 1522 81 104 F A I H S L S G M 16 16 16 16 16 16 16					<u>1514</u>			
1465 24 206 1 S A I G L D S L 22 1516 75 281 Y L L V P P V L N 17 1466 25 11 A T Y F I L I G L 21 1518 77 56 S L H E P M Y I F 16 1468 27 135 A T V L T L P R V 21 1519 78 70 G I D I L I S T S 16 1469 28 160 P L P V F I K Q L 21 1520 79 78 S M P K M L A I 16 1470 29 174 N I L S H S Y C L 21 1522 81 104 F A I H S L S G M 16 sd-63712 167					<u> 1515</u>			
1466 25 11 A T Y F I L I G L 21 1517 76 31 F P L C S L Y L I 16 1467 26 60 P M Y I F L C M L 21 1518 77 56 S L H E P M Y I F 16 1468 27 135 A T V L T L P R V 21 1519 78 70 G I D I L I S T S 16 1469 28 160 P L P V F I K Q L 21 1520 79 78 S S M P K M L A I F 16 1470 29 174 N I L S H S Y C L 21 1522 81 104 F A I H S L S G M 16 sd-63712 167		_			<u>1516</u>	_		
1467 26 60 PMYIFIBIGE 21 1518 77 56 SLHEPMYIF 16 1468 27 135 ATVLTLPRV 21 1519 78 70 GIDILISTS 16 1469 28 160 PLPVFIKOL 21 1520 79 78 SSMPKMLAI 16 1470 29 174 NILSHSYCL 21 1522 80 79 SMPKMLAIF 16 1522 81 104 FAIHSLSGM 16 16 16 16 16 16 16					<u>1517</u>			
1468 27 135 A T V L T L P R V 21 1520 79 78 70 G I D I L I S T S 16 16 1469 28 160 P L P V F I K Q L 21 1520 79 78 S S M P K M L A I 16 1470 29 174 N I L S H S Y C L 21 1522 81 104 F A I H S L S G M 16 sd-63712 167						77	·	
1469 28 160 PLPVFIKQL 21 1520 79 78 SSMPKMLAI 16 1470 29 174 NILSHSYCL 21 1522 80 79 SMPKMLAIF 16 1522 81 104 FAIHSLSGM 16						<u>78</u>		
1470 29 174 N I L S H S Y C L 21 1521 80 79 S M P K M L A I F 16 sd-63712 167							<u> 78 </u>	
<u>sd-63712</u> <u>167</u> <u>1522</u> <u>81</u> <u>104</u> <u>FAIHSLSGM</u> <u>16</u>							<u>79 SMPKMLAI</u>	
		<u>= 3</u>	<u> </u>	<u> </u>	<u>1522</u>	<u>81</u>	104 FAIHSLSGN	
	sd-637	12	· ·		_167			

H	LA-B	5101	nonomers			SEO				
	Pos	1	2 3 4 5 6 7 8 9	ecere		<u>ID</u> NO:	<u>H</u>	LA A*(0203 nonomers	
<u>1523</u>	<u>82</u>	119	<u> </u>	<u>16</u>			_	Pos	_	score
<u>1524</u>	83		T K I G V A A V V			1569			1 2 3 4 5 6 7 8 9	
<u>1525</u>	84	147	G V A A V V R G A	<u>16</u>		1570	<u>21</u>	<u>230</u> 240	L T R E A Q A K A G T C V S H V C A	<u>ଧା ଧା ଧା ଧା</u>
1526	<u>85</u>		VMKLACDDI			1571	23	270 270	D S P L P V I L A	≟ 9
<u>1527</u> <u>1528</u>	<u>86</u>		LTREAQAKA			1572	22 23 24	303	RILRLFHVA	<u>=</u> <u>9</u>
1529	<u>87</u> 88		A F G T C V S H V V F I F Y V P F I			<u>1573</u>	25	306	RLFHVATHA	<u>9</u>
1530	89		Q R I L R L F H V			SEO.				
1531	90		RILRLFHVA			<u>SEO</u> ID				
<u>1532</u>	<u>91</u>	<u>18</u>	G L P G L E E A Q	<u>15</u>		NO:	H	LA-A2	5 nonomers	
<u>1533</u> 1534	<u>92</u>		SLYLIAVLG					Pos	100456700	score
$\frac{1534}{1535}$	<u>93</u> 94		LGNLTIIYI TIIYIVRTE			<u>1574</u>	1	<u>299</u>	1 2 3 4 5 6 7 8 9 E I R Q R I L R L	30
1536	9 <u>5</u>		SGIDILIST			1575	≛ 2	<u>233</u> 72	DILISTSSM	<u>30</u> 27
1537	96		STSSMPKML			<u>1576</u>	<u>=</u> <u>3</u>	248	AVFIFYVPF	<u>27</u>
<u>1538</u>	97	131	PLRHATVLT	<u>15</u>		<u>1577</u>	<u>4</u>	210	GLDSLLISF	<u>26</u>
1539 1540	<u>98</u>	<u>137</u>	V L T L P R V T K			1578	<u>5</u>	. 14	FILIGLPGL	<u>24</u>
<u>1540</u> 1541	<u>99</u> 100		RGAALMAPL			<u>1579</u> 1580	<u>6</u>	<u>56</u>	S L H E P M Y I F	<u>24</u>
	101		A C D D I R V N V C D D I R V N V			1581	1 2 3 4 5 6 7 8 9 0 10	$\frac{117}{222}$	L L A M A F D R Y L I L K T V L G L	<u>24</u> <u>24</u>
	102		VIISAIGLD			1582	<u>s</u>	245	HVCAVFIFY	24
	103	241	T C V S H V C A V	<u>15</u>		<u>1583</u>	10	11	ATYFILIGL	23 23
	<u>104</u>	<u>251</u>	I F Y V P F I G L			1584 1585	<u>11</u>	<u>37</u>	YLIAVLGNL	<u>23</u>
15.45	105		RDSPLPVIL			<u>1585</u> 1586	<u>12</u>	114 156	STVLLAMAF	<u>23</u>
45.40	<u>106</u> 107	<u>280</u> 306	I Y L L V P P V L R L F H V A T H A			1587	13 14	<u>156</u> 162	ALMAPLPVF PVFIKQLPF	2 <u>3</u> 23
	<u> </u>	300	<u> </u>	===		1588	15 15	181	CLHQDVMKL	<u>23</u>
SEO						<u>1589</u>	16	28	WLAFPLCSL	<u>22</u>
<u>ID</u> NO:	H	LA A*(0203 nonomers			1590	<u>17</u>	<u>92</u>	TTIQFDACL	<u>22</u> <u>22</u>
		<u>Pos</u>		80	ore	<u>1591</u> 1592	<u>18</u>	<u>160</u>	P L P V F I K Q L	<u>22</u>
1540			1 2 3 4 5 6 7 8	9 —		1593	<u>19</u> 20	203 213	IVIISAIGL SLLISFSYL	<u>22</u> <u>22</u>
<u>1549</u> 1550	<u>1</u> 2	148	VAAVVRGA		14	1594	<u>20</u> 21	275	VILANIYLL	<u>22</u>
1551	<u>≼</u> <u>3</u>	<u>119</u> <u>147</u>	A M A F D R Y V G V A A V V R G		<u>13</u> <u>12</u>	<u>1595</u>	22	193	DIRVNVVYG	21
1552	<u>4</u>	97	DACLLQIF		<u>11</u>	<u>1596</u>	<u>23</u>	242	CVSHVCAVF	<u>21</u>
<u>1553</u>	<u>=</u> <u>5</u>	127	AICHPLRH		10	<u>1597</u> <u>1598</u>	24	<u>76</u>	STSSMPKML	<u>20</u>
<u>1554</u>	<u>6</u>	<u>3</u>	DPNGNESS			1599 1599	<u>25</u> 26	<u>253</u> 274	YVPFIGLSM PVILANIYL	<u>20</u> 20
<u>1555</u> <u>1556</u>	<u>7</u> <u>8</u>	<u>17</u>	IGLPGLEE	_	<u>9</u>	1600	<u>27</u>	23	EEAQFWLAF	19
1557	<u>8</u>	<u>22</u> <u>32</u>	L E E A Q F W L P L C S L Y L I		9 .	1601	28	41	VLGNLTIIY	<u>19</u>
1558	<u>10</u>	<u> 77</u>	T S S M P K M L		<u>⊒</u> 9	<u>1602</u>	<u>29</u>	<u>49</u>	YIVRTEHSL	<u>19</u>
1559	11	90	NSTTIQFD		<u>≅</u> 9	1603	<u>30</u>	<u>150</u>	AVVRGAALM	<u>19</u>
<u>1560</u>	<u>12</u>	111	GMESTVLL		9	<u>1604</u> 1605	31 32	<u>174</u> 192	NILSHSYCL DDIRVNVVY	<u>19</u> <u>19</u>
1561	<u>13</u>	113	ESTVLLAM	_	<u>9</u>	1606	33	$\frac{132}{214}$	LLISFSYLL	19
<u>1562</u> <u>1563</u>	14 15	141 142	P R V T K I G V		ତ୍ୟା ତ୍ୟା ତ୍ୟା ତ୍ୟା ତ୍ୟା ତ୍ୟା ତ୍ୟା ତ୍ୟା	1607	34	251	IFYVPFIGL	<u>19</u>
$\frac{1563}{1564}$	<u>15</u> <u>16</u>	<u>142</u> <u>151</u>	R V T K I G V A V V R G A A L M		<u>⊒</u> 9	<u>1608</u>	<u>35</u>	<u>8</u>	ESSATYFIL	<u>18</u>
1565	<u>17</u>	182	LHQDVMKL		<u>≠</u> 9	1609	<u>36</u>	<u>21</u>	GLEEAQFWL	<u>18</u>
<u>1566</u>	18	200	YGLIVIIS		<u>=</u>	$\frac{1610}{1611}$	<u>37</u>	<u>45</u>	LTIIYIVRT	<u>18</u>
<u>1567</u>	<u>19</u>	<u>226</u>	TVLGLTRE		<u>9</u>	1612	<u>38</u> 39	<u>54</u> 59	E H S L H E P M Y E P M Y I F L C M	<u>18</u> <u>18</u>
<u>1568</u>	<u>20</u>	<u>228</u>	LGLTREAQ	<u>A</u>	<u>9</u>	1613	40	<u>33</u> 88	WFNSTTIQF	18 18
1.625	10				1.0		-	. —		

<u>sd-63712</u> _____168

SEQ ID					<u>SEQ</u> ID				
NO:	HLA	-A26	nonomers		NO:	<u>H</u>	LA-A3	nonomers	
	<u>Po</u>	<u>s</u>	1 2 3 4 5 6 7 8 9	score			<u>Pos</u>	1 2 3 4 5 6 7 8 9	score
<u> 1614</u>	<u>41</u>	<u>93</u>	TIQFDACLL	<u>18</u>	<u> 1659</u>	<u>5</u>	<u>290</u>	PIVYGVKTK	<u>24</u>
1615		<u>85</u>	DVMKLACDD	<u>18</u>	1660	5 6 7 8 9	<u>35</u>	S L Y L I A V L G	<u>23</u>
<u>1616</u> 1617		98	VVYGLIVII	<u>18</u>	<u>1661</u> 1662	<u>7</u>	<u>156</u>	ALMAPLPVF	<u>23</u>
1618		<u>62</u> 70	Y I F L C M L S G G I D I L I S T S	<u>17</u> 17	1663	<u>e</u> 9	<u>47</u> 50	I I Y I V R T E H I V R T E H S L H	<u>22</u> <u>22</u>
1619		7 9	SMPKMLAIF	<u> </u>	1664	<u>≟</u> 10	142	RVTKIGVAA	22
<u>1620</u>	<u>47</u>	96	FDACLLQIF	<u>17</u>	<u>1665</u>	11	151	V V R G A A L M A	<u>22</u>
1621		04	FAIHSLSGM	<u>17</u>	<u>1666</u>	<u>12</u>	<u>242</u>	<u>CVSHVCAV</u>	<u>22</u>
<u>1622</u> 1623		<u>38</u> 43	LTLPRVTKI	<u>17</u>	<u>1667</u> 1668	13 14	248 116	AVFIFYVPF	<u>22</u>
1624		04	V T K I G V A A V V I I S A I G L D	<u>17</u> 17	1669	15 15	<u>116</u> 192	<u>V L L A M A F D R</u> <u>D D I R V N V V Y</u>	<u>21</u> <u>21</u>
1625		12	DSLLISFSY	<u> </u>	1670	16	303	RILRLFHVA	<u>21</u>
<u>1626</u>	<u>53</u> <u>2</u>	20	YLLILKTVL	<u>17</u>	<u>1671</u>	<u>17</u>	304	ILRLFHVAT	<u>21</u>
1627 1622		56	FIGLSMVHR	<u>17</u>	<u>1672</u>	<u>18</u>	<u>108</u>	SLSGMESTV	<u>20</u>
<u>1628</u> 1629		83	LVPPVLNPI	<u>17</u>	<u>1673</u> 1674	<u>19</u>	<u>198</u>	VVYGLIVII	<u>20</u>
1630		<u>29</u> 40	LAFPLCSLY AVLGNLTII	<u>16</u> 16	$\frac{1674}{1675}$	<u>20</u> 21	<u>291</u> 15	<u> </u>	<u>20</u> <u>19</u>
1631		46	TIIYIVRTE	16	1676	22	44	NLTIIYIVR	<u>19</u>
<u>1632</u>	<u>59</u>	<u>52</u>	RTEHSLHEP	16	<u> 1677</u>	23	<u>73</u>	I L I S T S S M P	<u>19</u>
1633		<u>75</u>	ISTSSMPKM	<u>16</u>	1678	<u>24</u>	<u>74</u>	LISTSSMPK	<u>19</u>
<u>1634</u> 1635		<u>91</u>	STTIOFDAC	<u>16</u>	<u>1679</u> 1680	<u>25</u>	<u>99</u>	C L L Q I F A I H	<u>19</u>
1636		. <u>35</u> .47	ATVLTLPRV GVAAVVRGA	<u>16</u> 16	1681	<u>26</u> 27	<u>162</u> 203	P V F I K Q L P F I V I I S A I G L	<u>19</u> <u>19</u>
1637		01	GLIVIISAI	16	1682	28	221	LLILKTVLG	<u>19</u>
<u>1638</u>	<u>65</u> 2	57	IGLSMVHRF		1683	29	245	HVCAVFIFY	<u>19</u>
1639		79	NIYLLVPPV	<u>16</u> <u>16</u>	<u>1684</u>	<u>30</u>	<u>306</u>	RLFHVATHA	<u>19</u>
<u>1640</u> 1641		<u>30</u>	AFPLCSLYL	<u>15</u>	<u>1685</u> 1686	<u>31</u>	<u>40</u>	AVLGNLTII	<u>18</u>
$\frac{1642}{1642}$. <u>01</u> .15	LQIFAIHSL TVLLAMAFD	<u>15</u> <u>15</u>	1687	<u>32</u> <u>33</u>	<u>85</u> 205	A I F W F N S T T I I S A I G L D S	<u>18</u>
1643		27	AICHPLRHA	15	1688	<u>34</u>	220	YLLILKTVL	<u>18</u> <u>18</u>
<u> 1644</u>		53	RGAALMAPL	<u>15</u>	1689	<u>35</u>	253	YVPFIGLSM	<u>18</u>
<u>1645</u>	_ =	63	VFIKQLPFC	<u>15</u>	1690	<u>36</u>	<u>37</u>	Y L I A V L G N L	<u>17</u>
<u>1646</u> 1647		15	LISFSYLLI	<u>15</u>	1691	<u>37</u>	41	V L G N L T I I Y	<u>17</u>
1648		16 25	I S F S Y L L I L K T V L G L T R E	<u>15</u> <u>15</u>	<u>1692</u> <u>1693</u>	<u>38</u>	<u>117</u> <u>131</u>	<u>L L A M A F D R Y</u> P L R H A T V L T	17 17
1649		<u>72</u>	PLPVILANI	15 15	1694	<u>39</u> <u>40</u>	136 136	T V L T L P R V T	17 17
1650		82	LLVPPVLNP	<u>15</u>	1695	41	180	YCLHQDVMK	<u> </u>
1651	<u>78</u> <u>2</u>	<u>86</u>	PVLNPIVYG	<u>15</u>	1696	41 42 43 44 45 46	<u>201</u>	G L I V I I S A I	17 17 17 17 17 17
<u>1652</u> 1653		87	V L N P I V Y G V	<u>15</u>	1697	<u>43</u>	<u>213</u>	SLLISFSYL	<u>17</u>
1654		<u>96</u> 03	KTKEIRORI RILRLFHVA	<u>15</u> 15	<u>1698</u> <u>1699</u>	44	<u>256</u> 261	F I G L S M V H R M V H R F S K R R	<u>17</u> <u>17</u>
	<u> </u>	<u> </u>	<u> </u>	12	1700	46	276	I L A N I Y L L V	<u>= /</u> 17
<u>SEQ</u>					1701	47	281	Y L L V P P V L N	<u>17</u> <u>17</u>
<u>ID</u> NO:	HLA-	-A3	nonomers		<u>1702</u>	<u>48</u>	<u> 286</u>	PVLNPIVYG	<u>17</u>
	Po			score	1703	<u>49</u>	288	LNPIVYGVK	<u>17</u>
1655		_	1 2 3 4 5 6 7 8 9		<u>1704</u> <u>1705</u>	<u>50</u> <u>51</u>	<u>309</u> 1	H V A T H A S E P M V D P N G N E S	<u>17</u> 16
<u>1655</u> 1656	1 1	<u>37</u>	V L T L P R V T K	<u>30</u>	1706	<u>51</u>	<u>1</u> 56	SLHEPMYIF	<u>16</u> <u>16</u>
1657		<u>29</u> 45	G L T R E A Q A K K I G V A A V V R	<u>27</u> 26	1707	53	70	G I D I L I S T S	16
1658	<u>≚</u>	<u> </u>	AVVRGAALM	<u>20</u> 24	<u>1708</u>	<u>54</u>	72	DILISTSSM	<u>16</u> <u>16</u>
sd-637				169)				
= <u> </u>	·			•					

SEO ID NO:	<u>H</u>	LA-A3	nonomers		SEO ID NO:	<u>H</u>	LA-B*(0702 nonomers	
		<u>Pos</u>		score			Pos		score
1700			1 2 3 4 5 6 7 8 9		1754			1 2 3 4 5 6 7 8 9	
<u>1709</u> 1710	<u>55</u>	<u>115</u> 125	TVLLAMAFD	<u>16</u>	<u>1754</u> 1755	<u>28</u>	<u>216</u>	ISFSYLLI L	<u>13</u>
1711	<u>56</u> <u>57</u>	$\frac{123}{144}$	Y V A I C H P L R T K I G V A A V V	<u>16</u> <u>16</u>	1756	<u>29</u> <u>30</u>	<u>268</u> <u>280</u>	R R D S P L P V I I Y L L V P P V L	<u>13</u> <u>13</u>
1712	<u>58</u>	167	QLPFCRSNI	<u>16</u>	1757	<u>31</u>	<u>11</u>	ATYFILIGL	<u>12</u>
1713	59	175	I L S H S Y C L H	16	1758	32	34	C S L Y L I A V L	12
<u>1714</u>	<u>60</u>	195	RVNVVYGLI	<u>16</u>	<u>1759</u>	<u>33</u>	<u>57</u>	LHEPMYIFL	12
<u>1715</u>	<u>61</u>	<u> 197</u>	NVVYGLIV I	<u>16</u>	<u>1760</u>	<u>34</u>	<u>76</u>	STSSMPKML	<u>12</u>
<u>1716</u>	<u>62</u>	<u>210</u>	G L D S L L I S F	<u>16</u>	<u>1761</u>	<u>35</u>	<u>142</u>	RVTKIGVAA	<u>12</u>
<u>1717</u>	<u>63</u>	<u>282</u>	L L V P P V L N P	<u>16</u>	1762	<u>36</u>	<u>151</u>	V V R G A A L M A	<u>12</u>
<u>1718</u> 1719	<u>64</u>	<u>299</u>	EIRORILRL	<u>16</u>	1763	<u>37</u>	<u>190</u>	ACDDIRVN V	<u>12</u>
$\frac{1719}{1720}$	<u>65</u>	<u>301</u>	RQRILRLFH	<u>16</u>	<u>1764</u> <u>1765</u>	<u>38</u>	<u>194</u>	IRVNVVYGL	<u>12</u>
1721	<u>66</u> <u>67</u>	<u>16</u> 46	L I G L P G L E E T I I Y I V R T E	<u>15</u> <u>15</u>	$\frac{1765}{1766}$	<u>39</u> <u>40</u>	<u>206</u> 207	ISAIGLDSL SAIGLDSLL	<u>12</u>
1722	<u>68</u>	102	O I F A I H S L S	15 15	1767	41	220 220	YLLILKTVL	<u>12</u> <u>12</u>
1723	69	193	DIRVNVVYG	<u>15</u>	1768	42	267	KRRDSPLPV	<u>12</u>
1724	70	208	AIGLDSLLI	· <u>15</u>	1769	43	304	ILRLFHVAT	. 12
<u>1725</u>	<u>71</u>	223	I L K T V L G L T	<u>15</u>	<u>1770</u>	44	14	FILIGLPGL	11
<u>1726</u>	<u>72</u>	<u>237</u>	KAFGTCVSH	<u>15</u>	<u>1771</u>	<u>45</u>	<u>23</u>	E E A Q F W L A F	<u>11</u>
SEO.					<u>1772</u>	<u>46</u>	<u>37</u>	YLIAVLGNL	<u>11</u>
<u>SEQ</u> ID					1773	<u>47</u>	40	AVLGNLTII	<u>11</u>
NO:	H	LA-B*(0702 nonomers		<u>1774</u> 1775	48	77	T S S M P K M L A	<u>11</u>
		Pos		score	$\frac{1775}{1776}$	<u>49</u> 50	<u>78</u> 80	S S M P K M L A I M P K M L A I F W	<u>11</u>
1707			1 2 3 4 5 6 7 8 9		<u>1777</u>	<u>50</u>	<u>92</u>	TTIQFDACL	<u>11</u> <u>11</u>
<u>1727</u> 1728	1	<u>130</u>	HPLRHATVL	<u>22</u>	1778	<u>52</u>	<u>112</u>	MESTVLLAM	<u>11</u>
1729	2 ≥	<u>59</u> 168	E P M Y I F L C M L P F C R S N I L	<u>21</u>	1779	<u>53</u>	119	A M A F D R Y V A	<u>11</u>
1730	3 4 5 6 7	289	N P I V Y G V K T	<u>20</u> <u>19</u>	<u>1780</u>	. <u>54</u>	127	AICHPLRHA	<u>11</u>
1731	≐ 5	<u>203</u>	D P N G N E S S A	<u>18</u>	<u>1781</u>	<u>55</u>	<u>131</u>	PLRHATVLT	<u>11</u>
1732	<u>=</u>	<u>19</u>	LPGLEEAQF	<u>18</u>	<u>1782</u>	<u>56</u>	<u>155</u>	<u> </u>	<u>11</u>
<u>1733</u>	<u></u>	140	L P R V T K I G V	18	1783	<u>57</u>	<u>157</u>	L M A P L P V F I	<u>11</u>
<u>1734</u>	8	284	V P P V L N P I V	<u>17</u>	<u>1784</u> 1785	<u>58</u>	<u>181</u>	C L H Q D V M K L	<u>11</u>
<u>1735</u>	<u>9</u>	<u>31</u>	FPLCSLYL I	<u>16</u>	$\frac{1785}{1786}$	<u>59</u>	203 208	I V I I S A I G L A I G L D S L L I	<u>11</u>
<u>1736</u>	10	<u>254</u>	<u>V P F I G L S M V</u>	<u>16</u>	1787	<u>60</u> <u>61</u>	<u>208</u> 213	SLLISFSYL	<u>11</u> <u>11</u>
<u>1737</u> 1738	11	<u>269</u>	RDSPLPVIL	<u>16</u>	1788	62	248	AVFIFYVPF	<u>11</u>
$\frac{1730}{1739}$	<u>12</u> <u>13</u>	<u>149</u> 153	<u> </u>	<u>15</u>	<u>1789</u>	<u>63</u>	265	F S K R R D S P L	<u> </u>
1740	14 14	156 156	ALMAPLPVF	<u>15</u> <u>15</u>	<u>1790</u>	64	275	V I L A N I Y L L	<u>11</u>
1741	<u>15</u>	<u>251</u>	I F Y V P F I G L	<u>15</u>	<u>1791</u>	<u>65</u>	<u> 285</u>	PPVLNPIVY	<u>11</u>
1742	16	299	EIRQRILRL	<u>15</u>	SEQ				
<u>1743</u>	<u>17</u>	8	ESSATYFI L	<u>14</u>	ID ID				
<u>1744</u>	<u>18</u>	<u>28</u>	WLAFPLCSL	14	NO:	Ħ	LA-B*(08 nonomers	
<u>1745</u>	<u>19</u>	<u>30</u>	AFPLCSLYL	<u>14</u>			Pos		score
<u>1746</u>	<u>20</u>	<u>110</u>	SGMESTVL L	<u>14</u>	1700			1 2 3 4 5 6 7 8 9	
1747 1748	<u>21</u>	<u>132</u>	LRHATVLTL	<u>14</u>	1792 1793	1	<u>299</u>	EIRORILR L	<u>31</u>
<u>1748</u> 1749	<u>22</u>	<u>159</u>	APLPVFIKO	<u>14</u>	<u>1793</u> 1794	2	<u>265</u>	F S K R R D S P L	<u>29</u>
1750	<u>23</u> <u>24</u>	<u>222</u> 271	LILKTVLG L S P LPVILA N	<u>14</u> 14	1795	<u>3</u> 4	<u>149</u> 168	AAVVRGAAL LPFCRSNII.	<u>24</u>
1751	<u>25</u>	271 25	AQFWLAFPL	<u>14</u> <u>13</u>	1796	<u>4</u> <u>5</u>	<u> 294</u>	LPFCRSNIL GVKTKEIRQ	<u>24</u> <u>21</u>
1752	<u>26</u>	<u>109</u>	LSGMESTVL	<u>13</u>	1797	<u>≅</u> 6	$\frac{234}{120}$	MAFDRYVAI	<u>21</u> 20
1753	27	124	RYVAICHPL	13	1798	<u>~</u>	292	VYGVKTKEI	20
<u>sd-637</u>				170					

<u>SEQ</u> <u>ID</u> NO: HL	A-B*08 nonomers	SEQ ID NO: HLA-B*1510 nonomers	
1799 8 1800 9 1801 10 1802 11 1803 12 1804 13 1805 14 1806 15 1807 16 1808 17 1809 18 1810 19 1811 20 1812 21 1813 22 1814 23 1815 24 1816 25 1817 26	1	Score Pos 1	12 12 12 12 12 12 12 12 12 12 12 12 12 1
1819 28 1820 29 1821 30 1822 31 1823 32 1824 33 1825 34 1826 35	233	16	11 11 11 11 11 11 11
<u>ID</u> NO: HL	A-B*1510 nonomers	<u>Pos</u> <u>1 2 3 4 5 6 7 8 9</u>	score
1827 1828 2 1829 3 1830 4 1831 5 1832 6 1833 7 1834 1835 9 1836 10 1837 11 1838 12 1839 133 14 15 16 1831	S 1 2 3 4 5 6 7 8 9 57 L H E P M Y I F L 244 S H V C A V F I F 269 R D S P L P V I L 280 I Y L L V P P V L 262 V H R F S K R R D 299 E I R Q R I L R L 206 I H S L S G M E S 206 I S A I G L D S L 220 Y L L I K T V L 297 T K E I R Q F <td> 1870</td> <td>25 24 23 23 23 21 18 18 17 17 17 17 17 17 17 17 17 17 17</td>	1870	25 24 23 23 23 21 18 18 17 17 17 17 17 17 17 17 17 17 17

SEQ ID NO:	HLA-B*	2705 nonomers	,	SEQ ID NO:	H	LA-B*2	2705 nonomers	
<u>110.</u>				<u></u>	_		700 110110110210	
	<u>Pos</u>	1.23456789	score			Pos	1 2 3 4 5 6 7 8 9	score
<u>1889</u>	<u>20</u> <u>14</u>	FILIGLPGL	<u>16</u>	<u>1939</u>	<u>70</u>	<u>213</u>	S L L I S F S Y L	<u>14</u>
	<u>21</u> <u>21</u>	GLEEAQFWL	<u>16</u>	<u>1940</u>	<u>71</u>	<u>214</u>	L L I S F S Y L L	<u>14</u>
	<u>22</u> <u>25</u>	AQFWLAFPL	<u>16</u>	<u>1941</u>	<u>72</u>	<u>260</u>	SMVHRFSK R	<u>14</u>
	<u>23</u> <u>37</u>	Y L I A V L G N L	<u>16</u>	1942	<u>73</u>	<u>263</u>	H R F S K R R D S	<u>14</u>
	<u>24</u> <u>92</u>	TTIQFDACL	<u>16</u>	<u>1943</u> 1944	$\frac{74}{25}$	<u>267</u>	KRRDSPLPV	<u>14</u>
	25 <u>101</u>	LQIFAIHSL RYVAICHPL	<u>16</u>	1944 1945	<u>75</u>	<u>293</u>	YGVKTKEIR	<u>14</u>
	26 124 27 130	HPLRHATVL	<u>16</u> <u>16</u>	1946	<u>76</u> 77	<u>301</u> 302	R Q R I L R L F H Q R I L R L F H V	<u>14</u> <u>14</u>
	28 <u>141</u>	PRVTKIGVA	16	1947	<u>77</u> 78	<u> 502</u>	NGNESSATY	13
	29 <u>153</u>	R G A A L M A P L	<u>16</u>	1948	7 9	<u>23</u>	EEAQFWLAF	<u>13</u>
	30 181	CLHQDVMKL	<u> 16</u>	1949	80	 28	WLAFPLCSL	<u>13</u>
<u>1900</u>	31 201	G L I V I I S A I	<u>16</u>	<u>1950</u>	81	44	NLTIIYIVR	<u>13</u>
<u>1901</u>	<u>32</u> <u>203</u>	I V I I S A I G L	<u>16</u>	<u> 1951</u>	82	<u>51</u>	VRTEHSLHE	<u>13</u>
<u>1902</u>	<u>33</u> <u>216</u>	ISFSYLLI L	<u>16</u>	<u>1952</u>	<u>83</u>	<u>56</u>	SLHEPMYIF	<u>13</u>
	<u>34</u> <u>222</u>	LILKTVLG L	<u>16</u>	<u>1953</u>	<u>84</u>	<u>60</u>	P M Y I F L C M L	<u>13</u>
	<u>35</u> <u>255</u>	PFIGLSMVH	<u>16</u>	1954	<u>85</u>	<u>72</u>	D I·L I S T S S M	<u>13</u>
	<u>36</u> <u>257</u>	I G L S M V H R F	<u>16</u>	<u>1955</u> 1956	86	<u>74</u>	LISTSSMPK	<u>13</u>
	<u>37</u> <u>275</u>	V I L A N I Y L L I I Y I V R T E H	<u>16</u>	$\frac{1950}{1957}$	<u>87</u>	<u>75</u>	I S T S S M P K M	<u>13</u>
	38 <u>47</u> 39 <u>109</u>	L S G M E S T V L	<u>15</u> <u>15</u>	1958	<u>88</u> 89	<u>98</u> 104	A C L L Q I F A I F A I H S L S G M	<u>13</u> <u>13</u>
	<u>40</u> <u>114</u>	STVLLAMAF	15 15	1959	<u>90</u>	110	SGMESTVLL	<u>13</u>
	41 123	DRYVAICHP	15	1960	91	116	V L L A M A F D R	13
	<u>42</u> <u>145</u>	KIGVAAVVR	<u>15</u>	<u>1961</u>	92	126	V A I C H P L R H	13
	<u>43</u> <u>156</u>	ALMAPLPVF	15	1962	93	149	AAVVRGAAL	<u>13</u>
<u>1913</u>	<u>44</u> <u>168</u>	LPFCRSNI L	<u>15</u>	<u> 1963</u>	94	<u>158</u>	MAPLPVFIK	<u>13</u>
	<u>45</u> <u>172</u>	R S N I L S H S Y	<u>15</u>	<u>1964</u>	<u>95</u>	<u>164</u>	F I K Q L P F C R	<u>13</u>
	<u>46</u> <u>198</u>	<u> </u>	<u>15</u>	<u>1965</u>	<u>96</u> .	<u>170</u>	FCRSNILSH	<u>13</u>
	<u>47</u> <u>206</u>	I S A I G L D S L	<u>15</u>	<u>1966</u>	<u>97</u>	<u>171</u>	CRSNILSH S	<u>13</u>
	48 229	G L T R E A O A K	<u>15</u>	1967	<u>98</u>	<u>187</u>	MKLACDDIR	<u>13</u>
	<u>49</u> <u>248</u>	A V F I F Y V P F I F Y V P F I G L	<u>15</u>	<u>1968</u> 1969	<u>99</u> 100	<u>217</u>	SFSYLLILK	<u>13</u>
	50 <u>251</u> 51 <u>274</u>	PVILANIYL	<u>15</u> 15	1970	101	<u>224</u> 242	LKTVLGLTR CVSHVCAVF	<u>13</u> <u>13</u>
1001	52 290	PIVYGVKTK	<u>15</u>	4004	102	<u>256</u>	FIGLSMVHR	13 13
	53 298	KEIRQRILR	<u>15</u>	1972	103	<u>261</u>	MVHRFSKRR	13
	<u>54</u> <u>19</u>	LPGLEEAQF	14	1973	104	49	YIVRTEHSL	12
<u>1924</u>	<u>55</u> <u>29</u>	LAFPLCSLY	<u>14</u>	<u>1974</u>	105	<u>57</u>	LHEPMYIFL	12
· <u>1925</u>	<u>56</u> <u>30</u>	AFPLCSLYL	14	<u> 1975</u>	106	88	WFNSTTIQF	<u>12</u>
1926	<u>57</u> <u>39</u>	IAVLGNLTI	<u>14</u>	<u>1976</u>	<u> 107</u>	<u>96</u>	FDACLLQIF	<u>12</u>
<u>1927</u>	<u>58</u> <u>40</u>	AVLGNLTII	<u>14</u>	<u>1977</u>	<u> 108</u>	<u>134</u>	HATVLTLPR	<u>12</u>
<u>1928</u> 1929	<u>59</u> <u>79</u>	SMPKMLAIF	<u>14</u>	1978	<u>109</u>	<u>152</u>	VRGAALMAP	<u>12</u>
	60 <u>81</u>	PKMLAIFWF	<u>14</u>	<u>1979</u> <u>1980</u>	110	<u>179</u>	SYCLHODVM	<u>12</u>
	61 <u>99</u> 62 <u>137</u>	CLLQIFAI H VLTLPRVT K	<u>14</u> 14	1981	111	<u>197</u>	N V V Y G L I V I	<u>12</u>
	62 137 63 138	L T L P R V T K I	<u>14</u> <u>14</u>	1982	112 113	<u>244</u> <u>265</u>	S H V C A V F I F F S K R R D S P L	<u>12</u> <u>12</u>
	64 <u>150</u>	AVVRGAALM	<u>14</u>	1983	114	<u>273</u>	LPVILANIY	12
	65 <u>160</u>	PLPVFIKQL	<u>14</u>	1984	115	285	PPVLNPIVY	12
<u>1935</u>	66 174	NILSHSYCL	14	1985	116	288	L N P I V Y G V K	12
<u> 1936</u>	67 <u>180</u>	YCLHQDVMK	<u>14</u>	<u> 1986</u>	117	296	KTKEIRORI	12
<u>1937</u>	<u>192</u>	D D I R V N V V Y	<u>14</u>	<u> 1987</u>	<u>118</u>	297	TKEIRORIL	. 12
<u>1938</u>	<u>69</u> <u>212</u>	DSLLISFSY	<u>14</u>					
	_							

SEO ID NO:	<u>H</u>	LA-B*:	2709 nonomers		SEQ ID NO:	<u>H</u>	LA-B*2	2709 nonomers	
		Pos		score		_	Pos		score
<u>1988</u> <u>1989</u>	<u>1</u> <u>2</u>	<u>194</u> 268	1 2 3 4 5 6 7 8 9 I R V N V V Y G L R R D S P L P V I	24 24	2038 2039	<u>51</u> 52	248 279	1 2 3 4 5 6 7 8 9 A V F I F Y V P F N I Y L L V P P V	12 12
<u>1990</u> 1991	<u>3</u> <u>4</u>	132 267	LRHATVLTL KRRDSPLPV	<u>22</u> <u>21</u>	<u>2040</u> 2041	<u>53</u> <u>54</u>	287 296	<u>V L N P I V Y G V</u> K T K E I R Q R I	12 12
1992	<u>5</u>	300	IRQRILRLF	<u>20</u>	2042	<u>55</u>	299	EIRQRILRL	<u>12</u>
<u>1993</u> 1994	<u>6</u> <u>7</u> <u>8</u>	<u>231</u> 302	TREAQAKAF QRILRLFHV	<u>19</u> <u>19</u>	SEQ				
1995	8	124	RYVAICHPL	<u>16</u>	<u>ID</u> NO:	н	LA-B*5	3101 nonomers	
<u>1996</u> 1997	<u>9</u> <u>10</u>	<u>269</u>	RDSPLPVIL	<u>16</u>		_	Pos		score
1998	11 11	<u>43</u> 216	GNLTIIYIV ISFSYLLIL	<u>15</u> <u>15</u>	2043			1 2 3 4 5 6 7 8 9	
<u>1999</u>	12	11	ATYFILIGL	<u>14</u>	2044	<u>1</u> <u>2</u>	<u>39</u> 31	FPLCSLYLI	<u>26</u> <u>25</u>
<u>2000</u> 2001	<u>13</u> <u>14</u>	<u>25</u> 153	AQFWLAFPL RGAALMAPL	<u>14</u> <u>14</u>	2045	<u>3</u>	120	MAFDRYVAI	<u>24</u>
2002	15	174	NILSHSYCL	<u>14</u>	<u>2046</u> 2047	<u>4</u> <u>5</u>	<u>130</u> 118	H P L R H A T V L L A M A F D R Y V	<u>23</u> 22
<u>2003</u> 2004	<u>16</u>	222	LILKTVLGL	· <u>14</u>	2048	<u>6</u>	140	L P R V T K I G V	· <u>22</u> <u>22</u>
2005	<u>17</u> <u>18</u>	<u>257</u> 280	I G L S M V H R F I Y L L V P P V L	<u>14</u> <u>14</u>	<u>2049</u> 2050	<u>7</u>	<u>155</u>	AALMAPLPV	<u>22</u>
2006	<u>19</u>	<u></u>	GNESSATYF	<u>13</u>	<u>2050</u> 2051	<u>8</u> <u>9</u>	<u>42</u> 254	LGNLTIIYI VPFIGLSMV	<u>21</u> <u>21</u>
<u>2007</u> 2008	<u>20</u> 21	<u>14</u> 21	FILIGLPGL	<u>13</u>	2052	<u>10</u>	284	V P P V L N P I V	<u></u>
2009	<u>21</u> <u>22</u>	<u>66</u>	G L E E A Q F W L C M L S G I D I L	<u>13</u> <u>13</u>	<u>2053</u> 2054	11	<u>168</u>	L P F C R S N I L	<u>20</u>
<u>2010</u>	<u>23</u>	130	HPLRHATVL	<u>13</u>	2055	<u>12</u> <u>13</u>	<u>235</u> <u>138</u>	<u>Q A K A F G T C V</u> L T L P R V T K I	<u>20</u> <u>19</u>
<u>2011</u> 2012	<u>24</u> 25	<u>201</u> 203	<u>G L I V I I S A I</u> I V I I S A I G L	<u>13</u>	2056	<u>14</u>	<u>159</u>	APLPVFIKQ	<u>18</u>
2013	<u>25</u> 26	214	LLISFSYLL	<u>13</u> <u>13</u>	<u>2057</u> 2058	15 16	189	LACDDIRVN	<u>18</u>
2014	<u>27</u>	<u>251</u>	IFYVPFIGL	<u>13</u>	<u>2059</u>	<u>16</u> <u>17</u>	<u>198</u> 277	V V Y G L I V I I L A N I Y L L V P	<u>18</u> <u>18</u>
<u>2015</u> 2016	<u>28</u> 29	<u>263</u> 275	HRFSKRRDS VILANIYLL	<u>13</u>	2060	<u>18</u>	207	SAIGLDSL L	<u>17</u>
2017	<u>30</u>	305	LRLFHVATH	<u>13</u> <u>13</u>	<u>2061</u> 2062	<u>19</u>	<u>283</u>	LVPPVLNPI	<u>17</u>
<u>2018</u>	<u>31</u>	30	AFPLCSLYL	<u>12</u>	2063	<u>20</u> 21	<u>63</u> 86	I F L C M L S G I I F W F N S T T I	<u>16</u> <u>16</u>
<u>2019</u> <u>2020</u>	<u>32</u> 33	<u>34</u> <u>37</u>	<u>C S L Y L I A V L</u> <u>Y L I A V L G N L</u>	<u>12</u>	<u>2064</u>	22	110	SGMESTVLL	16
<u>2021</u>	33 34	<u>51</u>	VRTEHSLHE	<u>12</u> <u>12</u>	<u>2065</u> 2066	23 24	144	TKIGVAAVV	<u>16</u>
<u>2022</u>	<u>35</u>	<u>60</u>	PMYIFLCML	<u>12</u>	2067	<u>24</u> 25	<u>149</u> <u>197</u>	A A V V R G A A L N V V Y G L I V I	<u>16</u> <u>16</u>
2023 2024	<u>36</u> <u>37</u>	<u>75</u> <u>93</u>	ISTSSMPKM TIQFDACLL	<u>12</u> <u>12</u>	2068	<u>26</u>	<u>271</u>	SPLPVILAN	<u>16</u>
2025	38	123	DRYVAICHP	<u>12</u>	<u>2069</u> 2070	<u>27</u> 28	<u>280</u> 3	<u>I Y L L V P P V L</u> D P N G N E S S A	<u>16</u> 15
<u>2026</u> 2027	<u>39</u>	<u>135</u>	ATVLTLPRV	<u>12</u>	2071	29	<u>3</u> 40	AVLGNLTII	<u>15</u> <u>15</u>
2028	<u>40</u> <u>41</u>	<u>138</u> <u>149</u>	<u>L T L P R V T K I</u> A A V V R G A A L	<u>12</u> <u>12</u>	2072	<u>30</u>	<u>97</u>	DACLLQIFA	<u>15</u>
2029	42	155	AALMAPLPV	12	<u>2073</u> 2074	<u>31</u> <u>32</u>	<u>132</u> 222	L R H A T V L T L L I L K T V L G L	<u>15</u> <u>15</u>
<u>2030</u> <u>2031</u>	<u>43</u>	168 101	LPFCRSNIL	12	2075	33	279	NIYLLVPPV	15
<u>2031</u> 2032	<u>44</u> <u>45</u>	<u>181</u> 188	C L H Q D V M K L K L A C D D I R V	<u>12</u> <u>12</u>	<u>2076</u>	<u>34</u>	285	P P V L N P I V Y	<u>15</u>
2033	46	190	ACDDIRVNV	<u>12</u>	<u>2077</u> 2078	<u>35</u> <u>36</u>	<u>289</u> <u>9</u>	N P I V Y G V K T S S A T Y F I L I	<u>15</u> <u>14</u>
<u>2034</u> 2035	<u>47</u>	<u>195</u>	RVNVVYGLI	<u>12</u>	2079	<u>37</u>	<u>≟</u> <u>65</u>	LCMLSGIDI	14 14
<u>2033</u> 2036	<u>48</u> 49	<u>210</u> 213	G L D S L L I S F S L L I S F S Y L	<u>12</u> <u>12</u>	2080	<u>38</u>	84	LAIFWFNST	<u>14</u>
2037	50	220	YLLILKTVL	12	<u>2081</u> 2082	<u>39</u> 40	<u>126</u> <u>157</u>	<u>V A I C H P L R H</u> <u>L M A P L P V F I</u>	<u>14</u> <u>14</u>
rd 627				173		<u></u>	<u> </u>		17

HLA-B*5101 nonomers

	=			
		<u>Pos</u>	1 2 3 4 5 6 7 8 9	score
<u> 2083</u>	<u>41</u>	<u>158</u>	MAPLPVFIK	<u>14</u>
2084	42	191	CDDIRVNVV	14
<u>2085</u>	<u>43</u>	200	YGLIVIISA	<u>14</u>
2086	44	209	IGLDSLLIS	14
<u>2087</u>	<u>45</u>	215	LISFSYLLI	14
<u>2088</u>	46	219	SYLLILKTV	14
2089	<u>47</u>	220	YLLILKTVL	<u>14</u>
2090	<u>48</u>	<u>237</u>	KAFGTCVSH	<u>14</u>
<u> 2091</u>	<u>49</u>	247	CAVFIFYVP	<u>14</u>
<u> 2092</u>	<u>50</u>	249	V F I F Y V P F I	14
<u> 2093</u>	<u>51</u>	<u>251</u>	I F Y V P F I G L	<u>14</u>
<u>2094</u>	<u>52</u>	<u>257</u>	I G L S M V H R F	14
<u> 2095</u>	<u>53</u>	<u> 268</u>	RRDSPLPVI	14
<u> 2096</u>	<u>54</u>	<u>273</u>	<u>LPVILANIY</u>	14
<u> 2097</u>	<u>55</u>	<u>29</u>	LAFPLCSLY	<u>13</u>
<u>2098</u>	<u>56</u>	<u>33</u>	LCSLYLIAV	<u>13</u>
<u> 2099</u>	<u>57</u>	<u>55</u>	H S L H E P M Y I	<u>13</u> <u>13</u>
<u>2100</u>	<u>58</u>	<u>67</u>	MLSGIDILI	<u>13</u>
<u>2101</u>	<u>59</u>	<u>80</u>	MPKMLAIFW	<u>13</u>
<u>2102</u>	<u>60</u>	<u>95</u>	<u>Q F D A C L L Q I</u>	<u>13</u>
<u>2103</u>	<u>61</u>	<u>98</u>	ACLLQIFAI	<u>13</u>
<u>2104</u>	<u>62</u>	<u>104</u>	FAIHSLSGM	<u>13</u>
<u>2105</u>	<u>63</u>	<u>146</u>	IGVAAVVRG	<u>13</u>
<u>2106</u>	<u>64</u>	<u>148</u>	V A A V V R G A A	<u>13</u>
<u>2107</u>	<u>65</u>	<u>153</u>	RGAALMAPL	<u>13</u>
<u>2108</u>	<u>66</u>	<u>233</u>	E A Q A K A F G T	<u>13</u>
<u>2109</u>	<u>67</u>	243	V S H V C A V F I	<u>13</u> <u>13</u>
<u>2110</u>	<u>68</u>	292	V Y G V K T K E I	<u>13</u>
<u>2111</u>	69	296	KTKEIRORI	<u>13</u>

Table				Ħ	LA-A*	0201 decamers	
HLA C1	ass .	I decamers			Pos	1-2-3-4-5-6-7-8-9-0	score
13*	T N _ N 1	. decamers		27	29	<u> </u>	20
-	Pos	1 2 3 4 5 6 7 8 9 0	score	28	142	RVTKIGVAAV	20
1	191	-C D D I R V N V V Y	27	29	148	<u> </u>	20
± 2		SHVCAVFIFY	27 24	30	167	Q L P F C R S N I L	20
3	40	AVLGNLTIIY	21	31	180	Y C L H Q <u>D</u> V M K L	20
4	284	V P P V L N P I V Y	21 21	32	222	- L - I - L - K - T - V - L - C - L - T	20
1 5		<u>V L L A M A F D R Y</u>	21	33	240	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	20
5	28	W L A F P L C S L Y	20 18	34	248	AVFIFYVPFI	20
7	297	T K E I R O R I L R	17	35	250	FIFYV <u>P</u> FI C L	20
<i>∓</i> 8	21	-C L E E A O F W L A	16	36	271	SPLPV <u>I</u> LA N I	20
9	22	LEEAQFWLAF	16	37	279	NIYLL <u>V</u> PPVL	20
10	52	RTEHSLHEPM	16	38	304	ILRLFHVATH	20
11	53	TEHSLHEPMY	16	39	10	SATYF <u>I</u> LICL	19
12	57	LHEPMYIFLC	16	40	15	I L I C L P C L E E	19
13	111	G M E S T V L L A M	16	41	27	FWLAFPLCSL	19
14	272	PLPVILANIY	16	42	35	SLYLI <u>AVLC</u> N	19
15	1	MVDPNCNESS	15	43	37	Y L I A V L G N L T	19
16		P'N G N E S S A T Y	15 15	44	44	N L T I I Y I V R T	19
17		A F D R Y V A I C H	15 15	45	64	FLCMLSCIDI	19
18		CRSNILSHSY	15 15	46	83	MLAIFWFNST	19
19	211	LDSLLISFSY	15 15	47	159	APLPV <u>FIKQ</u> L	19
20	8	ESSATYFILI	13 13	48	189	LACDDIRVNV	19
21	_	A C D D I R V N V V	13 13	49	207	SAICL <u>D</u> SLLI	19
		. <u> </u>	13	50	253	YVPFIGLSMV	19
H	LA-A*	0201 decamers		51	276	ILANIYLL V P	19
7	Pos	1 2 3 4 5 6 7 8 9 0	score	52	281	YLLVPPVLNP	19
1	221	L I I K T V I G I	30	53	283	<u> </u>	-
2	100	LLQIFAIH S L	29	54	286	-P V-L N P I V Y C V	19
3	282	LLVPPVLN P I	27	55	33	-L-C-S-L-Y-L-I-A-V-L	18
4	205	<u>-</u> I	26	56	36	LYLIAVLCNL	18
5	213	-S L L I S F S Y L L	25	57	39	IAVLGNLTI	18
6	56	<u>-</u> 	24	58	42	LCNLTIIYIV	18
7	62	Y I F L C M L S C I	24	59	66	CMLSG <u>I</u> DILI	18
8	108	-S-L S G M E S T-V-L	24	60	111	-C M E S T <u>V</u> L L A M	18
9	117	<u> </u>	24			<u>-I C H P L R H A T V</u>	
10	131	- P L R H A T V L T L	24			<u> </u>	
11	137	<u> </u>	24			CAALMAPLPV	
12	215	<u> </u>	24			LMAPLEVFIK	
13	38	LIAVL <u>G</u> NLTI	23			-ACDDIRVNVV	
14	41	V L G N L T I I Y I	23			GLTREAQAKA	
15	156	-A-L M A P L P V F I	23			HVCAVEIFYV	
16	193	-DIRVNVVYCL	23			-P V I L A N I Y L L	
17	214	<u>-L L I S F S Y L L I</u>	23			ANIYLLV PPV	
18	32	PLCSLYLIAV	22			- I V Y G V <u>K</u> T K E I	
19	119	- A M A F D R Y V A I	22			KEIRQRILRL	
20	237	<u>K A F C T C V S H V</u>	22	72		I Y I V R T E H S L	
21	275	<u> </u>	22			LCMLSCIDIL	
22	85	AIFWFNSTTI	21	74		M L S G I D I L I S	
23	139	-T L P R V T K I G V	21	75		LISTSSMPKM	
24	202	LIVIISAIGL	21	76		STTIQFDACL	
25	13	-Y F I L I <u>C</u> L P C L	20			IQFDAGLLQI	
26	16	-LI-G-LP-G-LEEA	20			K L A C D D I R V N	
				. <u></u>	19/	N V V Y C <u>L</u> I V I I	17

H	ILA-A*	:0201 decamers	HLA-A*0203-decamers					
-	Pos	-1 2 3 4 5 6 7 8 9 0	score	_	Pos	1234567890	score	
80	200	Y C L I V I I S A I	17	30	22	LEBAQFWLAF	9	
81	218	<u>F 8 Y L L I L K T V</u>	17	31	32	P L C S L Y L I A V	9	
82	227	V L C L T R E A Q A	17	32	77		9	
83	303	RILRLFHVAT	17	33	90	NSTTIQFDAC	9	
84	21	G L E E A Q F W L A	16	34	97	DACLLOIFAI	و	
85	92	TTIQFDACL	16	35	111		9	
86	97	DACLLQIFAI	16	36	113	ESTVLLAMAF	9	
87	127	- A I C H P L R H A T	16	37	119	-A M A F D R Y V A I	9	
88	143	<u> </u>	16	38	127	A I C H P L R H A T	9	
89	195	-R V N V V Y G L I V	16	39	151	- V R G A A L M A P	9	
90	220	<u> </u>	16	40	182	_ L H Q D V M K L A C	9	
91	296	-K T K E I R Q R I L	16	41	200	-Y G L - I V I I S A I	9	
92	18	-G L P G L E E A Q F	15	42	226		9	
93	30	AFPLCSLYLI	15	43	228	LCLTREAQAK	9	
94	126	<u> </u>	15	44	230		9	
95	145	<u>K I G V-A A V V R-G</u>	15	45	240	-G T C V S H V C A V	9	
96	173	SNILSHSYCL	15	46	270		9	
97	201	G L I V I I S A I G	15	47	303	RILRLFHVAT	9	
98	208	AIGLD <u>S</u> LLIS	15	48	306		٠ ۽	
99	210	GLDSLLISFS	15			- -		
100	267	KRRDS <u>P</u> LP V I	15					
				_		26 decamero		
H	ILA-A*	0203 decamers			Pos	1 2 3 4 5 6 7 8 9 0	score	
	Pos	1 2 3 4 5 6 7 8 9 0	score	1	299	EIRQRILRLF	31	
1	141	PRVTKIGVAA	19	2	193	DIRVNVVYGL	29	
2	147	-G V A A V V R C A A	19	3	250	-FIFYVPFIGL	25	
3	112	MESTVLLAMA	18	4	256	FIGLSMVHRF	25	
4	227	V L G L T R E A Q A	18	5	74	LISTSSMPKM	24	
5	229	G L T R E A Q A K A	18	6	274	PVILANIYLL	24	
6	142	-R <u>V T K I G V</u> A A V	17	7	18	G L P G L E E A Q F	23	
7	148	V A V V R G A A L	17	8	116	V L L A M A F D R Y	23	
용	2	V D P N C N E S S A	10	9	205	I I S A I C L D S L	23	
9	16	LIGLPGLEEA	10	10	221	L L I L K T V L G L	23	
10	21	G L E E A Q F W L A	10	11	230	LTREAQAKAF	23	
11	31	FPLCSLYLIA	10	12	13	Y F I L I C L P C L	22	
12	76		10	13	40	AVLGNLTIIY	22	
13	89	- -	10	14		S L H E P M Y I F L	22	
14		F D A C L L Q I F A	10	15		Q F D A C L L Q I F	22	
15		SCMESTVLLA	10	16 17		LISFSYLLIL	22	
16		LAMAFDRYVA	10			TTIQFDACLL LLQIFAIHSL	21	
17		V A I C H P L R H A	10	18		I F A I H S L S G M	21	
18		LPRVTKIGVA	10	19			21	
19		I G V A A V V R G A	10	20		KTKEIRQRIL	21	
20	150	A V V R G A A L M A	10	21 22		W L A F P L C S L Y P L R H A T V L T L	20	
21	181	C L H Q D V M K L A	10				20	
22	199	VYCLIVIISA	10	23 24		EPMYIFLOML	19	
23	225	K T V L G L T R E A	10	24 25		STTIQFDACL LIVIISAICL	19	
24	239	FGTCVSHVCA	10			DSLLISFSYL	19	
25	269	RDSPLPVILA	10	26 27		- P L P V - I L A N I Y	19	
26		QRILRLEHVA	10	2 / 28		NIYLLVPPVL	19	
27		LRLFHVATHA	10	28		RTEHSLHEPM	19	
28		DPNCNESSAT	9	29 30		-Y I F L C M L S C I	18 18	
29	+-	I G L P G L E E A Q	9	30	02	1 1 1 1 0 11 1 0 0 1	10	

<u>sd-63712</u>_____176

HLA-A26 decamers			HLA-A		3 decamers		
_	Pos	1 2 3 4 5 6 7 8 9 0	score	_	Pos	1-2-3-4-5-6-7-8-9-0	score
31	72	-DILISTSSM-P	18	16	125	YVAICHPLRH	20
32	108	SLSCMESTVL	18	17	131	-P-L R H A T V-L T-L	20
33	113	-E-S-T-V-L-A-M-A-F	18	18	144	T K I G V A A V V R	20
34	151	-V-V-R-G-A-A-L-M-A-P	18	1.9	156	-A L M A P L P V F I	20
35	78	SSMPKMLAIF	17	20	195	-R -V N V V Y -C L I V	20
36	142	RVTKIGVAAV	17	21	35	SLYLIAVLCN	19
37	162	PVFIKQLPFC	17	22	272	-P L P V I L A N I Y	19
38	164	FIKQLPFCRS	17	23	37	YLIAVLGNLT	18
39	167	Q L P F C R S N I L	17	24	49	YIVRTEHSLH	18
40	185	DVMKLACDDI	17	25	50	<u> </u>	18
41	248	AVFIFYVPFI	17	26	108	<u> </u>	18
42	253	YVPFIGLSMV	17	27	142	<u>-R V <u>T</u> K I <u>G</u> <u>V</u> A A V</u>	18
43	45	-LTIIYIVRTE	16	28	188	KLACDDIRVN	18
4-4	145	-K-I-G-V-A-A-V-V-R-G	16	29	279	<u>NIYLLVPPVL</u>	18
45	198	VVYGLIVIIS	16	30	291	<u>-I V Y C V K T K E I</u>	18
4-6	203	-I-V-I-I-S-A-I-G-L-D	16	31	294	GVKTKEIRQR	18
47	209	-I C L D S L L I S F	16	32	46	T I <u>I</u> Y I <u>V</u> <u>R</u> T E H	17
48	213	-S-L-L-I-S-F-S-Y-L-L	16	33	102	QIFAIHSLSC	17
49	255	PFIGLSMVHR	16	34	151	<u> </u>	17
50	264	RFSKRRDSPL	16	35	179	<u>-S ¥ C L H Q D V M K</u>	17
51	294	CVKTKEIRQR	16	36	203	IV<u>I</u>IS<u>AI</u>GLD	17
52	16	LICLPCLEEA	15	37	204	-VIISAIGLDS	17
53	80	-M-P-K-M-L-A-I-F-W-F	15	38	220	<u> </u>	17
54	114	STVLLAMAFD	15	39	221	LLIKTVLC	17
55	155	AALMAPLPVF	15	40	227	V L C L T R E A Q A	17
56	159	-APLPVFIKQL	15	41	242	<u>C V S H V C A V F I</u>	17
57	174	NILSHSYC-LH	15	42	289	<u>N P I V Y G V K T K</u>	17
58	197	NVYCLIVII	15	43	38	LI<u>V</u>APENTI	16
59	210	G L D S L L I S F S	15	44	85	AIFWFNSTTI	16
60	214	-L I S F S Y L L I	15	45	147	GVANVYRCAA	16
61	222	- L I L K T V L C L T	15	46	198	VVYGLIVIIS	16
62	240	CTCVSHVCA-V	15	47	201	GLIVIISAIG	16
63	247	C A V F I F Y V P F	15	48	214	-LLISFSYLLI	16
64	286	PVLNPIVYGV	15	49	226	TVLGLTREAQ	16
65	298	KEIRQRIL-RL	15	50	228	LCLTREAQAK	16
10	T 7_72	- decamers		51		CLTREAQAKA	16
_	Pos	-1 2 3 4 5 6 7 8 9 0	score	52		MVDPNGNESS	15
		T V L T L P R V T K	31	53		NLTIIYIVRT	15
2		V L N P I V Y C V K	28	54		-IIYIVRTEHS	15
3		<u>-I L K T V L G L T R</u>	27	55	67	-M L S G I D I L I S	15
4		I L R L F H V A T H	27	56		-DILISTSSMP	15
<u>.</u> 5		I L I S T S S M P K	26	57		-C L L Q I F A I H S -A I H S L S G M E S	15
6		I L I G L P G L E E	23	58 59			15
7	40	- A V L G N L T I I Y	23			-K I G V A A V V R G	15
8	150	AVVRGAALMA	23	60 61		<u>-I L S H S Y C L H Q</u> -C D D I R V N V V Y	15
9	258	G L S M V H R F S K	23	62	-	AIGLDS LIS	15 15
10	18	CLPGLEEAQF	22	63		VILANIY LV	. 15
11		RILRLFHVAT	22			Y L L V P P V L N P	15 15
12		I L A N I Y L L V P	21			EIRQRILRIF	1-5
13	28	WLAFPLCSLY	20			RLFHVATHAS	15
14		TVLLAMAFDR	20	99	J00		10
15		<u> </u>	20	H	LA-B	10702 decamers	
				_			

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	Pos	1234567890	score	H	LA-B*	0702	-decamers	
1	159	APLPVFIKQL	23	_	Pos		3 4 5 6 7 8 9 0 see	re
2	59	EPMYIFLCM-L	22	55	267	-K-F	RDSPLPVI	11
3	273	LPVILANI¥L	20	56	269	-R-E	S P L P V I L A	11
4	3	DPNGNESSAT	19	57	278	-A-N	I I Y L L V P P V	11
5	130	HPLRHATVLT	19	58	28 4	- V- E	PVLNPIVY	11
6	140	LPRVTKICVA	19	59	289	-N-E	Y C V K T K	11
7	161	LPVFIKQLPF	19	60	296	-K 1	'KEIRQRIL	11
8	31	FPLCSLYLIA	18	SEQ				
9	271	SPLPVILANI	18	<u>ID</u>				
10	80	-MPKMLAIFWF	16	<u>NO:</u>	_		decamers	
11	108	SLSCMESTVL	16	2112	_	Pos	1 2 3 4 5 6 7 8 9 0	score
12	131	- P - L - R - H - A - T - V - L - T - L	15	$\frac{2112}{2113}$	1	<u>191</u>	C D D I R V N V V Y	<u>27</u>
13	264	RFSKRRDSPL	15	$\frac{2113}{2114}$	<u>2</u>	244	SHVCAVFIFY	<u>24</u>
14	33	-L-C-S-L Y L I A V L	14	$\frac{2114}{2115}$	<u>3</u>	<u>40</u>	AVLGNLTIIY	<u>21</u>
15	109	LS-C-MESTVLL	14	$\frac{2115}{2116}$	<u>4</u>	284 116	V P P V L N P I V Y	<u>21</u>
16	152	VRGAALMAPL	14	<u>2110</u> 2117	<u>2</u>	<u>116</u>	V L L A M A F D R Y	<u>20</u>
17	205	IISAIGLD S L	14	<u>2117</u> 2118	유	<u>28</u> 297	WLAFPLCSLY	<u>18</u>
18	215	LISFSYLLIL	14	2119	<u>-</u>		TKEIRORILR GLEEAOFWLA	<u>17</u>
19	268	RRDSPLPVIL	14	2120	5 <u> </u> 6 <u> </u> 7 <u> </u> 8 <u> </u> 9 <u> </u>	<u>21</u> 22	LEEAOFWLAF	<u>16</u> <u>16</u>
20	29	LAFPLCSLY L	13	2121	≟ 10	<u>52</u>	RTEHSLHEPM	16 16
21	148	VAAVVRGAAL	13	2122	11	<u>52</u>	T E H S L H E P M Y	16 16
22	156	ALMAPLPVFI	13	2123	12	<u>55</u>	L H E P M Y I F L C	<u>16</u>
23	193	DIRVNVYGL	13	2124	13	111	G M E S T V L L A M	<u>16</u>
24	221	LLILKTVLGL	13	2125	14	272	PLPVILANIY	<u>16</u>
25	298	KEIRQRILRL	13	2126	<u>15</u>	1	MVDPNGNESS	15
26	7	NESSATYFIL	12	2127	<u>16</u>	<u> 4</u>	PNGNESSATY	<u>15</u>
27	19	LPCLEEAQFW	12	2128	17	121	AFDRYVAICH	<u>15</u>
28	24	EAQFWLAFPL	12	2129	18	171	CRSNILSHSY	<u>15</u>
29	119	AMAFDRYVAI	12	2130	19	211	LDSLLISFSY	<u>15</u>
30	129	CHPLRHATVL	12	2131	20	8	ESSATYFILI	<u>13</u>
31	206	ISAIGLDSLL	12	2132	21	190	A C D D I R V N V V	13
32	219	SYLLILKTVL	12					
33	279 285	PPVLNPIV Y G	12 12	SEO				
34 35	203	ESSATYFILI	11	<u>ID</u> NO:	12	IT. A A .	0201 decamers	
36	13	YFILICLPG L	11	<u>NO.</u>	=	Pos	1 2 3 4 5 6 7 8 9 0	score
37		FWLAFPLCS L	11	<u>2133</u>	1	221	LLILKTVLGL	30
38	48	IYIVRTEHSL	11	2134	≐ 2	100	L L Q I F A I H S L	<u>29</u>
39	56	SLHEPMYIFL	11	2135	= 3	282	L L V P P V L N P I	<u>27</u>
40	65	LCMLSCIDIL	11	2136	<u>≃</u> 4	205	I I S A I G L D S L	26
41	75	ISTSSMPKML	11	2137	2 3 4 5 6 7 8 9	213	SLLISFSYLL	<u>25</u>
. 42	77	TSSMPKMLAI	11	2138	= 6	56	SLHEPMYIFL	24
43	91	STTIQFDACL	11	<u>2139</u>	7	62	YIFLCMLSGI	<u>24</u>
44	123	DRYVAICHPL	11	<u>2140</u>	8	108	SLSGMESTVL	24
45	142	RVTKICVAAV	11	2141	=	117	LLAMAFDRYV	24
46	180	YCLHQDVMKL	11	<u>2142</u>	10	131	PLRHATVLTL	24
47	190	ACDDIRVNVV	11	<u>2143</u>	11	137	V L T L P R V T K I	24
48	212	DSLLISFSYL	11	<u>2144</u>	12	215	LISFSYLLIL	<u>24</u>
49	234	A Q A K A F C T C V	11	<u>2145</u>	13	<u>38</u>	LIAVLGNLTI	<u>23</u>
50	242	CVSHVCAVFI	11	<u>2146</u>	<u>14</u>	41	V L G N L T I I Y I	<u>23</u>
51	248	<u> </u>	11	<u>2147</u>	<u>15</u>	156	A L M A P L P V F I	<u>23</u>
52	250	FIFYVPFICL	11	<u>2148</u>	<u>16</u>	193	D I R V N V V Y G L	<u>23</u>
53	254	<u> </u>	11	<u>2149</u>	<u>17</u>	<u>214</u>	LLISFSYLLI	<u>23</u>
54	266	SKRRDSPLPV	11	<u>2150</u>	<u>18</u>	<u>32</u>	PLCSLYLIAV	<u>22</u>
sd-6371	12			178				
								٠

SEQ ID				SEQ ID				
<u>NO:</u>	HLA-A		NO:	Ħ				
0151	<u>Pos</u>	1 2 3 4 5 6 7 8 9 0	score	0000		<u>Pos</u>	1 2 3 4 5 6 7 8 9 0	score
<u>2151</u>	<u>19</u> <u>119</u>	AMAFDRYVAI	<u>22</u>	<u>2202</u>	<u>70</u>	<u>291</u>	I v Y G V K T K E I	<u>18</u>
<u>2152</u>	<u>20</u> <u>237</u>	KAFGTCVSHV	<u>22</u>	2203	<u>71</u>	<u>298</u>	KEIRQRILRL	<u>18</u>
<u>2153</u>	<u>21</u> <u>275</u>	V I L A N I Y L L V	<u>22</u>	<u>2204</u>	<u>72</u>	<u>48</u>	IYIVRTEHSL	<u>17</u>
2154	<u>22</u> <u>85</u>	A I F W F N S T T I	<u>21</u>	2205	<u>73</u>	<u>65</u>	LCMLSGID I L	<u>17</u>
<u>2155</u>	<u>23</u> <u>139</u>	T L P R V T K I G V	<u>21</u>	<u>2206</u>	<u>74</u>	<u>67</u>	MLSGIDILIS	<u>17</u>
<u>2156</u>	<u>24</u> <u>202</u>	LIVIISAIGL	<u>21</u>	2207	<u>75</u>	<u>74</u>	LISTSSMPKM	<u>17</u>
<u>2157</u> 2158	<u>25</u> <u>13</u>	Y F I L I G L P G L	<u>20</u>	2208	<u>76</u>	<u>91</u>	STTIQFDACL	<u>17</u>
$\frac{2156}{2159}$	<u>26</u> <u>16</u> <u>27</u> <u>29</u>	LIGLPGLEEA	<u>20</u>	<u>2209</u> 2210	<u>77</u>	94	IQFDACLLQI	<u>17</u>
$\frac{2155}{2160}$	<u>27</u> <u>29</u>	LAFPLCSLYL	. 20	<u>2210</u> 2211	<u>78</u>	<u>188</u>	K L A C D D I R V N	<u>17</u>
$\frac{2160}{2161}$	<u>28</u> <u>142</u>	RVTKIGVAAV	<u>20</u>	$\frac{2211}{2212}$	<u>79</u>	<u>197</u>	N V V Y G L I V I I	<u>17</u>
$\frac{2101}{2162}$	<u>29</u> <u>148</u>	V A A V V R G A A L	<u>20</u>	2213	<u>80</u>	<u>200</u>	YGLIVIISAI	<u>17</u>
$\frac{2162}{2163}$	<u>30</u> <u>167</u>	Q L P F C R S N I L Y C L H Q D V M K L	<u>20</u>	2214	<u>81</u>	<u>218</u>	FSYLLILKTV	<u>17</u>
2164	$\frac{31}{32}$ $\frac{180}{222}$	LILKTVLGLT	<u>20</u> 20	2215	<u>82</u>	<u>227</u> 303	V L G L T R E A Q A	<u>17</u>
2165	<u>32</u> <u>222</u> <u>33</u> <u>240</u>	G T C V S H V C A V	<u>20</u> 20	2216	<u>83</u> <u>84</u>	<u>303</u> 21	RILRLFHVAT	<u>17</u>
2166	$\frac{33}{34} \frac{248}{248}$	AVFIFYVPFI	<u>20</u> 20	2217	<u>85</u>	<u>≥±</u> 92	G L E E A Q F W L A T T I Q F D A C L L	<u>16</u> <u>16</u>
2167	35 <u>250</u>	FIFYVPFI·GL	<u>20</u>	2218	<u>86</u>	<u>97</u>	DACLLQIFAI	16
2168	36 <u>271</u>	SPLPVILANI	20	2219	<u>87</u>	<u>127</u>	AICHPLRHAT	<u>16</u>
2169	37 279	NIYLLVPPVL	20	2220	88	143	V T K I G V A A V V	<u>16</u>
2170	38 304	ILRLFHVATH	20	2221	89	195	R V N V V Y G L I V	<u>16</u>
2171	<u>39</u> <u>10</u>	SATYFILIGL	19	2222	90	220	YLLILKTVLG	<u>16</u>
2172	<u>40</u> <u>15</u>	I L I G L P G L E E	19	2223	91	296	KTKEIRORIL	<u>16</u>
2173	41 27	F W L A F P L C S L	19	2224	92	<u>18</u>	G L P G L E E A Q F	<u>15</u>
<u>2174</u>	42 35	SLYLIAVLGN	19	2225	93	30	AFPLCSLY L I	<u>15</u>
<u>2175</u>	43 37	YLIAVLGNLT	19	2226	94	126	V A I C H P L R H A	<u>15</u>
<u>2176</u>	<u>44</u> <u>44</u>	NLTIIYIVRT	<u>19</u>	<u>2227</u>	95	145	KIGVAAVVRG	<u>15</u>
<u>2177</u>	<u>45</u> <u>64</u>	F L C M L S G I D I	<u>19</u>	<u>2228</u>	96	173	SNILSHSYCL	<u>15</u>
<u>2178</u>	46 83	MLAIFWFNST	<u>19</u>	<u>2229</u>	97	201	G L I V I I S A I G	<u>15</u>
<u>2179</u>	<u>47</u> <u>159</u>	APLPVFIKQL	19	<u>2230</u>	98	208	AIGLDSLLIS	<u>15</u>
<u>2180</u>	<u>48</u> <u>189</u>	LACDDIRVNV	<u>19</u>	<u>2231</u>	<u>99</u>	<u>210</u>	G L D S L L I S F S	<u>15</u>
<u>2181</u>	<u>49</u> <u>207</u>	SAIGLDSLLI	<u>19</u>	<u>2232</u>	<u>100</u>	<u> 267</u>	KRRDSPLPVI	<u>15</u>
2182	<u>50</u> <u>253</u>	Y V P F I G L S M V	<u>19</u>	aro.				
<u>2183</u>	<u>51</u> <u>276</u>	I L A N I Y L L V P	<u>19</u>	SEQ ID				
2184	<u>52 281</u>	YLLVPPVLNP	<u>19</u>	NO:	н	LA-A*	0203 decamers	
<u>2185</u>	<u>53</u> <u>283</u>	LVPPVLNPIV	<u>19</u>		_	Pos	1 2 3 4 5 6 7 8 9 0	score
<u>2186</u>	<u>54</u> <u>286</u>	PVLNPIVYGV	<u>19</u>	2233	<u>1</u>	141	PR V TKIGV A A	<u>19</u>
<u>2187</u>	<u>55</u> <u>33</u>	LCSLYLIAVL	<u>18</u>	<u>2234</u>	<u></u>	147	G V A A V V R G A A	<u>19</u>
<u>2188</u> 2189	<u>56</u> <u>36</u>	LYLIAVLGNL	<u>18</u>	<u>2235</u>	<u>3</u>	112	M E S T V L L A M A	<u>18</u>
$\frac{2109}{2190}$	<u>57</u> <u>39</u>	I A V L G N L T I I	<u>18</u>	<u>2236</u>	<u>4</u>	227	V L G L T R E A Q A	<u>18</u>
$\frac{2190}{2191}$	<u>58</u> <u>42</u>	LGNLTIIYIV	<u>18</u>	<u>2237</u>	<u>5</u>	229	G L T R E A Q A K A	<u>18</u>
$\frac{2191}{2192}$	<u>59</u> <u>66</u>	CM ESTVII	<u>18</u>	<u>2238</u>	<u>6</u>	<u>142</u>	RVTKIGVAAV	<u>17</u>
2193	60 <u>111</u>	G M E S T V L L A M	<u>18</u>	<u>2239</u>	5 <u>1</u> 6 <u>1</u> 7 <u>1</u> 8 <u>1</u> 9 <u>1</u>	<u>148</u>	V A A V V R G A A L	<u>17</u>
2194	<u>61</u> <u>128</u>	I C H P L R H A T V	<u>18</u>	<u>2240</u>	<u>8</u>	<u>2</u>	V D P N G N E S S A	<u>10</u>
2195	<u>62</u> <u>134</u>	HATVLTLPRV	<u>18</u>	<u>2241</u>	<u>9</u>	<u>16</u>	LI G LPGLE E A	<u>10</u>
2196	63 <u>154</u> 64 <u>157</u>	G A A L M A P L P V L M A P L P V F I K	<u>18</u>	2242	<u>10</u>	<u>21</u>	GLEEAOFWLA	<u>10</u>
2197		A C D D I R V N V V	<u>18</u>	2243	<u>11</u>	<u>31</u>	F P L C S L Y L I A	<u>10</u>
2198	65 190 66 229	G L T R E A Q A K A	<u>18</u> 18	2244	<u>12</u> <u>13</u>	<u>76</u>	<u>S T S S M P K M L A</u>	<u>10</u>
2199	67 <u>245</u>	HVCAVFIFYV	<u>18</u> 18	2245		<u>89</u>	F N S T T I Q F D A	<u>10</u>
2200	68 <u>274</u>	PVILANIYLL	<u>18</u> <u>18</u>	2246	14	<u>96</u>	FDACLLQIFA	<u>10</u>
2201	69 <u>274</u>	ANIYLLVPPV	18 18	2247	<u>15</u>	110	SGMESTVLLA	<u>10</u>
				2248	<u>16</u>	<u>118</u>	LAMAFDRYVA	<u>10</u>
<u>sd-6371</u>	2		179					

SEQ				<u>SEQ</u>			
<u>ID</u> NO:	HLA-A	0203 decamers		ID NO:	HLA-A2	6 decamers	
	Pos	1 2 3 4 5 6 7 8 9 0	score		Pos	1234567890	score
<u>2249</u>	<u>17 126</u>	VAICHPLR H A	<u>10</u>	<u>2295</u>	<u>15</u> 95	QFDACLLQIF	22
<u>2250</u>	<u>18 140</u>	L P R V T K I G V A	10	<u>2296</u>	<u>16 215</u>	LISFSYLLIL	<u>22</u>
<u>2251</u>	<u>19 146</u>	IG V AAVVR G A	<u>10</u>	<u>2297</u>	<u>17</u> <u>92</u>	TTIQFDACLL	<u>21</u>
<u>2252</u>	<u>20</u> <u>150</u>	AVVRGAAL M A	<u>10</u>	<u>2298</u>	<u>18 100</u>	LLQIFAIHSL	<u>21</u>
<u>2253</u>	<u>21</u> <u>181</u>	C L H Q D V M K L A	<u>10</u>	<u>2299</u>	<u>19 103</u>	<u>IFAIHSLSGM</u>	<u>21</u>
<u>2254</u>	<u>22</u> <u>199</u>	VYGLIVIISA	<u>10</u>	<u>2300</u>	<u>20</u> <u>296</u>	KTKEIRQRIL	<u>21</u>
<u>2255</u>	<u>23</u> <u>225</u>	KTVLGLTREA	<u>10</u>	<u>2301</u>	<u>21</u> <u>28</u>	WLAFPLCSLY	<u>20</u>
<u>2256</u>	<u>24</u> <u>239</u>	F G T C V S H V C A	<u>10</u>	2302	<u>22</u> <u>131</u>	PLRHATVLTL	<u>20</u>
<u>2257</u> 2258	<u>25</u> <u>269</u>	RDSPLPVILA	<u>10</u>	<u>2303</u> 2304	<u>23</u> <u>59</u>	E P M Y I F L C M L	19 19 19 19 19
<u>2258</u> 2259	<u>26</u> <u>302</u>	ORILRLFHVA	<u>10</u>	2304 2305	<u>24</u> <u>91</u>	STTIQFDACL	<u>19</u>
2260	27 <u>305</u> 28 <u>3</u>	L R L F H V A T H A D P N G N E S S A T	10	2306	<u>25</u> <u>202</u> <u>26</u> <u>212</u>	L I V I I S A I G L D S L L I S F S Y L	19
2261	29 <u>17</u>	IGLPGLEEAQ	<u>≥</u> a	2307	<u>26</u> <u>212</u> <u>272</u>	P L P V I L A N I Y	<u>≟2</u> 10
2262	30 22	LEEAQFWLAF	<u>≅</u> 9	2308	28 <u>279</u>	NIYLLVPPVL	<u>13</u>
2263	31 32	P· L C S L Y L I A V	≟ 9	2309	<u>29</u> <u>52</u>	RTEHSLHEPM	18
2264	32 77	T S S M P K M L A I	<u>=</u> 9	2310	30 62	YIFLCMLSGI	<u>18</u>
2265	33 90	N S T T I Q F D A C	<u>=</u> 9	2311	<u>31</u> <u>72</u>	DILISTSSMP	<u>18</u>
<u> 2266</u>	<u>34</u> <u>97</u>	DACLLQIFAI	<u>9</u>	<u>2312</u>	<u>32</u> <u>108</u>	SLSGMESTVL	18
<u>2267</u>	<u>35</u> <u>111</u>	G M E S T V L L A M	<u>9</u>	<u>2313</u>	<u>33</u> <u>113</u>	ESTVLLAMAF	18
<u>2268</u>	<u>36</u> <u>113</u>	ESTVLLAMAF	<u>9</u>	<u>2314</u>	<u>34 151</u>	VVRGAALMAP	<u>18</u> <u>17</u>
<u>2269</u>	<u>37</u> <u>119</u>	AMAFDRYVAI	<u>9</u>	<u>2315</u>	<u>35</u> <u>78</u>	SSMPKMLAIF	<u>17</u>
<u>2270</u>	<u>38</u> <u>127</u>	AICHPLRHAT	<u>9</u>	<u>2316</u>	<u>36</u> <u>142</u>	RVTKIGVAAV	<u>17</u>
<u>2271</u>	<u>39</u> <u>151</u>	V V R G A A L M A P	<u>9</u>	2317	<u>37</u> <u>162</u>	PVFIKQLPFC	<u>17</u>
<u>2272</u> 2273	<u>40</u> <u>182</u>	L H Q D V M K L A C	ର୍ଜା ପା ରା ବା	2318	<u>38</u> <u>164</u>	FIKQLPFCRS	17 17 17 17 17 17 17
<u>2273</u> 2274	<u>41</u> <u>200</u>	YGLIVIISAI	<u>9</u>	<u>2319</u> 2320	<u>39</u> <u>167</u>	<u>QLPFCRSNIL</u>	<u>17</u>
<u>2274</u> 2275	<u>42</u> <u>226</u> <u>43</u> <u>228</u>	T V L G L T R E A Q L G L T R E A Q A K	볼	<u>2320</u> 2321	<u>40</u> <u>185</u>	DVMKLACDDI	<u>1 /</u>
2276	<u>43</u> <u>228</u> <u>44</u> <u>230</u>	LTREAQAKAF	<u>⊒</u> 0	2322	<u>41</u> <u>248</u> <u>42</u> <u>253</u>	A V F I F Y V P F I Y V P F I G L S M V	17
· <u>2277</u>	<u>45</u> <u>240</u>	G T C V S H V C A V	2 9	2323	43 45	LTIIYIVRTE	
2278	46 270	D S P L P V I L A N	<u>≅</u> 9	2324	<u>44</u> <u>145</u>	KIGVAAVVRG	16
2279	<u>47</u> <u>303</u>	RILRLFHVAT	9	2325	45 198	VVYGLIVIIS	16 16 16 16
2280	48 306	RLFHVATHAS	<u>=</u> 9	2326	46 203	IVIISAIGLD	16
			-	<u>2327</u>	47 209	IGLDSLLISF	<u>16</u>
SEO.	٠			<u>2328</u>	48 213	SLLISFSYLL	<u>16</u>
<u>SEQ</u> ID				<u>2329</u>	<u>49 255</u>	PFIGLSMVHR	<u>16</u>
NO:	HLA-A2	?6 decamers		<u>2330</u>	<u>50</u> <u>264</u>	RFSKRRDSPL	<u>16</u>
	Pos	1 2 3 4 5 6 7 8 9 0	score	<u>2331</u>	<u>51</u> <u>294</u>	GVKTKEIRQR	<u>16</u>
<u>2281</u>	<u>1</u> 299	EIRQRILRLF	<u>31</u>	2332	<u>52</u> <u>16</u>	LIGLPGLEEA	<u>15</u>
<u>2282</u>	<u>2</u> <u>193</u>	DIRVNVVYGL	<u>29</u>	<u>2333</u> 2334	<u>53</u> <u>80</u>	MPKMLAIFWF	<u>15</u>
<u>2283</u>	<u>3</u> <u>250</u>	FIFYVPFIGL	<u>25</u>	2335	<u>54</u> <u>114</u> 55	STVLLAMAFD	<u>15</u>
<u>2284</u> 2285	<u>4</u> <u>256</u>	FIGLSMVHRF	<u>25</u>	2336	55 155 56 159	A A L M A P L P V F A P L P V F I K Q L	15 15 15
<u>2285</u> 2286	<u>5</u> <u>74</u>	LISTSSMPKM	<u>24</u>	2337	57 <u>174</u>	NILSHSYCLH	<u></u>
2287	4 256 5 74 6 274 7 18 8 116 9 205	PVILANIYLL	<u>24</u>	2338	58 197	NVVYGLIVII	15
2288	7 <u>18</u> 8 <u>116</u>	G L P G L E E A Q F V L L A M A F D R Y	<u>23</u>	2339	59 210	GLDSLLISFS	15
2289	9 205	IISAIGLDSL	<u>23</u> <u>23</u>	2340	<u>60</u> <u>214</u>	LLISFSYLLI	15
2290	10 221	LLILKTVLGL	23	<u>2341</u>	61 222	LILKTVLGLT	15 15 15
2291	11 230	LTREAQAKAF	23	<u>2342</u>	<u>62 240</u>	GTCVSHVCAV	<u>15</u>
<u>2292</u>	<u>12</u> <u>13</u>	YFILIGLPGL	22	2343	<u>63</u> <u>247</u>	CAVFIFYVPF	<u>15</u>
<u>2293</u>	<u>13</u> <u>40</u>	AVLGNLTIIY	22	2344	<u>64</u> <u>286</u>	PVLNPIVYGV	<u>15</u>
<u>2294</u>	<u>14</u> <u>56</u>	SLHEPMYIFL	<u>22</u>	<u>2345</u>	<u>65</u> <u>298</u>	KEIRQRILRL	<u>15</u>
<u>sd-6371</u>	2		180				
<u> </u>	=		100				
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<u>SEQ</u>				SEO ID			
ID				NO:	HLA-A	decamers	
NO:	HLA-A	3 decamers			Pos	1 2 3 4 5 6 7 8 9 0	score
	<u>Pos</u>	1 2 3 4 5 6 7 8 9 0	score	<u>2396</u>	<u>51 229</u>	G L T R E A Q A K A	<u>16</u>
<u>2346</u>	<u>1</u> <u>136</u>	TVLTLPRVTK	<u>31</u>	2397	<u>52</u> <u>1</u>	M V D P N G N E S S	<u>15</u>
<u>2347</u>	<u>2</u> <u>287</u>	V L N P I V Y G V K	28	2398	<u>53</u> <u>44</u>	NLTIIYIVRT	<u>15</u>
<u>2348</u>	1 136 2 287 3 223 4 304 5 73 6 15 7 40 8 150 9 258	I L K T V L G L T R	<u>27</u>	2399	<u>54</u> <u>47</u>	I I Y I V R T E H S	<u>15</u>
<u>2349</u>	4 304	I L R L F H V A T H	<u>27</u>	2400	<u>55</u> <u>67</u>	MLSGIDILIS	<u>15</u>
<u>2350</u>	<u>5</u> <u>73</u>	I L I S T S S M P K	26	2401	56 72	D I L I S T S S M P	<u>15</u>
<u>2351</u>	6 15	I L I G L P G L E E	23	2402	57 99	C L L Q I F A I H S	<u>15</u>
<u>2352</u>	7 40	AVLGNLTIIY	<u>23</u>	2403	58 <u>105</u>	AIHSLSGMES	<u>15</u>
<u>2353</u>	<u>8 150</u>	AVVRGAALMA	<u>23</u>	2404	<u>59</u> <u>145</u>	KIGVAAVVRG	<u>15</u>
<u>2354</u>	9 258	G L S M V H R F S K	<u>23</u>	2405	60 <u>175</u>	I L S H S Y C L H Q	<u>15</u>
<u>2355</u>	<u>10</u> <u>18</u>	G L P G L E E A Q F	23 22 22 21	2406	61 <u>191</u>	C D D I R V N V V Y	<u>15</u>
<u>2356</u>		RILRLFHVAT	<u>22</u>	2407	62 208	AIGLDSLLIS	<u>15</u>
<u>2357</u>	<u>12 276</u>	I L A N I Y L L V P	<u>21</u>	2408	63 275	V I L A N I Y L L V	<u>15</u>
<u>2358</u>	<u>13</u> <u>28</u>	WLAFPLCSLY	<u>20</u>	<u>2409</u>	64 281	YLLVPPVLNP	<u>15</u>
<u>2359</u>	<u>14 115</u>	TVLLAMAFDR	<u>20</u>	<u>2410</u>	65 299	EIRQRILRLF	<u>15</u>
<u>2360</u>	11 303 12 276 13 28 14 115 15 116	<u> </u>	20 20 20	<u>2411</u>	66 306	RLFHVATHAS	<u>15</u>
<u>2361</u>	<u>16 125</u>		<u>20</u>			•	
<u>2362</u>	16 125 17 131	<u> </u>	<u>20</u>	SEQ			
<u>2363</u>	<u>18</u> <u>144</u>	TKIGVAAVVR	<u>20</u>	<u>ID</u> NO:	UT.A_D	0702 decamers	
<u>2364</u>	<u>19</u> <u>156</u>	ALMAPLPVFI	20 20 20 20 19 18 18 18 18 18 18 18 18 18	NO:	Pos	1 2 3 4 5 6 7 8 9 0	score
<u>2365</u>	<u>20 195</u>	RVNVVYGLIV	<u>20</u>	2412	 .	A P L P V F I K Q L	
<u>2366</u>	<u>21</u> <u>35</u>	S L Y L I A V L G N	<u>19</u>	2413		E P M Y I F L C M L	<u>23</u>
<u>2367</u>	<u>22</u> <u>272</u>	PLPVILANIY	<u>19</u>	2414	3 273	L P V I L A N I Y L	<u>≗≗</u> 20
<u>2368</u>	<u>23</u> <u>37</u>	YLIAVLGNLT	<u>18</u>	2415	<u>2</u> <u>2/3</u>	D P N G N E S S A T	<u> 20</u> 19
<u>2369</u>	20 195 21 35 22 272 23 37 24 49 25 50 26 108 27 142 28 188	YIVRTEHSLH	<u>18</u>	2416	2 59 3 273 4 3 5 130 6 140 7 161 8 31 9 271	HPLRHATVLT	22 20 19 19 19 19 18 16 16
<u>2370</u>	<u>25</u> <u>50</u>	I V R T E H S L H E	<u>18</u>	2417	$\frac{3}{6} \frac{130}{140}$	LPRVTKIGVA	19
<u>2371</u>	<u>26</u> <u>108</u>	SLSGMESTVL	<u>18</u>	2418	7 161	LPVFIKQLPF	19
2372	<u>27</u> <u>142</u>	RVTKIGVAAV	<u>18</u>	2419	8 31	F P L C S L Y L I A	18
2373	<u>28 188</u>	KLACDDIRVN	<u>18</u>	2420	9 271	SPLPVILANI	18
2374	<u>29</u> <u>279</u>	NIYLLVPPVL	<u>18</u>	2421	10 80	MPKMLAIFWF	16
<u>2375</u>	30 <u>291</u> 31 <u>294</u>	I V Y G V K T K E I	<u>18</u>	2422	<u>11</u> 108	SLSGMESTVL	16
<u>2376</u> 2377	<u>31</u> <u>294</u>	GVKTKEIRQR	<u>18</u>	2423	<u>12</u> <u>131</u>	PLRHATVLTL	15
<u>2377</u> <u>2378</u>	<u>32</u> <u>46</u>	T I I Y I V R T E H	17	2424	13 264	R F S K R R D S P L	15
<u>2378</u> 2379	<u>33</u> <u>102</u>	OIFAIHSLS G	<u>17</u>	2425	14 33	LCSLYLIAVL	14
2380	<u>34</u> <u>151</u>	V V R G A A L M A P	<u>17</u>	<u>2426</u>	<u>15</u> 109	L S G M E S T V L L	14
<u>2381</u>	<u>35</u> <u>179</u>	SYCLHQDVMK	<u>17</u>	2427	<u>16</u> <u>152</u>	VRGAALMAPL	14
2382	<u>36</u> <u>203</u>	IVIISAIGLD	<u>17</u>	<u>2428</u>	<u>17 205</u>	I I S A I G L D S L	14
2383	37 <u>204</u> 38 <u>220</u>	V I I S A I G L D S Y L L I L K T V L G	<u>17</u>	<u>2429</u>	<u>18</u> <u>215</u>	LISFSYLLIL	14
2384	39 <u>221</u>	L L I L K T V L G L	<u>17</u>	<u>2430</u>	<u>19 268</u>	R R D S P L P V I L	14
2385	40 227	V L G L T R E A Q A	<u>17</u> <u>17</u>	<u>2431</u>	<u>20 29</u>	LAFPLCSLYL	<u>13</u>
2386	$\begin{array}{c c} 40 & 227 \\ 41 & 242 \end{array}$	C V S H V C A V F I	<u>17</u>	<u>2432</u>	<u>21 148</u>	V A A V V R G A A L	<u>13</u>
2387	42 289	N P I V Y G V K T K	<u>17</u>	<u>2433</u>	<u>22 156</u>	<u> </u>	
2388	<u>42</u> <u>209</u> <u>43</u> <u>38</u>	LIAVLGNLTI	16 16	<u>2434</u>	<u>23</u> <u>193</u>	DIRVNVVYGL	<u>13</u>
2389	44 <u>85</u>	AIFWFNSTTI	16	<u>2435</u>	<u>24</u> <u>221</u>	LLILKTVLGL	<u>13</u>
2390	45 147	G V A A V V R G A A	16 16	<u>2436</u>	<u>25 298</u>	KEIRQRILRL	<u>13</u>
2391	46 198	V V Y G L I V I I S	16 16	2437	<u>26</u> <u>7</u>	NESSATYFIL	<u>12</u>
2392	47 <u>201</u>	G L I V I I S A I G	<u>16</u>	2438	<u>27</u> <u>19</u>	LPGLEEAQFW	<u>12</u>
2393	48 <u>214</u>	L L I S F S Y L L I	16	2439	<u>28</u> <u>24</u>	EAQFWLAFPL	<u>12</u>
2394	49 226	TVLGLTREAQ	16	2440	<u>29 119</u>	AMAFDRYVAI	<u>12</u>
2395	50 228	LGLTREAQAK	16	2441	<u>30 129</u>	CHPLRHATVL	13 13 13 13 12 12 12 12 12 12
		· · · · · · · · · · · · · · · · · · ·		<u>2442</u>	<u>31</u> <u>206</u>	ISAIGLDSLL	<u>12</u>
	_		404				

sd-63712

	SEO				<u>SEO</u>			
	ID NO.	117 B D4	0702 da assesse		ID	177 B D4	0700 3	
	<u>NO:</u>		0702 decamers		NO:		0702 decamers	
		Pos	1234567890	score		Pos	<u>1234567890</u>	score
	<u>2443</u>	<u>32 219</u>	SYLLILKT V L	<u>12</u>	<u>2460</u>	<u>49 234</u>	AQAKAFGT CV	<u>11</u>
	<u>2444</u>	<u>33 279</u>	NIYLLVPP V L	<u>12</u>	<u>2461</u>	<u>50 242</u>	CVSHVCAVFI	<u>11</u>
	<u>2445</u>	34 285	PPVLNPIV Y G	<u>12</u>	<u>2462</u>	<u>51</u> <u>248</u>	AVFIFYVPFI	11
	2446	<u>35</u> <u>8</u>	ESSATYFI L I	<u>11</u>	<u>2463</u>	<u>52</u> <u>250</u>	F I F Y V P F I G L	<u>11</u>
	2447	<u>36</u> <u>13</u>	Y F I L I G L P G L	<u>11</u>	<u>2464</u>	53 254	V P F I G L S M V H	<u>11</u>
	<u>2448</u>	<u>37</u> <u>27</u>	F W L A F P L C S L	11	<u>2465</u>	<u>54</u> <u>266</u>	SKRRDSPLPV	<u>11</u>
	<u>2449</u>	<u>38</u> <u>48</u>	I Y I V R T E H S L	11	<u>2466</u>	<u>55</u> <u>267</u>	KRRDSPLPVI	<u>11</u>
•	<u>2450</u>	<u>39</u> <u>56</u>	SLHEPMYIFL	11	<u>2467</u>	<u>56</u> <u>269</u>	RDSPLPVILA	11
	<u>2451</u>	40 65	LCMLSGIDIL	<u>11</u>	<u>2468</u>	<u>57</u> <u>278</u>	ANIYLLVPPV	<u>11</u>
	<u>2452</u>	<u>41</u> <u>75</u>	I S T S S M P K M L	<u>11</u>	<u>2469</u>	<u>58</u> <u>284</u>	V P P V L N P I V Y	<u>11</u>
	<u>2453</u>	<u>42</u> <u>77</u>	T S S M P K M L A I	<u>11</u>	<u>2470</u>	<u>59</u> <u>289</u>	NPIVYGVKTK	<u>11</u>
	<u> 2454</u>	<u>43</u> <u>91</u>	STTIQFDACL	<u>11</u>	2471	60 296	KTKEIRQRIL	11
	<u>2455</u>	44 123	DRYVAICH P L	<u>11</u>				
	<u>2456</u>	<u>45</u> <u>142</u>	RVTKIGVAAV	<u>11</u>				
	<u>2457</u>	<u>46 180</u>	YCLHQDVMKL	<u>11</u>				
	<u>2458</u>	<u>47</u> <u>190</u>	A C D D I R V N V V	11				
	<u>2459</u>	48 212	D S L L I S F S Y L	11				
5				=				

<u>HLA-DRB1*0101 15-mers</u>

	Pos	-1 2 3 4 5 6 7 8 9 0 1 2 3 4 5	score
1	200	Y G L I V I I S A I G L D S L	36
2		-LSCIDILISTSSMPK	34
3		Y I F L C M L S C I D I L I S	33
4		I F A I H S L S C M E S T V L	32
1 5		LTITYIVRTEHSLHE	32 31
5			
♦ 7		DIRVNVYCLIVIIS	31
•		LANIYLLVPPVLNPI	31
8		DACLUQIFAIHSUSC	30
9		I H S L S G M E S T V L L A M	30
10		CTCVSHVCAVFIFYV	30
		S A T Y F I L I C L P C L E E	29
		N P I V Y G V K T K E I R Q R	29
		ATYFILIGLEGA	28
		-FIFYVPFICLSMVHR	27
15		L P R V T K I G V A A V V R G	26
16		HQDVMKLACDDIRVN	26
17		SFSYLLILKTVLCLT	26
		LIGLPGLEEAQFWLA	25
		EAQFWLAFPLCSLYL	25
20	36	LYLIAVLGNLTIIYI	25
21	70	GIDILISTSSMPKML	25
22	111	C M E S T V L L A M A F D R Y	25
23	148	<u>V A A V V R G A A L M A P L P</u>	25
24	162	-PVFIKQLPFCRSNIL	25
25	197	-N V V Y G L I V I I S A I G L	25
26	211	LDSLLISFSYLLILK	25
27	218	FSYLLILKTVLCLTR	25
58	13	Y F I L I C L P C L E E A Q F	24
29	30	AFPLCSLYLIAVLGN	24
30	39	-I-A V L C N L T I I Y I V R T	24
31	77	TSSMPKMLAIFWFNS	24
32	85	A I F W F N S T T I Q F D A C	24
33	$\frac{137}{1}$	V L T L P R V T K I C V A A V	24
34	$\frac{151}{1}$	V V R G A A L M A P L P V F I	24
35	$\frac{161}{1}$	LPVFIKQLPFCRSNI	24
36	$\frac{196}{1}$	VNVYCLIVIISAIC	24
37	202	LIVIISAICLDSLLI	24
38	208	A	24
39	248	AVFIFYVPFIGLSMV	24
40	$\frac{251}{2}$	IFYVPFICLSMVHRF	24
41	83	MLAIFWFNSTTIQFD	23
42	101	LQIFAIHSLSCMEST	23
43	165	IKQLPFCRSNILSHS	23
44	203	IVIISAICLDSLLIS	23
45	221	L I I L K T V L C L T R E A Q	23
46	278	A N I Y L L V P P V L N P I V	23
47	27	FWLAFPLC-SLYLIAV	
48	35	S L Y L I A V L C N L T I I Y	
49	61	MYIFLCMLSCIDILI	22

HLA-DRB1*0101 15-mers

	Pos	1	2	3	_1	5	-6-	7	Q	۵	Δ.	1	2	3	1	5	score
50	65	<u></u>		-		_	-										22
50		-M															22
~ -	145																22
	146																22
	154																22
	205																22
	243																22
	270																22
	274																22
	281																22
	34																22 21
	69																21
	152																21 21
	299																21 21
	100				_												21 20
	135																20
	141																20
	191																20
-	199											_		_		_	20
	262												-			_	20
	271																20
	271 28																19
	20 58																19
	59																19
	55																
74 75																	19
	215	- A				_											19
70 77																	19
	219																19
70 79	228																19
	232 246																19 19
	297													-	_	_	
	3																19
83																	18
	25	-F												_			18
	23 42																18
	46																18 18
	70 78																
	70 84																18
	89																18 18
	93																18
	115																18
92																	18
93	127																18
94	129																18
95																	18
	149																18
	216																18
	227																18
	249									_							18
	243 253																18 18
	233 284																18
sd-637					.					, —	1-	<u></u>	,	_ · ·		- 1 184	
									-			_				=- V '	

HLA-DRB1*0101 15-mers

•	Pos	-1	2	2	Λ	5	6	7	0	۵.	۵	_1_	2	2	1	5	score
			7	-5	4	-	-0-	,	0		0		~		7		
102	286	- P	-V	Ъ	N	P	I	V	Y	G	V	K	T	K	E	-I	18
103	303	-R	I	_ <u>L</u>	R	Ь	F	Н	V	A	Т	Н	Λ	s	E	<u>-</u> P	18

HLA-DRB1*0301 (DR17) 15-mers

	Pos	123456789012345	score
1	16	LIGLPCLEEAQFWLA	26
2	206	ISAIGLDSLLISFSY	23
3	91	-S-T-T I Q-F D-A-C L L Q I F A	22
4	117	LLAMAFDRYVAICHP	22
5	38	LIAVLCNLTIIYIVR	21
6	179	SYCLHQDVMKLACDD	21
7	211	LDSLLISFSYLLILK	21
용	219	<u> </u>	21
9	272	P L P V I L A N I Y L L V P P	21
10	26	QFWLAFPLCSLYLIA	20
11		STVLLAMAFDRYVAI	20
12	129	CHPLRHÄTVLTLPRV	20
13	134	<u> </u>	20
14	186	V M K L A C D D I R V N V Y	20
15	200	YGLIVII SAI-GLDSL	20
16	270	- D S P L P V I L A N I Y L L V	20
17	297	TKEIRQRILRLFHVA	20
18	11	ATYFILICLPCLEEA	19
19	54	E H S L H E P M Y I F L C M L	19
20	106	I H S L S C M E S T V L L A M	19
21	165	IKQLPFCRSNILSHS	19
22	191	CDDIRVNVVGLIVI	19
23	203	IVIISAIGLDSLLIS	19
24	213	-S L L I S F S Y L L I L K T V	19
25		LKTVLGLTR-EAQAKA	19
26	227	V L C L T R E A Q A K A F C T	19
27		AVFIFYVPFIGLSMV	19
28		V P F I C L S M V H R F S K R	19
29		LANIYLLVPPVLNPI	19
30		LYLIAVLGNLTIIYI	18
31		TIQFDACLLQIFAIH	18
32		ACLLQIFAIHSLSGM	18
		YVAICHPLRHATVLT	18
		MAPLPVFIKQLPFCR	18
35		M K L A C D D I R V N V V Y G	18
		<u> </u>	18
37		KTVLCLTREAQAKAF	18
38		Y L L V P P V L N P I V Y C V	18
39		LNPIVYGVKTKEIRQ	18
40		C L P C L E E A Q F W L A F P	17
41		N L T I I Y I V R T E H S L H	17
42		KICVAAVVRCAALMA	17
		A P L P V F I K Q L P F C R S	17
		FIGLSMVHRFSKRRD	17
		L S M V H R F S K R R D S P L	17
4-0	±3/	V L T L P R V T K I G V A A V	16

HLA-DRB1*0301 (DR17) 15-mers

	Pos	1 2 3 4 5 6 7 8 9 0 1 2 3 4 5	score
47	262	VHRFSKRRDSPLPVI	16
48	29 4	GVKTKEIRQRILRLF	16
49	46	T I-I Y I V R T E H S L H E P	15
50	51	VRTEHSLHEPMYIFL	15
51	172	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	15
52	189	LACDIRVNVVYGLI	15
53	212	-D-S-L-L-I-S-F-S-Y-L-L-I-L-K-T	15
54	218	FSYLLILKTVLGLTR	15
55	271	SPLPVILANIYLLVP	15
56	279	N I Y L L V P P V L N P I V Y	15
57	12	TYFILICLP-C-L-E-A-Q	14
58	35	<u>S L Y L I A V L C N L T I I Y</u>	14
59	64	FLCMLSCIDILISTS	14
60	140	LPRVTKICVAAVVRG	14
61	273	LPVILANIYLLVPPV	14
62	301	RQRILRLFHVATHAS	14
63	13	YFILICLPCLEEAQF	13
64	47	IIYIVRTEHSLHEPM	13
65	71	I D I L I S T S S M P K M L A	. 13
66	80	MPKMLAIFWFNSTTI	13
67	109	LSCMESTVLLAMAFD	13
68	113	ESTVLLAMAFDRYVA	13
69	135	ATVLTLPRVTKIGVA	13
70	195	RVNVYCLIVISAI	13
71	202	LIVIISAICLDSLLI	13
72	220	Y L L I L K T V L G L T R E A	13
73	221	LLILKTVLGLTREAQ	13
74	264	RFSKRRDSPLPVILA	13
75	280	IYLLVPPVLNPIVYC	13
76	302	QRILRLFHV,ATHASE	13

HLA-DRB1*0401 (DR4Dw4) 15-mers

	Pos	123456789012345	score
1	36	LYLIAVLGNLTIIYI	26
2	45	LTITYVRTEHSLHE	26
3	68	LSCIDILISTSSMPK	26
4	83	<u>M L A I F W F N S T T I Q F D</u>	26
5	134	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	26
6	145	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	26
7	224	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	26
용	227	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	26
9	256	FICLSMVHRFSKRRD	26
10	281	<u>Y L L V P P V L N P I V Y C V</u>	26
11	289	NPIVYCVKTKEIRQR	26
12	301	RQRILRLFHVATHAS	26
13	11	ATYFILICLPGLEEA	22
14	24	-E-A-Q-F-W-L-A-F-P-L-C-S-L-Y-L	22
15	25	AQFWLAFPLCSLYLI	22
16	34	-C S L Y L I A V L G N L T I I	22
17	84	LAIFWFNSTTIQFDA	22
18	122	F D R Y V A I C H P L R H A T	22

HLA-DRB1*0401 (DR4Dw4) 15-mers

	Pos	1 2 3 4 5 6 7 8 9 0 1 2 3 4 5	score
19	197	NVVYGLIVIISAIGL	22
20		LISFSYLLILKTVLG	22
21		SFSYLLILKTVLGLT	22
22		FIFYVPFICLSMVHR	22
23		ANIYLLVPPVLNPIV	22
24		LPGLEEAQFWLAFPL	22
25		A F P L C S L Y L I A V L C N	20
20		LCSLYLIAVLCNLTI	20
27		S L Y L I A V L G N L T I I Y	20
2 7		IAVLGNLTIIYIVRT	20
29		L C N L T I I Y I V R T E H S	20
30		N L T I I Y I V R T E H S L H	20
31		I Y I V R T E H S L H E P M Y	
32		HEPMYIFLEMLSGID	20
33		Y I F L C M L S C I D I L I S	20
33			20
35		-L C M L S G I D I L I S T S S -I D I L I S T S S M P K M L A	20
36			20
30		M P K M L A I F W F N S T T I	20
-		PKMLAIFWFNSTTIQ	20
38		STTIQFDACLLQIFA	20
39		DACLIQIFAIHSLSG	20
40		ACLLQIFAIHSLSCM	20
41		LLQIFAIHSLSGMES	20
42		I F A I H S L S C M E S T V L	20
43		I H S L S G M E S T V L L A M	20
44		TVLLAMAFDRYVAIC	20
45		LLAMAFDRY VAICHP	20
46		YVAICHPLRHATVLT	20
47		CHPLRHATVLTLPRV	20
48		V L T L P R V T K I G V A A V	20
		LPRVTKIGVAAVV-RC	20
		A A L M A P L P V F I K Q L P	20
		-PVFIKQLPFCRSNIL	20
		I K Q L P F C R S N I L S H S	20
		SYCLHQDVMKLACDD	20
		- H Q D V M K L A C D D I R V N V Y Y	20
		DIRVNVVYGLIVIIS	20
		V N V V Y C L I V I I S A I C	20
		V Y G L I V I I S A I G L D S	20
		-Y G L I V I I S A I G L D S L	20
		LIVIISAICLDSLLI	20
		IVIISAICLDS LLIS	20
		I S A I C L D S L L I S F S Y	20
		A I C L D S L L I S F S Y L L	20
		LDSLLISFSYLLILK	20
65		-D S L L I S F S Y L L I L K T	20
66		-F S Y L L I L K T V L G L T R	20
		-GTCVSHVCAVFIFYV	20
68		-V S H V C A V F I F Y V P F I	20
69		-V C A V F I F Y V P F I C L S	20 20
		-A V F I F Y V P F I G L S M V	20
70 1727		AVELET VEET OF SMV	

HLA-DRB1*0401 (DR4Dw4) 15-mers

Pos	1 2 3	1 5 6	7.0	<u>a n 1 ′</u>	2-3-4-5	score
71 25	1 - I F Y					20
	2 -P L P					20
	7 - L A N					20
	5 P P V					20
	8 -C-L-P					18
	7 F W L			_		18
	9 S G I					18
	4 I Q F					18
	9 C L L					18
	7 - H S - L					18
	6 -V L L					18
- II II	6 V A I					18
	4 F I K					18
	6 L S H					18
	7 M K L			_		18
	7 I I S					18
	3 - E A Q					18
	3 - K A F					18
	- 					18
	3 - Y C V					18
	4 G V K					18
	0 - S A T					16
	8 -W L A					16
	9 E P M					16
	1 M Y I					16
	5 A I F					16
	1			-		16
	7 S H S					16
	6 -Л К Л					16
	9 V F I					16
	3 - Y V P					16
	3 Y F I					14
	4 F I L					14 14
	6 L I C					14
	8 1 1 A					14
	7 I I Y					14
	4 E H S					14
	0 P M Y					14
	4 F L C					14
	0 G I D					
	2 D I L					14
	9 1 s c					14
	3 EST					14
	5 -A T V					14
	3 -V T K					14
	8 V A A					14
	9 A A V					14
	4 G A A					
	8 M A P					14
	3 -S N I			_		14
	4 Q D V			-	-	14
	1 C D D					14
sd-63712					18	8
	_					

HLA-DRB1*0401 (DR4Dw4) -15-mers

	Pos	123456789012345	score
123	195	RVNVYCLIVIISAI	14
124	213	<u> </u>	14
125	220	<u> </u>	14
126	221	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	14
127	225	KTVLCLTREAQAKAF	14
128	259	LSMVHRFSKRRDSPL	14
129	270	DSPLPVILANIYLLV	14
130	273	LPVILANIXLLVPPV	14
131	274	-P-V I L A N I Y L L V P P V L	14
132	280	IYLLV PP VLNPIVYG	14
133	284	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	14
134	302	QRILRLFHVATHASE	14

HLA-DRB1*1101 15-mers

	Pos	1 2 3 4 5 6 7 8 9 0 1 2 3 4 5	score
1	145	KICVAAVVRGAALMA	28
2	122	FDRYVAICHPLRHAT	25
3	217	SFSYLLILKTVLGLT	25
4	197	NVVYCLIVIISAIGL	2 4
5	10	S A T Y F I L I C L P G L E E	23
6	255	PFICLSMVHRFSKRR	23
7	44	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	22
용	59	EPMYIFLCMLSGIDI	22
9	158	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	22
10	237	KAFCTCVSHVCAVFI	22
11	74	LISTSSMPKMLAIFW	21
12	134	<u> </u>	20
13	137	<u>V L T L P R V T K I G V A A V</u>	20
14	162	PVFIKQLPFCRSNIL	20
15	$\frac{199}{1}$	VYGLIVIIS-AIGLDS	20
16	224	LKTVLCLTREAQAKA	20
17		FIGLSMVHRFSKRRD	20
18	290	PIVYGVKTKEIRQRI	20
19	301	RQRILRLFHVATHAS	20
20	65	-L C M L S G I D I L I S T S S	19
21	100	LLQIFAIHSLSCMES	19
22	196	V N V V Y C L I V I I S A I G	19
23	218	FSYLLILKTVLGLTR	19
24		CAVFIFYVPFIGLSM	19
25	274	PVILANIYLLVPPVL	19
26	45	LTIIYIVRTEHSLHE	18
27	68	LSGIDILISTSSMPK	18
28		MPKMLAIFWFNSTTI	18
29		DACLLQIFAIHSLSC	18
30		I F A I H S L S C M E S T V L	18
31		AIGLDSLLISFSYLL	18
32		V F I F Y V P F I C L S M V H	18
33		MYIFLCMLSCIDILI	17
34		LISFSYLLILKTVLC	17
35		LSMVHRFSKRRDSPL	17
36	278	ANIYLLVPPVLNPIV	17

HLA-DRB1*1101 15-mers

	Pos	123456789012345	score
37	288	LNPIVYGVKTKEIRQ	17
38	11	ATYFILIGL PGLEEA	16
39	24	EAQFWLAFPLCSLYL	16
40	42	LCNLTIIYIVRTEHS	16
41	253	-Y V P F I C L S M V H R F S K	16
42	47	<u>-IIYIVRTEHSLHEPM</u>	15
43	99	C L L Q I F A I H S L S G M E	15
44	116	V L L A M A F D R Y V A I C H	15
45	143	VTKIGVAAVVRGAAL	15
4-6	179	- S Y C L H Q D V M K L A C D D	15
47	227	V L C L T R E A Q A K A F G T	15
48	260	<u>S M V H R F S K R R D S P L P</u>	15
49	261	MVHRFSKRRDSPLPV	15
50	277	-L A N I Y L L V P P V L N P I	15
51	285	PPVLNPIVYCVKTKE	15
52	114	STVLLAMAFDRYVAI	14
53	125	YVAICHPLRHATVLT	14
54	126	V A I C H P L R H A T V L T L	14
55	140	LPRVTKIGVAAVVRG	14
56	170	FCRSNILSHSYCLHQ	14
57	180	YCLHQDVMKLACDDI	14
58	193	DIRVNVYGLIVIIS	14
59	229	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	14
60	270	DSPLPVILANIYLLV	14
61	298	-KEIRQRILRLFHVAT	14

sd-63712

HLA-DRB1*0101 15-mers

		Pos	1 2 3 4 5 6 7 8 9 0 1 2 3 4 5	score
2472	1	200	YGLIVIISAIGLDSL	36
2473	= 2	68	LSGIDILISTSSMPK	3 <u>3</u>
2474	<u>=</u>	62	YIFLCMLSGIDILIS	<u>≖</u> = 33
2475	<u>⇒</u>	103	I F A I H S L S G M E S T V L	32
2476	5	45	LTIIYIVRTEHSLHE	3 <u>2</u> 31
2477	<u>≅</u>	45 193	DIRVNVVGLIVIIS	31
2478	<u>≚</u> 7	277	LANIYL LVPPVLNPI	<u>≅</u> 31
2479	= 8	<u>97</u>	DACLLQIFAIHSLSG	30
2480	1 2 3 4 5 6 7 8 9	106	I H S L S G M E S T V L L A M	36 34 33 32 31 31 30 30 29 28 27 26 26 26 25 25 25
2481	10	240	G T C V S H V C A V F I F Y V	30
2482	11	10	G T C V S H V C A V F I F Y V S A T Y F I L I G L P G L E	== 29
2483	12	289	NPIVYGVKTKEIRQR	<u>==</u> 29
2484	13		ATYFILIGLPGLEEA	28
2485	14	<u>11</u> 250	FIFYVPFIGLSMVHR	27
2486	<u>15</u>	140	L P R V T K I G V A A V V R G	26
2487	16	183	HODVMKLACDDIRVN	26
2488	<u>17</u>	217		26
2489	18		S F S Y L L I L K T V L G L T L I G L P G L E E A Q F W L A	25
2490	19	16 24 36 70	E A Q F W L A F P L C S L Y L	 25
2491	20	36	E A Q F W L A F P L C S L Y L L Y L I	
2492	21	70	G I D I L I S T S S M P K M L	25
2493	22	111	G M E S T V L L A M A F D R Y	25
2494	23	148		25
2495	24	162	V A A V R G A A L M A P L P P V F I K Q L P F C R S N I L	25 25 25 25
2496	25	197	N V V Y G L I V I I S A I G L	<u>25</u>
<u>2497</u>	26	211	LDSLLISFSYLLILK	<u>25</u>
<u>2498</u>	<u>27</u>	218	FSYLLILKTVLGLTR	<u>25</u>
<u> 2499</u>	<u>28</u>	<u>13</u>	YFILICLPGLEEAQF	24
<u>2500</u>	<u>29</u>	<u>30</u>	A F P L C S L Y L I A V L G N	<u>24</u>
<u>2501</u>	<u>30</u>	13 30 39 77	I A V L G N L T I I Y I V R T	24 24 24 24 24
<u>2502</u>	<u>31</u>		TSSMPKMLAIFWFNS	<u>24</u>
<u>2503</u>	<u>32</u>	85	AIFWFNSTTIQFDAC	<u>24</u>
2504	<u>33</u>	137	<u>V L T L P R V T K I G V A A V</u>	<u>24</u>
2505	<u>34</u>	<u>151</u>	V V R G A A L M A P L P V F I	24
2506	<u>35</u>	<u> 161</u>	L P V F I K Q L P F C R S N I V N V V Y G L I V I I S A I G	24
2507	<u>36</u>	<u> 196</u>	V N V V Y G L I V I I S A I G	<u>24</u>
<u>2508</u>	<u>37</u>	<u>202</u>	LIVIISAIGLDSLLI	<u>24</u>
<u>2509</u>	<u>38</u>	<u>208</u>	AIGLDSLLISFSYLL	<u>24</u>
<u>2510</u>	<u>39</u>	<u>248</u>	AVFIFYVPFIGLSMV	<u>24</u>
<u>2511</u>	<u>40</u>	<u>251</u>	I F Y V P F I G L S M V H R F	<u>24</u>
<u>2512</u>	<u>41</u>	<u>83</u> 101	M L A I F W F N S T T I Q F D	<u>23</u>
<u>2513</u>	<u>42</u>	101	L Q I F A I H S L S G M E S T	<u>23</u>
<u>2514</u>	<u>43</u>	<u>165</u>	IKQLPFCRSNILSHS	<u>23</u>
<u>2515</u>	44	<u>203</u>	IVIISAIGLDSLLIS	<u>23</u>
<u>2516</u>	<u>45</u>	221	L L I L K T V L G L T R E A Q	<u>23</u>
<u>2517</u>	46	278	ANIYLLVPPVLNPIV	<u>23</u>
<u>2518</u>	47	<u>27</u>	FWLAFPLCSLYLIAV	<u>22</u>
<u>2519</u>	48	<u>35</u>	S L Y L I A V L G N L T I I Y	24 24 24 23 23 23 23 23 23 22 22 22
<u>2520</u>	<u>49</u>	<u>61</u>	MYIFLCMLSGIDILI	<u>22</u>

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HLA-DRB1*0101 15-mers

	Pos	1 2 3 4 5 6 7 8 9 0 1 2 3 4 5	score
<u>2521</u> <u>50</u>	<u>65</u>	L C M L S G I D I L I S T S S	22
2522 51	80	M P K M L A I F W F N S T T I	22
2523 52	145	K I G V A A V V R G A A L M A	22
2524 53	$\frac{145}{146}$		22
	$\frac{140}{154}$		<u> </u>
			<u> </u>
2526 <u>55</u>	205		22
<u>2527</u> <u>56</u>	<u>243</u>	V S H V C A V F I F Y V P F I	<u>22</u>
<u>2528</u> <u>57</u>	270	DSPLPVILANIYLLV	22
<u>2529</u> <u>58</u>	274	PVILANIYLLVPPVL	22
<u>2530</u> <u>59</u>	281	Y L L V P P V L N P I V Y G V	22
<u>2531</u> <u>60</u>	34	C S L Y L I A V L G N L T I I	<u>21</u>
<u>2532</u> <u>61</u>	<u>69</u> 152	S G I D I L I S T S S M P K M	<u>21</u>
2533 62	<u>152</u>	V R G A A L M A P L P V F I K	<u>21</u>
<u>2534</u> <u>63</u>	299	EIRQRILRLFHVATH	<u>21</u>
2535 64	<u>100</u>	LLQIFAIHSLSGMES	<u>20</u>
<u>2536</u> <u>65</u>	<u>135</u>	A T V L T L P R V T K I G V A	<u>20</u>
2537 66 2538 67 2539 68	<u>141</u>	<u>P R V T K I G V A A V V R G A</u>	22 22 22 22 22 22 21 21 21 21 20 20 20 20 20 19
<u>2538</u> <u>67</u>	191	C D D I R V N V V Y G L I V I	<u>20</u>
<u>2539</u> <u>68</u>	<u>199</u>	V Y G L I V I I S A I G L D S	<u>20</u>
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2554 83	14	FILIGLPGLEEAQFW	18
2555 84	<u>25</u>	AQFWLAFPLCSLYLI	18
<u>2556</u> <u>85</u>	42	LGNLTIIYIVRTEHS	<u>18</u>
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<u>2558</u> 87	78	S S M P K M L A I F W F N S T	18
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2560 89	89	FNSTTIQFDACLLQI	18
<u>2561 90</u>	<u>93</u>	TIQFDACLLQIFAIH	18
<u>2562 91</u>	115	T V L L A M A F D R Y V A I C	18
<u>2563 92</u>	119	A M A F D R Y V A I C H P L R	18
2564 93	127	A I C H P L R H A T V L T L P	18
2565 94	129	C H P L R H A T V L T L P R V	18
2566 95	147	G V A A V V R G A A L M A P L	18
2567 96	149	A A V V R G A A L M A P L P V	18
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sd-63712

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2657

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sd-63712

7

<u> 224</u>

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194

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V L G L T R E A Q A K A F G

HLA-DRB1*0401 (DR4Dw4) 15-mers

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2661	11	289	N P I V Y G V K T K E I R Q R	26
2662	12	301	RORILRLFHVATHAS	26
2663	13	11	ATYFILIGLPGLEEA	22
2664	14	24	E A Q F W L A F P L C S L Y L	22
2665	15	25	AOFWLAFPLCSLYLI	26 26 22 22 22 22 22 22 22 22 22 22 22 2
2666	16	34	CSLYLIAVLGNLTII	22
2667	<u>17</u>	84	LAIFWFNSTTIQFDA	22
2668	18	122	F D R Y V A I C H P L R H A T	22
2669	19	197	N V V Y G L I V I I S A I G L	22
2670	20	215	LISFSYLLILKTVLG	22
2671	21	217	SFSYLLILKTVLGLT	22
<u> 2672</u>	22	250	FIFYVPFIGLSMVHR	22
<u> 2673</u>	23	278	ANIYLLVPPVLNPIV	22
<u> 2674</u>	24	<u>19</u>	L P G L E E A Q F W L A F P L	20
<u> 2675</u>	25	30	AFPLCSLYLIAVLGN	20
<u> 2676</u>	26	33	LCSLYLIAVLGNLTI	<u>20</u>
<u> 2677</u>	27	<u>35</u>	S L Y L I A V L G N L T I I Y	<u>20</u>
<u> 2678</u>	<u>28</u>	<u>39</u>	I A V L G N L T I I Y I V R T	20
<u> 2679</u>	<u>29</u>	<u>42</u>	LGNLTIIYIVRTEHS	<u>20</u>
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<u> 2681</u>	31	48	I Y I V R T E H S L H E P M Y	<u>20</u>
2682	32	<u>58</u>	H E P M Y I F L C M L S G I D	20
<u> 2683</u>	33	<u>62</u>	YIFLCMLSGIDILIS	20
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<u> 2687</u>	<u>37</u>	81	PKMLAIFWFNSTTIO	<u>20</u>
<u> 2688</u>	38	<u>91</u>	STTIQFDACLLQIFA	<u>20</u>
<u> 2689</u>	<u>39</u>	<u>97</u>	DACLLQIFAIHSLSG	<u>20</u>
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<u>2691</u>	<u>41</u>	<u>100</u>	LLQIFAIHSLSGMES	<u>20</u>
<u>2692</u>	<u>42</u>	<u>103</u>	I F A I H S L S G M E S T V L	<u>20</u>
<u>2693</u>	<u>43</u>	<u>106</u>	I H S L S G M E S T V L L A M	20
<u>2694</u>	44	115	T V L L A M A F D R Y V A I C	<u>20</u>
<u>2695</u>	<u>45</u>	<u>117</u>	L L A M A F D R Y V A I C H P	<u>20</u>
2696	46	<u>125</u>	Y V A I C H P L R H A T V L T	<u>20</u>
<u>2697</u>	47	129	<u>C </u>	<u>20</u>
<u>2698</u>	48	<u>137</u>	V L T L P R V T K I G V A A V	<u>20</u>
2699	<u>49</u>	140	L P R V T K I G V A A V V R G	<u>20</u>
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<u>2701</u>	<u>51</u>	<u>162</u>	PVFIKQLPFCRSNIL	<u>20</u>
2702	<u>52</u>	<u>165</u>	IKQLPFCRSNILSHS	<u>20</u>
2703	<u>53</u>	<u>179</u>	SYCLHQDVMKLACDD	<u>20</u>
2704	<u>54</u>	183	H Q D V M K L A C D D I R V N	<u>20</u>
2705	<u>55</u>	<u>186</u>	V M K L A C D D I R V N V V Y	<u>20</u>
<u>2706</u>	<u>56</u>	<u>193</u>	DIRVNVVGLIVIIS	<u>20</u>
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HLA-DRB1*0401 (DR4Dw4) 15-mers

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2711 61	203	<u>I V I I S A I G L D S L L I S</u> 20
2712 62	206	ISAIGLDSLLISFSY
2713 63	208	A I G L D S L L I S F S Y L L 20
2714 64	211	<u>L D S L L I S F S Y L L I L K</u> 20
2715 65	212	D S L L I S F S Y L L I L K T
2716 66	218	Y G L I V I I S A I G L D S L 20
2717 67	240	G T C V S H V C A V F I F Y V 20
2718 68	243	<u>V S H V C A V F I F Y V P F I</u>
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2720 70	248	A V F I F Y V P F I G L S M V
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2721 71 2722 72 2723 73	277	LANIYLLVPPVLNPI
2724 74	285	PPVLNPIVYGVKTKE
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2726 76	18 27 69 94 99	FWLAFPLCSLYLIAV 18
2727 77	6 9	SGIDILISTSSMPKM 18
2728 78	94	IQFDACLLQIFAIHS 18
2729 79	99	CLLQIFAIHSLSGME 18
2730 80	107	H S L S G M E S T V L L A M A 18
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2732 82	126	V A I C H P L R H A T V L T L
2733 83	164	FIKQLPFCRSNILSH 18
2734 84	176	F I K Q L P F C R S N I L S H 18 L S H S Y C L H Q D V M K L A 18
2735 85	187	M K L A C D D I R V N V V Y G
<u>2736</u> 86	205	IISAIGLDSLLISFS 18
<u>2737</u> 87	233	EAQAKAFGTCVSHVC
<u>2738</u> 88	237	KAFGTCVSHVCAVFI 18
2738 88 2739 89	271	S P L P V I L A N I Y L L V P
<u>2740 90</u>	293	YGVKTKEIRQRILRL 18
<u>2741</u> <u>91</u>	294	GVKTKEIRQRILRLF 18
<u>2742</u> <u>92</u>	10	SATYFILIGLPGLEE 16
<u>2743</u> <u>93</u>	28	W L A F P L C S L Y L I A V L 16 E P M Y I F L C M L S G I D I 16
<u>2744</u> <u>94</u>	<u>59</u>	EPMYIFLCMLSGIDI
<u>2745</u> 95	<u>61</u>	MYIFLCMLSGIDILI 16
2746 96	<u>85</u>	AIFWFNSTTIQFDAC 16
<u>2747</u> <u>97</u>	<u>101</u>	LQIFAIHSLSGMEST 16
<u>2748</u> <u>98</u>	<u>177</u>	SHSYCLHQDVMKLAC 16
<u>2749</u> <u>99</u>	<u>236</u>	AKAFGTCVSHVCAVF 16
<u>2750</u> <u>100</u>	<u>249</u>	VFIFYVPFIGLSMVH 16
<u> 2751 101</u>	<u>253</u>	Y V P F I G L S M V H R F S K 16
<u>2752</u> <u>102</u>	<u>13</u>	Y F I L I G L P G L E E A Q F
<u>2753</u> <u>103</u>	<u>14</u>	FILIGLPGLEEAQFW 14
<u>2754</u> <u>104</u>	<u>16</u>	LIGLPGLEEAQFWLA
<u>2755</u> <u>105</u>	<u>38</u>	<u>L I A V L G N L T I I Y I V R</u>
<u>2756</u> <u>106</u>	<u>47</u>	I I Y I V R T E H S L H E P M
<u>2757</u> <u>107</u>	<u>54</u>	S H S Y C L H Q D V M K L A C 16 A K A F G T C V S H V C A V F 16 V F I F Y V P F I G L S M V H 16 Y V P F I G L S M V H R F S K 16 Y F I L I G L P G L E E A Q F 14 F I G L P G L E E A Q F W 14 L I G L P G L E E A Q F W 14 L I G L P G L E E A Q F W 14 L I G L P G L E E A Q F W 14 L I G L P G L E E A Q F W L A 14 L I G L P G I E E A Q F W L A 14 L I G L P G I E E A Q F W L A 14 L I G L P G I E E A Q F W L A 14 L I G L P G I E E A Q F W L A 14 L I G I E E A Q F W L A 14 L I E E A C E E A Q F W L A 14 L I E E E A C E E E E E E E E E E E E E E E
<u>2758</u> <u>108</u>	<u>60</u>	PMYIFLCMLSGIDIL 14
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<u>SEO</u> ID			
	LA-DI	RB1*0401 (DR4Dw4) 15-mers	
<u>I</u>	?os	1 2 3 4 5 6 7 8 9 0 1 2 3 4 5	score
<u>2759</u> 109	<u>64</u>	F L C M L S G I D I L I S T S	<u>14</u>
<u>2760</u> <u>110</u>	<u>70</u>	G I D I L I S T S S M P K M L	<u>14</u>
<u>2761</u> <u>111</u>	<u>72</u>	DILISTSSMPKMLAI	<u>14</u>
<u>2762</u> <u>112</u>	109	LSGMESTVLLAMAFD	<u>14</u>
<u>2763</u> <u>113</u>	113	ESTVLLAMAFDRYVA	$\frac{14}{3}$
2764 <u>114</u> 2765 115	135	ATVLTLPRVTKIGVA	$\frac{14}{14}$
2765 <u>115</u> 2766 116	143 148	V T K I G V A A V V R G A A L V A A V V R G A A L M A P L P	$\frac{14}{14}$
$\frac{2760}{2767} \frac{110}{117}$	149	A A V V R G A A L M A P L P V	$\frac{14}{14}$
$\frac{2768}{2768}$ 118	154	GAALMAPLPVFIKQL	$\frac{14}{14}$
2769 119	158	MAPLPVFIKQLPFCR	14
2770 120	173	S N I L S H S Y C L H Q D V M	14
<u>2771</u> <u>121</u>	184	Q D V M K L A C D D I R V N V	14
<u>2772</u> <u>122</u>	<u> 191</u>	C D D I R V N V V Y G L I V I	<u>14</u>
<u>2773</u> <u>123</u>	<u>195</u>	RVNVYGLIVIISAI	14
2774 124 2775 125	213	S L L I S F S Y L L I L K T V	$\frac{14}{14}$
$\frac{2775}{2776} \frac{125}{126}$	220 221	<u>Y L L I L K T V L G L T R E A</u> <u>L L I L K T V L G L T R E A Q</u>	$\frac{14}{14}$
$\frac{2770}{2777} \frac{120}{127}$	225	K T V L G L T R E A Q A K A F	$\frac{14}{14}$
$\frac{2778}{2778} \frac{128}{128}$	259	L S M V H R F S K R R D S P L	$\frac{14}{14}$
2779 129	270	DSPLPVILANIYLLV	14
<u>2780</u> <u>130</u>	273	L P V I L A N I Y L L V P P V	$\overline{\overline{14}}$
<u>2781</u> <u>131</u>	274	P V I L A N I Y L L V P P V L	14
<u>2782</u> <u>132</u>	<u>280</u>	I Y L L V P P V L N P I V Y G	<u>14</u>
<u>2783</u> <u>133</u>	284	V P P V L N P I V Y G V K T K	<u>14</u>
<u>2784</u> <u>134</u>	<u>302</u>	Q R I L R L F H V A T H A S E	<u>14</u>
SEQ			
<u>ID</u>			
		B1*1101 15-mers	
	<u>08</u>	1 2 3 4 5 6 7 8 9 0 1 2 3 4 5 F T C W D D W W D C D D W D	score
	. <u>45</u> .22	K I G V A A V V R G A A L M A F D R Y V A I C H P L R H A T	<u>28</u>
$\frac{2787}{2787}$ $\frac{2}{3}$ $\frac{1}{2}$	217	SFSYLLILKTVLGLT	<u>43</u> 25
2788 4 1	97	N V V Y G L I V I I S A I G L	2 <u>3</u>
278 <u>9</u> 5		S A T Y F I L I G L P G L E E	23
<u>2790 6 2</u>	10 255	PFIGLSMVHRFSKRR	<u>23</u>
<u>2791</u> <u>7</u>	44 =	N L T I I Y I V R T E H S L H	<u>22</u>
<u>2792</u> <u>8</u>	<u>59</u> =	E P M Y I F L C M L S G I D I	<u>22</u>
<u>2793</u> <u>9</u> <u>1</u>	.58 =	MAPLPVFIKQLPFCR	<u>22</u>
		K A F G T C V S H V C A V F I L I S T S M P K M L A I F W	<u>22</u>
		<u>L I S T S S M P K M L A I F W</u> <u>H A T V L T L P R V T K I G V</u>	<u>41</u> 20
2797 13 1		V L T L P R V T K I G V A A V	<u>20</u> 20
	.62	PVFIKQLPFCRSNIL	20
<u>2799 15 1</u>	99	VYGLIVIISAIGLDS	20
	24	LKTVLGLTREAQAKA	<u>20</u>
	<u> </u>	FIGLSMVHRFSKRRD	25 24 23 23 22 22 22 21 20 20 20 20 20 20 20
<u>2802</u> <u>18</u> <u>2</u>	<u>90 </u>	PIVYGVKTKEIRQRI	<u>20</u>

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<u>HLA-DRB1*1101 15-mers</u>

<u>P</u>	<u>os</u> _1	2	3	4	5	6	7	8	9	0	1	2	3	4	<u>5</u>	score
<u>2803</u> 19 3	301 R	. Q	R	I	L	R	L	F	Н	V	A	Т	Н	A		<u>20</u>
2804 20	65 <u>I</u>	С	М	L	S	G	Ι	D	I	L	Ι	S	Т	S	S	19
	100 I	L	Q	I	F	Α	I	Н	s	L	S	G	М	Ε	S	<u>19</u>
<u>2806</u> <u>22</u> <u>1</u>	196 V	N	V	v	Y	G	L	I	v	Ι	Ι	S	Α	Ι	G	19
2807 23 2	218 F	S	Y	L	L	I	L	K	T	V	L	G	L	Т	R	<u>19</u>
2808 24 2	247 <u>C</u>	Α	V	F	I	F	Y	V	P	F	Ι	G	L	S	M	<u>19</u>
2808 24 2 2809 25 2	274 P	V	I	L	A	N	I	Y	L	L	V	P	Р	V	L	19 18
<u>2810</u> <u>26</u>	45 68 1		Ι	I	Y	I	V	R	T	E	Н	S	L	Н	E	18
<u>2811</u> <u>27</u>	68 I		G	I	D	I	L	Ι	S	T	S	S	М	Ρ	K	18
<u>2812</u> <u>28</u>	80 <u>M</u>		K	M	L	A	Ι	F	W	F	N	S	T	T	Ι	18
<u>2813</u> <u>29</u>	97 <u>D</u>	A	С	L	L	Q	I	F	Α	Ι	Н	S	L	S	G	18
<u>2814 30 1</u>	<u> 103</u>	F	A	I	Н	s	L	S	G	М	E	S	T	V	L	18
<u>2815</u> <u>31</u> <u>2</u>	208 A		G	L	D	s	L	L	I	S	F	<u>S</u>	Y	L	L	18
<u>2816</u> <u>32</u> <u>2</u>	249 v		I	F	Y	V	P	F	I	G	L	S	M	V	<u>H</u>	<u>18</u>
<u>2817</u> <u>33</u>	61 <u>M</u>	Y	I	F	L	С	М	L	S	G	I	D	I	Ļ	I	<u>17</u>
2818 34 2	215 1		S	F	S	Y	L	Ļ	I	L	K	Т	V	L	G	<u>17</u>
<u>2819 35 2</u>	259 I		М	V	Н	R	F	S	K	R	R	D	S	P	L	<u>17</u>
	278 A		I	Y	L	L	V	P	P	V	L	N	P	I	V	<u>17</u>
2821 37 2	288 <u>I</u>		P	Ι	V	Y	G	V	K	T	K	E	I	R	0	<u>17</u>
2822 38	11 A 24 E 42 I		Y	F	Ι	L	I	G	L	P	G	L	Ε	E	<u>A</u>	<u>16</u>
2823 39	<u>24</u> <u>E</u>		Q	F	W	L	A	F	P	L	C	S	Ļ	Y	L	<u>16</u>
2824 40	<u>42</u> <u>I</u>		<u>N</u>	L	T	I	I	Y	I	V	R	T	Ε	Н	<u>S</u>	<u>16</u>
2825 41 2	253 Y		P	F	I	G	L	s	M	V	Н	R	F	S	K	<u>16</u>
2826 42	47 <u>I</u>		Y	I	<u>V</u>	R	T	E	H	S	L	Н	E	P	M	<u>15</u>
<u>2827</u> <u>43</u>	47 I 99 C		L	0	I	F	A	I	H	S	L	S	G	M	<u>E</u>	<u>15</u>
2828 44 1	116 <u>V</u>		L	<u>A</u>	M	A	F	D	R	Y	V	A	I	C	<u>H</u>	<u>15</u>
<u>2829</u> <u>45</u> <u>1</u>	143 V		K	I	G	V	A	A	V	V	R	G	A	A	Ţ	<u>15</u>
	179 S		<u>C</u>	L	H	0	D	V	M	K	L	A	<u>C</u>	D	<u>D</u>	<u>15</u>
	227 v 260 s		G	<u>T.</u>	Ţ	R	<u>E</u>	A	0	A	K	A	F	G	T	16 16 15 15 15 15 15 15 15 15
		<u>M</u>	V H	H R	R	F S	S	K	R	R	D	S	P	F	<u>P</u>	75
	261 <u>M</u> 277 L		N N	I	F Y	L	K L	R V	R P	D P	S V	P L	L N	P P	<u>V</u>	15
	285 P	P	V	L	N	P P	I	V	Y	G	V	K	T	K	I E	12
	14 S	T	V	L	L	A	M	A	F	D	R	Y	v	A	I	13 14
2837 53 1	25 Y	V	A	I	C	Н	P	L	R	H	A	T	V	L	T	14 14
2837 53 1 2838 54 1	26 V		T	C	Н	P	L	R	Н	A	T	V	L	T	<u> </u>	
2839 55 1	40 L	P	R	v	Т	K	Ī	G	v	A	Ā	V	V	R	G	14
	70 F	C	R	s	N	I	L	S	Н	S	Y	Ċ	L	Н	<u>Q</u>	14
	80 Y		L	Н	Q	D	V	M	K	L	Ā	C	D	D	<u>I</u>	14
	93 D	I	R	V	N	٧	٧	Y	G	L	I	٧	Ī	I	S	=== 14
	229 G		Т	R	Ε	Α	Q	Α	K	Α	F	G	Т	С	V	==
	270 D	S	Р	L	Р	v	Ι	L	Α	N	Ι	Y	L	Ļ	V	$\frac{\overline{14}}{14}$
	98 K		Ι	R	Q	R	Ι	L	R	L	F	Н	V	A	T	14 14 14 14 14 14 14

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Table XXIX. Nucleotide sequence in the 5' region close to 101P3A11 gene. $\underline{\text{(SEQ ID NO: 2846)}}$

	NO: 2846	<u>_</u>					
	1	TGCGCTCCAC	CAAGCCTGGC	TAACTTTTGC	ATTTTTAATA	GAGGCAGGGT	TTCACCATGT
	61	TGGCCTGGCT	GGTCTCGAAC	CCCTGACCTT	GCGATCTGCC	CACCTCGGCC	TCCCAAAGTG
5	121	CTGGGATTAC	AGGCGTGAGC	CACTGTACCT	GGCGGGGCTT	ATTGTTTTTT	AAAAAGATTT
	181	CCAAAACCTT	GCCCTGGCAA	TTCTGATTTT	CTGGGCCTGG	AGCAGGACCT	GGAGGGATGG
	241	TGTTGTCAAT	TACTTTAGAT	GTTTCTATCA	GGAAAGTTTG	AGAAATGGTA	TTCAGGCCTA
	301	AACACAAACC	TCTCTTGAAA	TCTCATCCCA	GACTGAGCCC	CTGCTCCCTA	TCTTAAATTA
	361	GATTATAGTA	GGTCTTAAAG	TCAGCTGTAG	ACTGAGCCTC	TAAATCTGAA	CCCAGACCCA
10	421	CCCTAACCCC	AGGATACATC	AGAAGAGCTG	GTCAATGTGG	ACCATTCTGA	GCAATCCTGC
			TGATGGGAAA				
			CTGTGTTACA				
			ATCTGCTTGT				
			GTTGTATATA				
15			TAGTCCTGTT				
			CTGAGCCACA				
			TTGTATGCAG				
			CCACGCAGCA				
			GCTGGCTTTC				
20			CCTCATCATT				
-0			TTCCACTATT				
			GATGGGCATC				
			TCTGTCAGCC				
			TTGCCACCCA				
25			ATCTGCCCTG				
25			GTCCTACTGC				
			GCTGTCCTGT				•
			CATGGGTGTG				
			GGAGCTGTCC				
30			TGCTGTTCTG				
50			TCCCACCTCC				
			CAACCCCCTT				
			CTCACAAGGT				
			GGAATATTAG				
35	1921		TTCTTCCAGC			-	
55			TTATGACCCT				
			TTGCCTTCTC				
			AAGATCACAC				
			TGTGCCCATG				
40			TTACCCAGCC				
			CAACAAGACA				
			TGGCAGATTT				
			TAAGGGTGCC				
			TGCTTTTCAG				
45			TGTCCTGGTG				
			GATTGCGCAT				
			GAATGCTGGT				
			CTGCCTCTAC				
			GTGTGGAATG				
50			TGTGCATAAT				
50			TCGGTGAAAT				
			CATATGTCTA				
			CACGAATGGG				
55			TCTTGGAATG				
55			AAGTTGAGTG			· ·	
			AGCTCAGTCT				
			CAGACCACAC				
	3301	ACCICITECT	ACGTTTCCTG	CCICIGCTAT	CCGAGGCACT	GGCCTCCCTA	AACCCTGCCC

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	3361	TCCTGCCTCA	ATAGCAAGTC	ATGGTATCCT	CACCTCTCCC	TTCCCTTTTT	GGCTTATCT G
	3421	CCAAACATGT	ATAAAAGTCC	TTGGTTCCCC	ATCTCTACTA	AAAATACAAC	AATTAGCCGG
	3481	GTGTGATGGC	GCGTGCCTGT	AGTCCCAGCT	AGTTGGGAGG	CTGAGGCAGG	AGAAACGCTT
	3541	GAGCCCGCAA	GGTGGAGGTT	GCAGTGAGCC	GAGATCATGC	CACTGCACTC	CAGCCTGGTG
5	3601	ACAGAGCAAG	ACTCTGTGTC	AAAAAAAAA	AAAAAAAAA	AGCCTTGGTT	GTAGGGAGTT
	3661	TCTCCTAATC	CCTCTGGGAA	AGCAAGGGTG	GAGGGGAAGC	CAGTCAATCT	CCCTTCTGTT
	3721	GCCGCATGGA	AACTCCCTTA	AGGCAGGAAG	CTGAAAAAAC	TGTAGCATTC	ACCTCATTAT
	3781	TCACCTTGTC	TCATGTCTCA	CTGTCCTTCC	ACATGTCTCA	TTGTTACTCC	ATATTGGATG
	3841	GAAGTAGAAG	TCCCTTTGGT	${\tt ATTTTTTAAA}$	GTCTTTGCCA	TGTCTAAGTT	AATGAGGTTA
10	3901	ATGGAGGCAG	CAGAGATGGC	TCCAGGGTTC	TGATAGCAAG	TGTCAGGCTG	CGTGCTCTGT
	3961	AGGCACCAGA	AACTGTTGTC	ACCAGTAATT	TTGATGTGGT	CTGAGTTAGA	ATGGTCTGAT
	4021	TTGCCATGAT	CTATTTAACA	TAGCTTGATT	TAGCGTGTCC	TGTGTTCTGA	ATTTAAAACT
	4081	CACAGTTGTG	AAACTGATCA	GTAAAAAATA	AGGGGAGACC	AACTAAAAAC	CATGTTGTTC
	4141	TA TTTATAGA	TGTAGTTTTT	ACTTATTTCA	AAATACGAGG	TATTTAGTTT	TACATTCAAA
15	4201	TTGTTCTCTA	ACTCTCTAAA	ATGTTCTCTG	ACTATTTTTG	CCCTTAAGGG	AGAAACCAGA
	4261	TGTCATTGGT	CTTACGTGGC	TGGTGTTGGG	GGTGGGGAGG	GTTAAAGAAA	CCACGTTCTC
	4321	TGTCCTCAGC	CAGAAGTTCA	GTAATCCAAG	GCCAGAGAGT	GGACGGCAGA	GGCACTGTCC
	4381	CTGGGGACCT	TGGTTATAAG	TTATCCAGAC	ACAGGGACCA	GAGCCTGGGA	GACAAAAAAA
	4441	GATGTAGCCC	TAGGGCTTTG	GGAAAAGGAG	GATGGACCCA	GTGAATTCCA	CGCTTAGCAA
20	4501	GGACCTAAAC	AGTGTCCCCC	AAATGAGAGA	AGGGAGGACA	GAAAGAACAC	TTCAGGATGG
	4561	AAATGGGCTG	ACACTTAACC	GTGGAGTGTC	TCTGCAAACT	TCCTTTGCCA	TTCTCCTGTT
	4621				ATAAAGACCG	TCACGAAGAC	TACACTAATG
	4681	AGTTTCTTCT	AGCTTTTTTC	TACTCACTTT	CCCTATCTAT	CCTTCACATT	GGGAGTTGGC
	4741				ACTCTAGAGA		CATTGCCTCT
25	4801		CCCCCAGGCA	TATCCTCCCA	GGGCACGGAA	GCTGAGAAGC	AGTCCAGAAC
	4861	CACAGTGGGC	TAGTGAGGGG	TACCTGCTGA	TGTACCCTTT	GGACAGCATT	CTGCCCCACC
	4921				TG AGGCAGAG		
	4981	GAGGTCCAAG	GGAGTAGGCG	GAG Acagaga	ggctgtattt	cagtgcagcc	tgccagacct

³⁰ Note: The three high score predictions of promoters were bold and underlined. The lower case sequence indicates the beginning part of the transcript of 101P3A11 gene.

Table XXX: Promoters and their positions predicted by Neural Network Promoter 35 Prediction computer program.

			SEQ	
	Start	End	——Score ——— <u>I</u>	D NO Promoter Sequence
	25	75	-0.91 - <u>2847</u>	TTTTGCATTTTTAATAGAGGCAGGGTTTCACCATGTTGGCCTGGCTGG
40	665	715	0.95 - 2848	CAGGAAGTTGTATATAAGGAGAATCAGAGCAGAGAGAGACTAGGGTTCAG
	2477	2527	0.91 - 2849	TCAGTGTGCGTATATGTGAGAGAGAGGGTGCACACATGGAATACGTACTG
	3139	3189	0.82 - <u>2850</u>	TGACATGACTCTAAGATGCCCAGTTTCTCGGCCTGGGGTCAGCCTGGGTG
	3420	3470	<u>—</u> 0.96 – <u>2851</u>	GCCAAACATGTATAAAAGTCCTTGGTTCCCCATCTCTACTAAAAATACAA
	4092	4142	0.99 - 2852	AACTGATCAGTAAAAAATAAGGGGAGACCAACTAAAAACCATGTTGTTCT
45	4953	5003	$0.97 - \overline{2853}$	AGGCAGAGAATAAATAACCCTGACCAGGGAGGTCCAAGGGAGTAGGCGGA

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Table XXXI: Alignment of five homologous 5' upstream genomic regulatory regions of the human 101P3A11 and PSA genes.

Query: 5' upstream regulatory region of the PSA gene

5 Subject: -Putative 5' upstream regulatory region of the 101P3A11 gene. Nucleic acid sequences predicted to be binding sites for the indicated transcription factors are bolded, underlined, or italicized. 1. QUERY (SEQ ID NO: 2854) AND SUBJECT (SEQ ID NO: 2855) 10 NF-1 SP-1 NF-1 Query: 3864 ccaggctggagtgcagtggcgcagtctcggctcactgcaacctctgcctcccaggttcaa 3923 Sbjct: 3598 ccaggctggagtgcagtggcatgatctcggctcactgcaacctccaccttgcgggctcaa 3539 15 Query: 3924 gtgattctcctgcctcagcctcctgagttgctgggattacaggcatqcaqcaccatgccc 3983 Sbjct: 3538 gcgtttctcctgcctcagcctccaactagctgggactacaggcacgcgccatcacaccc 3479 Query: 3984 agctaatttttgtatttttagtagagatgggg 4015 20 Sbjct: 3478 ggctaattgttgtatttttagtagagatgggg 3447 2. QUERY (SEQ ID NO: 2856) AND SUBJECT (SEQ ID NO: 2857) 25 Query: 4670 cctgtaatcccaqctactgaggaggctgaggcaggagaatcacttgaacccagaaggcag 4729 Sbjct: 3496 cctgtagtcccagctagttgggaggctgaggcaggagaaacgcttgagcccgcaaggtgg 3555 SP1 30 NF-1 NF-1 Query: 4730 aggttgcaatgagccgagattgcgccactgcactccagcctgggtgacagagtgagactc 4789 Sbjct: 3556 aggttgcagtgagccgagatcatgccactgcactccagcct-gqtgacagagcaagactc 3614 35 Query: 4790 tgtctcaaaaaaaaaaa 4807 111 1111111111111 Sbjct: 3615 tgtgtcaaaaaaaaaa 3632 3. QUERY (SEQ ID NO: 2858) AND SUBJECT (SEQ ID NO: 2859) 40 **NF-1** SP1 Query: 142 tgagactgagtctcgctctgtgcccaggctggagtgcagtggtgcaaccttggctcactg 201 Sbjct: 3621 tgacacagagtcttgctctactgtcaccaggctggagtggcatgatctcggctcactg 3562 45 Query: 202 caagetccgcctcctgggttcacgccattctcctqcctcaqcctcctqaqtaqctqqqac 261 Sbjct: 3561 caacctccaccttgcgggctcaagcgtttctcctgcctcagcctcccaactagctgggac 3502 50 NF-1 Query: 262 tacaggcacccgccaccacgcctggctaannnnnnngtatttttagtagagatgggg 318 Sbjct: 3501 tacaggcacgcgccatcacacccggctaa--ttgttgtatttttagtagagatgggg 3447 55

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LF-A1 CP2 Query: 360 gtgatctgccaccttggcctcccaaagtgctgggattacaggcgtgagccactgcg		
SP1 LF-A1 CP2		159
NF-A1 CP2	acctt 9	3 0
Query: 360 gtgatctgcccaccttggcctcccaaagtgctgggattacaggcgtgagccactggcg	NF-1	
NF-1 Query: 420 ggc 422		119
Query: 420 ggc 422		.50
5. QUERY (SEQ ID NO: 2862) AND SUBJECT (SEQ ID NO: 2863) NF-1 Query: 4506 gccaggcacagtggctcacgcctgtaatcccaacaccatgggaggctgagatgggt		
5. QUERY (SEQ ID NO: 2862) AND SUBJECT (SEQ ID NO: 2863) NF-1 Query: 4506 gccaggcacagtggctcacgcctgtaatcccaacaccatgggaggctgagatgggt		
NF-1 CP2 Query: 4506 gccaggcacagtggctcacgcctgtaatcccaacaccatgggaggctgagatgggt Sbjct: 153 gccaggtacagtggctcacgcctgtaatcccaacaccatgggaggccgaggtgggc Query: 4566 cacgaggtcaggagtttgagaccagcctgaccaacatggtgaaactctgtctcta		
NF-1 CP2 Query: 4506 gccaggcacagtggctcacgcctgtaatcccaacaccatgggaggctgagatgggt Sbjct: 153 gccaggtacagtggctcacgcctgtaatcccaacaccatgggaggccgaggtgggc Query: 4566 cacgaggtcaggagtttgagaccagcctgaccaacatggtgaaactctgtctcta		
Query: 4506 gccaggcacagtggctcacgcctgtaatcccaacaccatgggaggctgagatgggt		
Query: 4506 gccaggcacagtggctcacgcctgtaatcccaacaccatgggaggctgagatgggt		
Sbjet: 153 gccaggtacagtggctcacgcctgtaatcccagcactttgggaggccgaggtgggc Query: 4566 cacgaggtcaggagtttgagaccagcctgaccaacatggtgaaactctgtctcta	.~+~~~+	1565
Sbjct: 153 gccaggtacagtggctcacgcctgtaatcccagcactttgggaggccgaggtgggc 25 Query: 4566 cacgaggtcaggagtttgagaccagcctgaccaacatggtgaaactctgtctcta		4500
Query: 4566 cacgaggtcaggagtttgagaccagcctgaccaacatggtgaaactctgtctcta		94
Query: 4566 cacgaggtcaggagtttgagaccagcctgaccaacatggtgaaactctgtctcta		
30		

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